

# *Water Quality Standards Academy*

## **Aquatic Life Criteria**

*March 24, 2021*

EPA Region 9 Tribal WQS Academy

# Disclaimer

- This Presentation does not:
  - Impose any binding requirements
  - Determine the obligation of the regulated community
  - Change or substitute for any statutory provision or regulation requirement
  - Represent, change or substitute for any Agency policy or guidance
  - Control in any case of conflict between this discussion and statute, regulation, policy, or guidance

The views expressed in this presentation are those of the author[s] and do not necessarily represent the views or policies of the U.S. Environmental Protection Agency.

# What are Aquatic Life Criteria?

## **What is an aquatic life criterion?**

The highest instream concentration of a toxicant to which organisms can be exposed for a period of time without causing an unacceptable adverse effect.

## **What is it intended to protect?**

Aquatic animals (e.g., fish, invertebrates, crustaceans) and plants from acute and chronic exposure to a toxicant or condition. More to the point, it is protection of the ecosystem so that it retains its “fishable, swimmable” designated use(s).

# National Recommended Water Quality Criteria

## Human Health Criteria (Fish consumption)

## Aquatic Life Criteria

Acute (Criterion Maximum Concentration)

Chronic (Criterion Continuous Concentration)

Freshwater and Saltwater

### National Recommended Water Quality Criteria for Arsenic

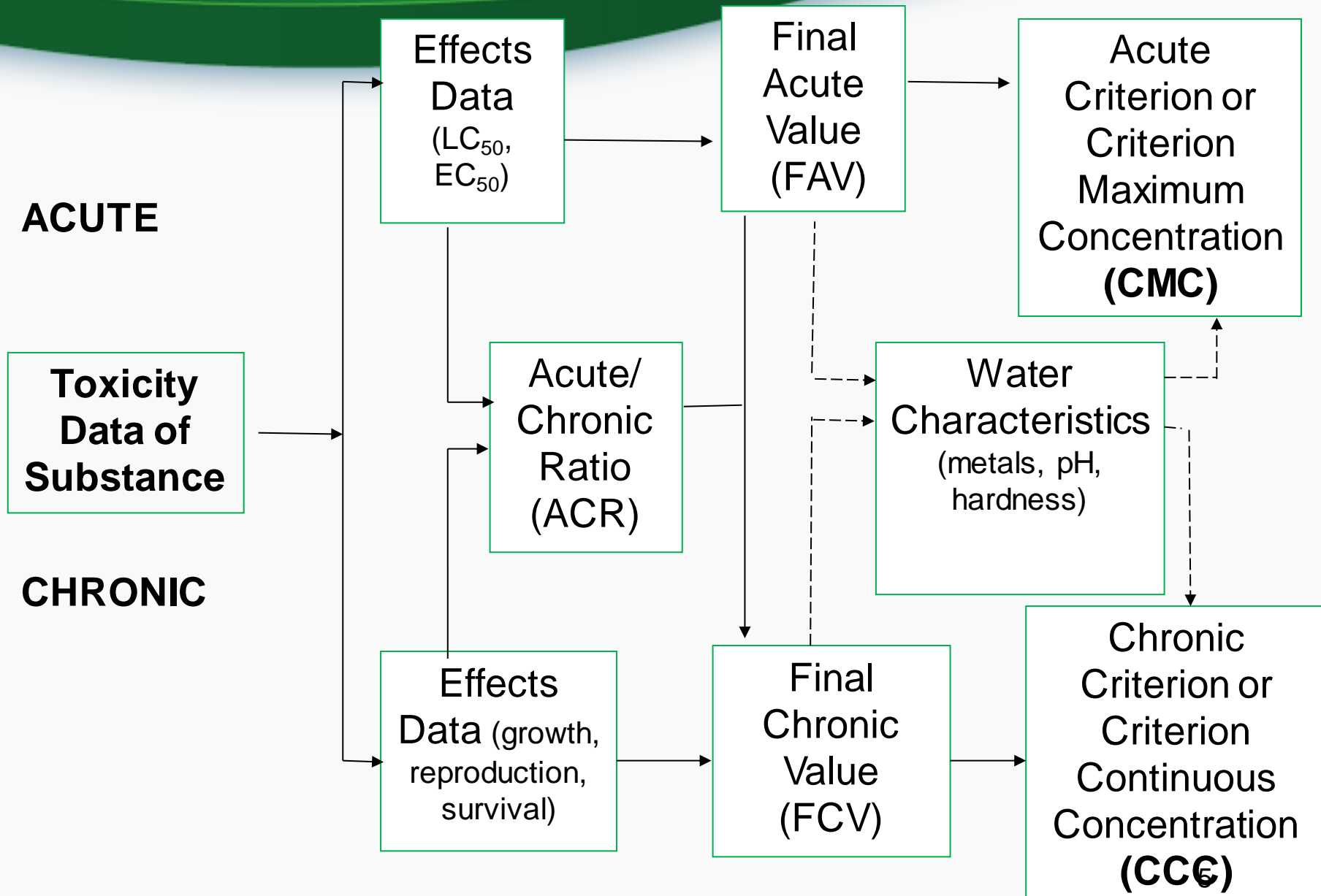
Human Health  
for the  
consumption of

Freshwater

Saltwater

Priority Pollutant	CAS Number	CMC	CCC	CMC	CCC	Water + Organism	Organism Only
Arsenic	7440382	340	150	69	36	0.018	0.14

# Criteria Derivation Overview



# Minimum Data Requirements (MDRs) Freshwater

SALMONID



SECOND  
FISH  
FAMILY



CHORDATA



PLANKTONIC  
CRUSTACEAN



BENTHIC  
CRUSTACEAN



INSECT



ROTIFERA,  
ANNELIDA,  
MOLLUSCA



OTHER  
INSECT OR  
MOLLUSC



# Minimum Data Requirements (MDRs)(Saltwater)

Images:<http://ian.umces.edu/imagelibrary/>

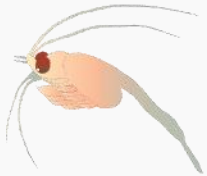
Two families in the phylum Chordata:



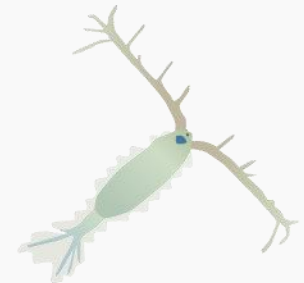
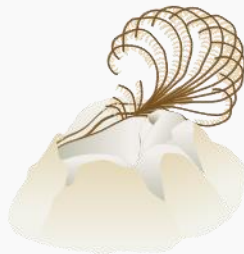
A family in a phylum other than Arthropoda or Chordata:



Either the Mysidae or Penaeidae family:



Three other families not in the phylum  
Chordata



Any other saltwater aquatic family.

# Acute Toxicity Testing

Acute toxicity - 96 hour (4 day) toxicity test for fish or other invertebrates (or 48 for *Daphnia*)

- Measured as LC<sub>50</sub> or EC<sub>50</sub> (immobilization)
- Lethal concentration [effects concentration] in which 50% of tested organisms display lethality/effect



# Acute Test Data Handling

96-hour  $LC_{50}$  values from the literature

Concentration:

0.0  $\mu\text{g/L}$

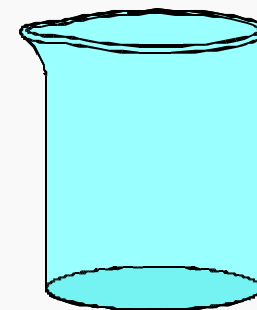
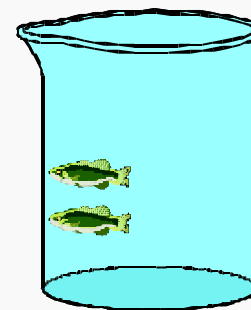
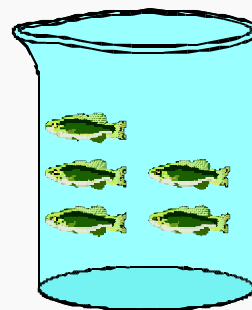
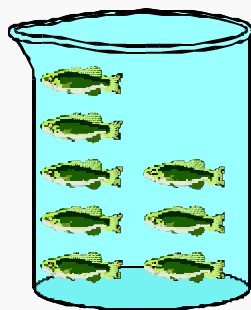
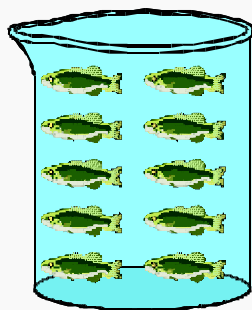
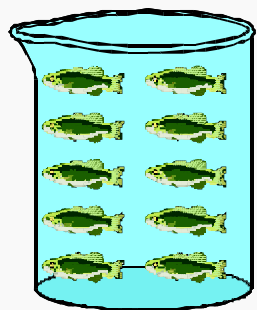
13  $\mu\text{g/L}$

25  $\mu\text{g/L}$

50  $\mu\text{g/L}$

100  $\mu\text{g/L}$

200  $\mu\text{g/L}$



**Control**  
(10)

**1**  
(10)

**2**  
(8)

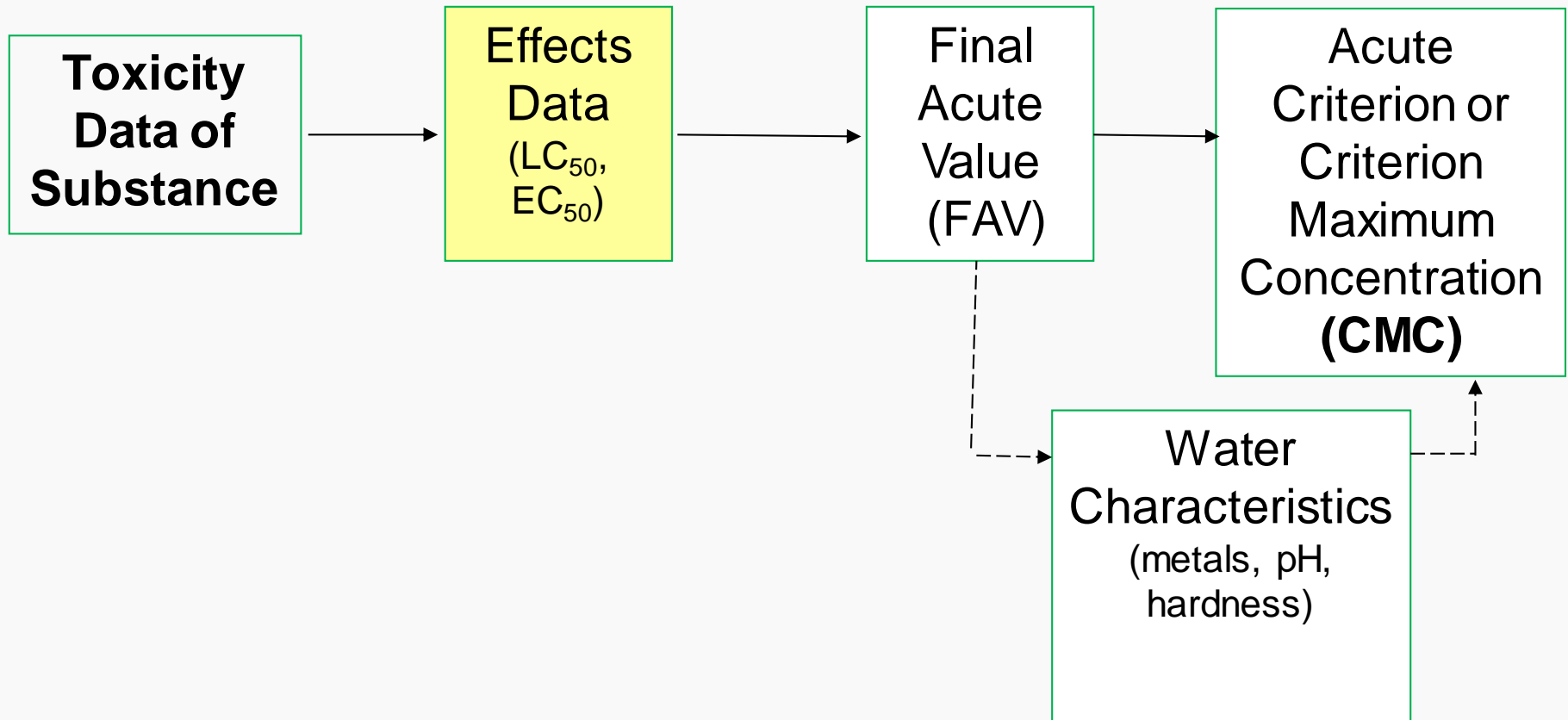
**3**  
(5)

**4**  
(2)

**5**  
(0)

**96-hr  $LC_{50} = 50 \mu\text{g/L}$**

# Acute Method - Step 1: Toxicity Data of Substance



# Data Sources and Endpoints

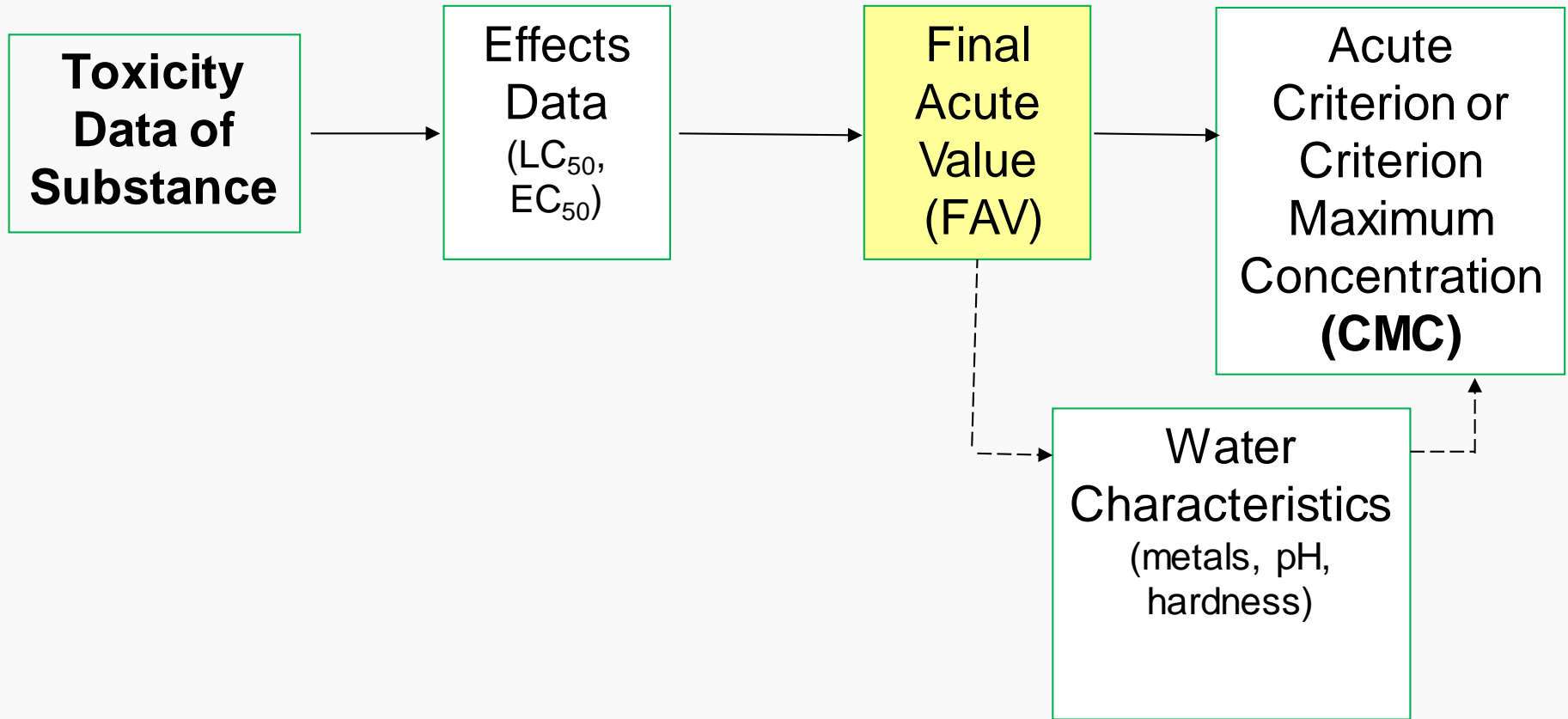
Data is collected from ecological toxicity database (ECOTOX, maintained by EPA). This is periodically updated, on a compound specific basis, from open peer-reviewed literature.

ECOTOX is a comprehensive, publicly available knowledgebase providing single chemical environmental toxicity data on aquatic life, terrestrial plants and wildlife (<https://cfpub.epa.gov/ecotox/>).

The database contains single chemical exposures using ecologically relevant species.

Main data limitations include reporting effects, concentration or dose, and exposure duration.

# Acute Method (cont'd)



# FAV Calculation Overview

- Step 1. Calculate Species Mean Acute Values (SMAVs)**
  - **geometric mean of all acceptable acute values for species**
  
- Step 2. Calculate Genus Mean Acute Values
  - geometric mean of all SMAVs for genus
  
- Step 3. Rank Genus Mean Acute Values
  - from most sensitive (#1 = lowest concentration to see an effect) to least sensitive (n)
  
- Step 4. Calculate Final Acute Value Using 4 Lowest GMAVs (or those GMAVs closest to the 5<sup>th</sup> percentile)

# FAV Calculation - SMAV

## Step 1 – Species Mean Acute Value

<i>Daphnia magna</i> EC <sub>50</sub>	25 µg/L
<i>Daphnia magna</i> EC <sub>50</sub>	30 µg/L
<i>Daphnia magna</i> EC <sub>50</sub>	35 µg/L
<i>Daphnia magna</i> EC <sub>50</sub>	28 µg/L

$$\text{SMAV} = 29 \mu\text{g/L}$$

# FAV Calculation Overview

- Step 1. Calculate Species Mean Acute Values (SMAVs)
  - geometric mean of all acceptable acute values for species
  
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  - geometric mean of all SMAVs for genus**
  
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  - from most sensitive (#1 = lowest concentration to see an effect) to least sensitive (n)
  
- Step 4. Calculate Final Acute Value Using 4 Lowest GMAVs (or those GMAVs closest to the 5<sup>th</sup> percentile)

# FAV Calculation - GMAV

## Step 2 – Genus Mean Acute Value

<i>Daphnia magna</i>	SMAV	29 µg/L
<i>Daphnia pulex</i>	SMAV	38 µg/L
<i>Daphnia ambigua</i>	SMAV	42 µg/L

$$\text{GMAV} = 36 \mu\text{g/L}$$



# Summary of steps 1 and 2

Step 1 – Calculate each SMAV (*Daphnia magna*)

<i>Daphnia magna</i> EC <sub>50</sub>	25 µg/L
<i>Daphnia magna</i> EC <sub>50</sub>	30 µg/L
<i>Daphnia magna</i> EC <sub>50</sub>	35 µg/L
<i>Daphnia magna</i> EC <sub>50</sub>	28 µg/L

$$\text{SMAV} = 29 \mu\text{g/L}$$

Step 2 – Calculate the GMAV (*Daphnia*)

<i>Daphnia magna</i>	SMAV	29 µg/L
<i>Daphnia pulex</i>	SMAV	38 µg/L
<i>Daphnia ambigua</i>	SMAV	42 µg/L

$$\text{GMAV} = 36 \mu\text{g/L}$$

# FAV Calculation Overview

- Step 1. Calculate Species Mean Acute Values (SMAVs)
  - geometric mean of all acceptable acute values for species
  
- Step 2. Calculate Genus Mean Acute Values
  - geometric mean of all SMAVs for genus
  
- Step 3. Rank Genus Mean Acute Values**
  - from most sensitive (#1 = lowest concentration to see an effect) to least sensitive (n)**
  
- Step 4. Calculate Final Acute Value Using 4 Lowest GMAVs (or those GMAVs closest to the 5<sup>th</sup> percentile)

# FAV Calculation - Ranking

## Step 3 – Rank (Percentile) Ordering by Sensitivity

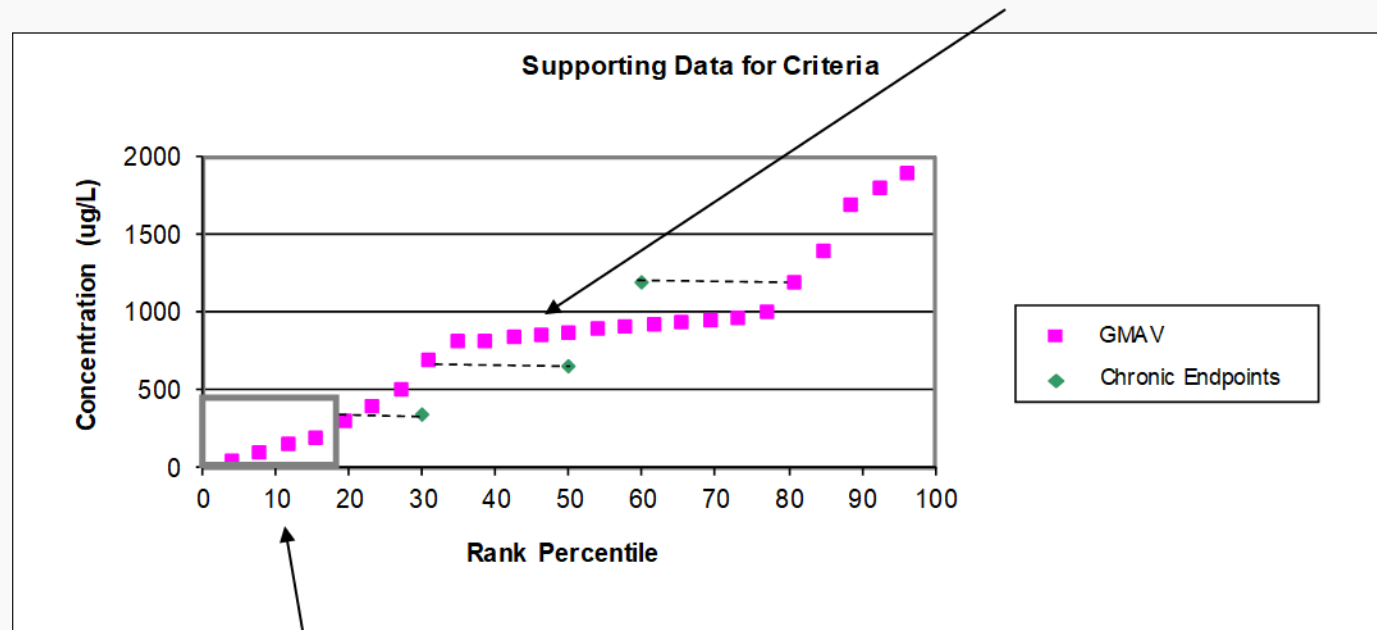
<u>RANK</u>	<u>GMAV</u>	<u>Species</u>	<u>SMAV</u>
4	100	<i>Oncorhynchus mykiss</i>	100
3	36	<i>Daphnia ambigua</i>	42
		<i>Daphnia pulex</i>	38
		<i>Daphnia magna</i>	29
2	25	<i>Gammarus pseudolimnaeus</i>	25
1	19	<i>Hyalella azteca</i>	19

# FAV Calculation Overview

- Step 1. Calculate Species Mean Acute Values (SMAVs)
  - geometric mean of all acceptable acute values for species
  
- Step 2. Calculate Genus Mean Acute Values
  - geometric mean of all SMAVs for genus
  
- Step 3. Rank Genus Mean Acute Values
  - from most sensitive (#1 = lowest concentration to see an effect) to least sensitive (n)
  
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# FAV Calculation

GMAV and Calculate the Percentile of each rank ( $100 R/(N+1)$ )

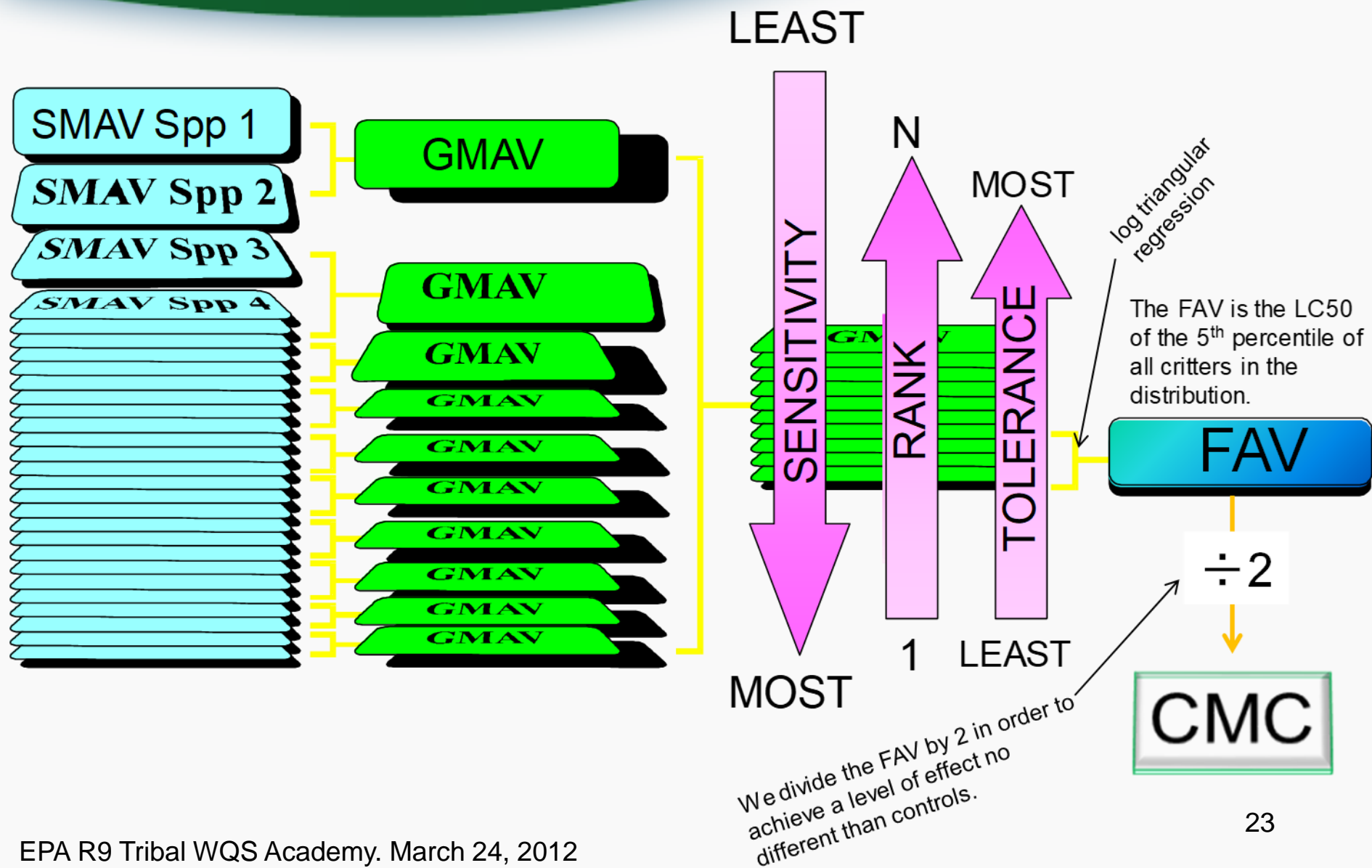


Using the 4 Most Sensitive Genera, Perform a Log Triangular Regression of the GMAV (log values) on the Percentile Ranks (square roots) to generate an  $HC_5 = FAV$

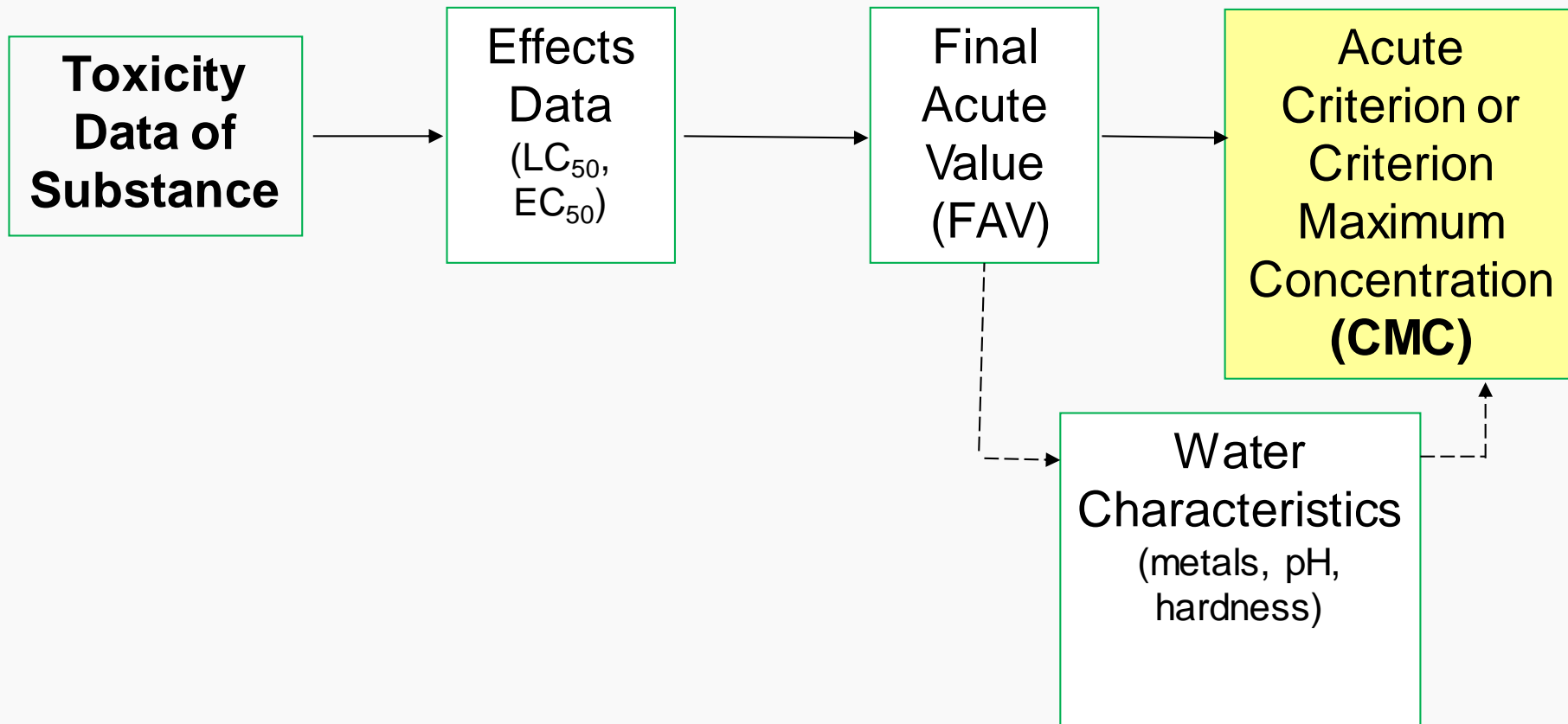
# Log Triangular Distribution

- Assume the available GMAVs follow a log-triangular distribution.
- Rank Order the GMAVs – least to most sensitive
- Assign Ranks (1 to N); Calculate Cumulative Probability  
 $P = R/(N+1)$
- Select the 4 GMAVs closest to 0.05 (often the lowest 4 GMAVs) – those are used to define the slope of the toxic response

# FAV Calculation Overview

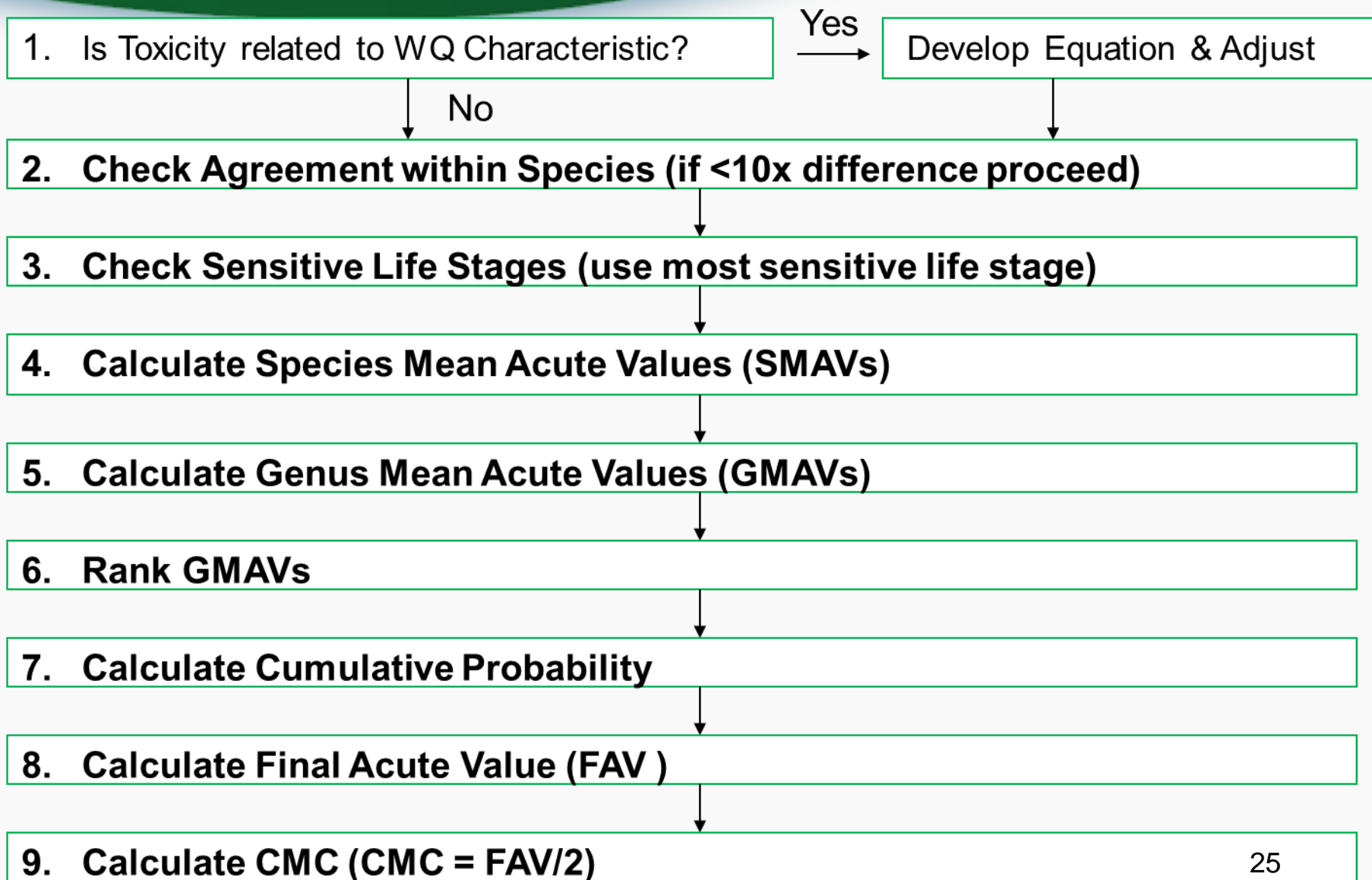


# Criterion Maximum Concentration (CMC)



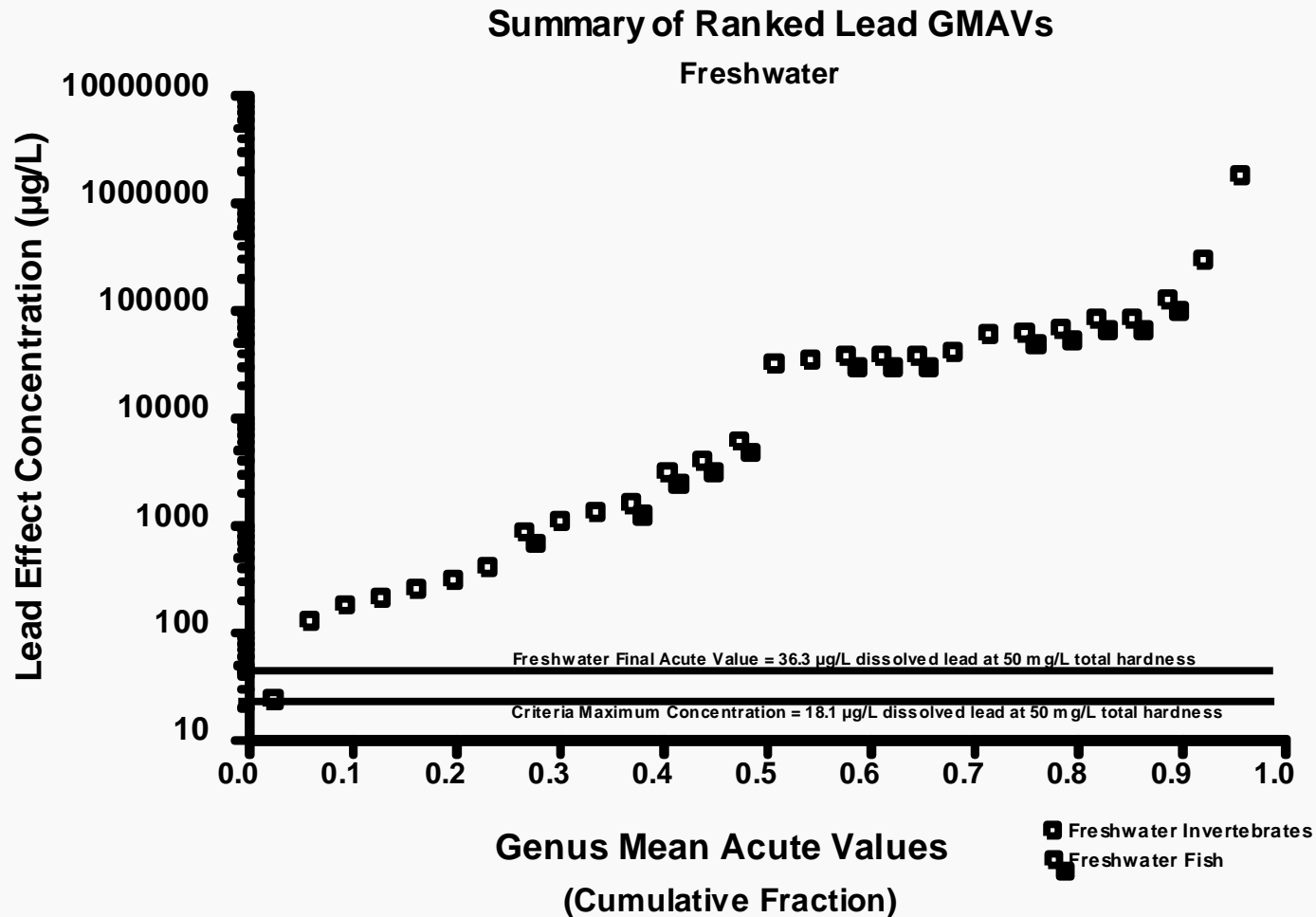


# Aquatic Life Criteria: Final Overview

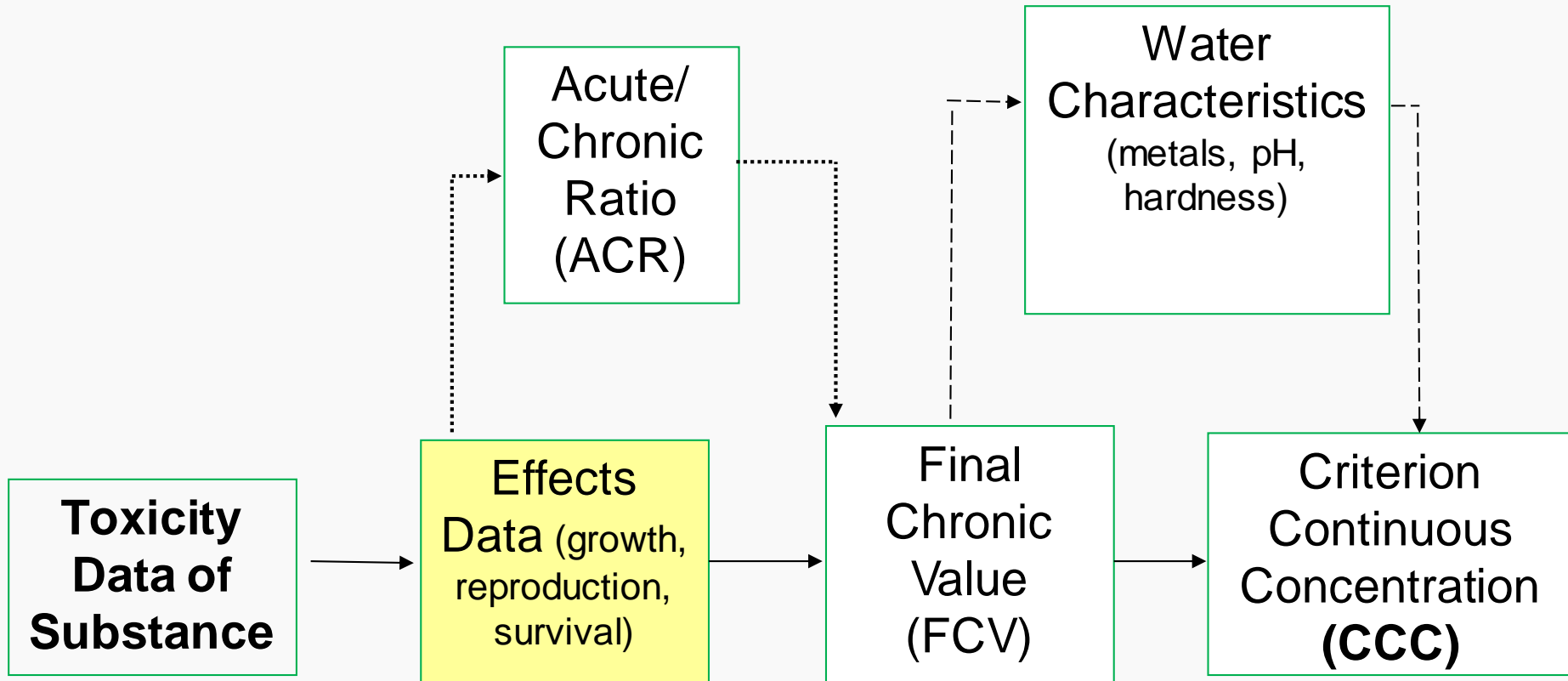


# Species Sensitivity Distribution

## Species Sensitivity Distribution (SSD)



# Chronic Criteria Derivation Overview



# Chronic Criteria Test Endpoints

- Species-appropriate test durations
  - Including full life cycle, early-life stage tests, etc as appropriate
- Endpoints include long term mortality, growth and reproduction
  - Or other endpoints that can be linked to those apical responses quantitatively
- Test endpoints include NOECs, LOECs, and EC20s

# Chronic Criteria Test Endpoint Calculation

Fathead Minnow Early Life Stage Test: Growth Measured as Length

**Chronic Value = 26.8  $\mu\text{g/L}$**

(NOEC = 15; LOEC = 30)

Concentration:

**Control**

**0.0  $\mu\text{g/L}$**

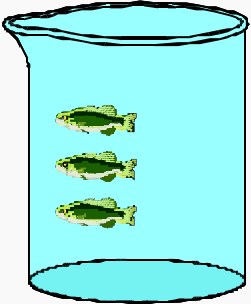
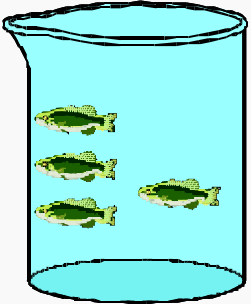
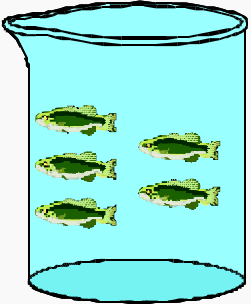
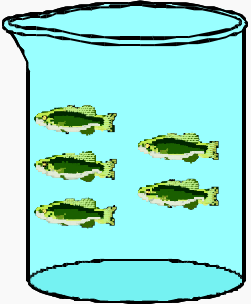
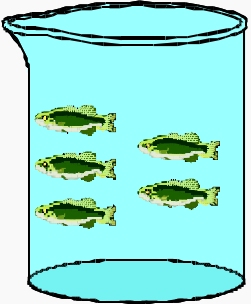
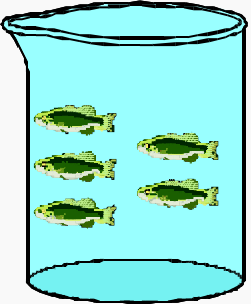
**3.8  $\mu\text{g/L}$**

**7.5  $\mu\text{g/L}$**

**15  $\mu\text{g/L}$**

**30  $\mu\text{g/L}$**

**60  $\mu\text{g/L}$**



Length:

**40 mm**

**41 mm**

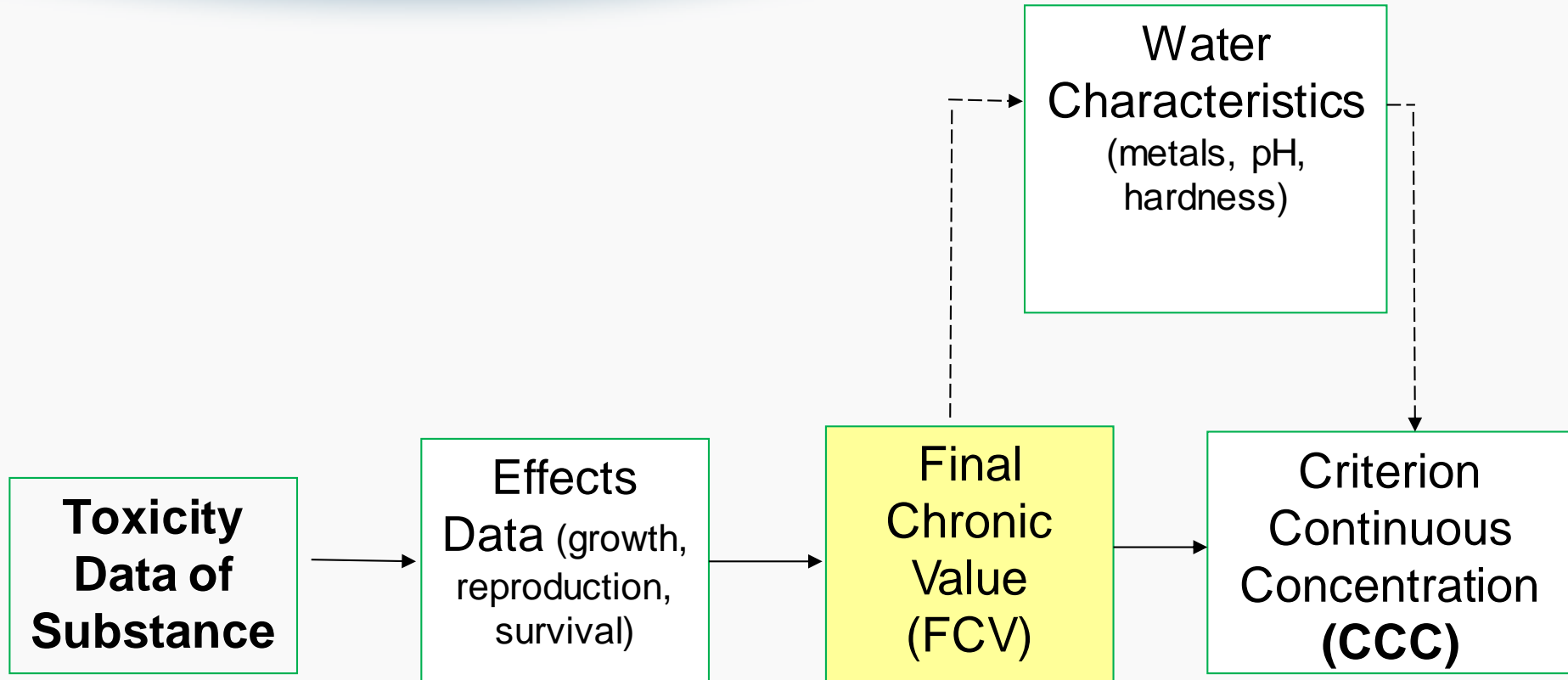
**38 mm**

**36 mm**

**20 mm**

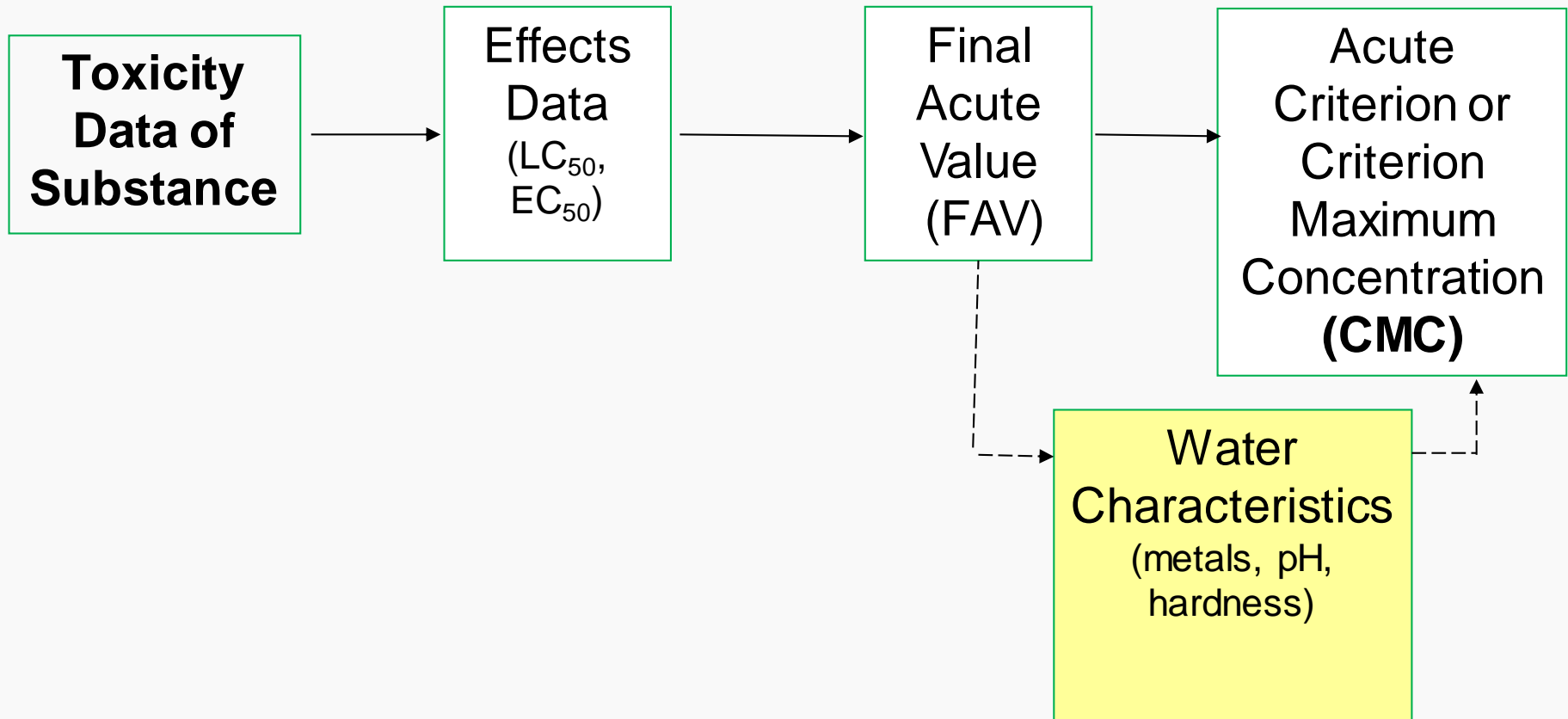
**5 mm**

# Final Chronic Value



FCV is calculated in the same manner as with acute criteria: values are ranked in sensitivity and then a regression is performed on the 4 most sensitive genera.

# Water Characteristics Affecting Criteria



# Water Chemistry Dependency

## Example: Cadmium Criteria Equation\*

$$= e^{(0.9789 \times (\ln \text{Hardness}) - 3.866)} \times \text{CF},$$

where CF (conversion factor from Total to Dissolved)

$$= 1.136672 - [(\ln \text{hardness}) \times (0.041838)]$$

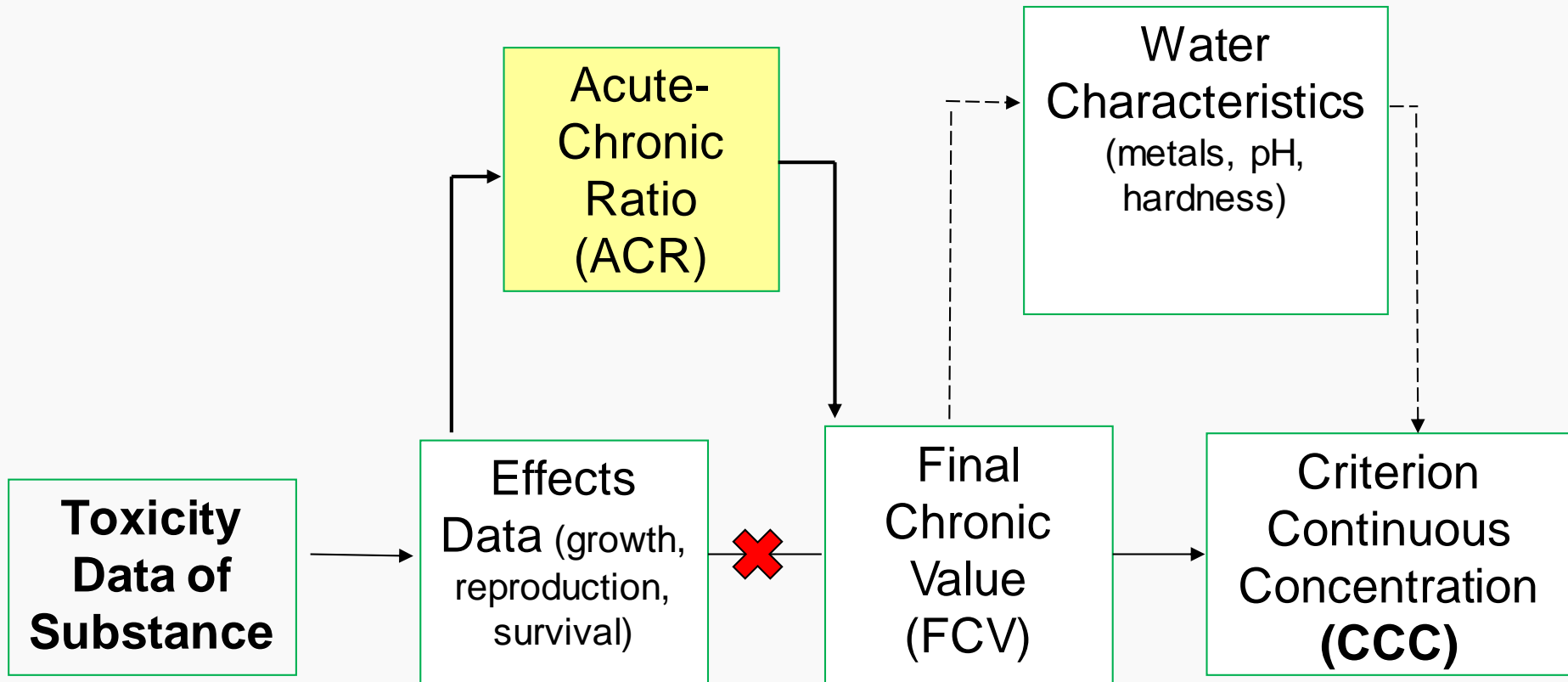
Hardness (mg/L)	Criteria Value ( $\mu\text{g/L}$ dissolved)
25	0.49
50	0.94
100	1.8
200	3.4

\* Based on dissolved concentration, 2016 Final Cd criteria document<sub>32</sub>



# Derivation of FCV when Insufficient Data

... OR ...



## Calculating and Applying the ACR

1. Acute & chronic tests using same species in same dilution water  
(*guidance on test matching and requirements in 1985 Guidelines*)



2. Use results of tests to calculate Acute-Chronic Ratios (ACR)

$$\text{ACR} = \frac{\text{Acute Value}}{\text{Chronic Value}}$$



3. Develop a Final Acute-Chronic Ratio (FACR) by taking a geometric mean of the appropriate ACRs (*3 minimum*)



4. Calculate the Final Chronic Value (FCV) using the FACR

$$\text{FCV} = \frac{\text{Final Acute Value}}{\text{FACR}}$$

# Site Specific Criteria (1)

Why would you develop a site-specific criterion?

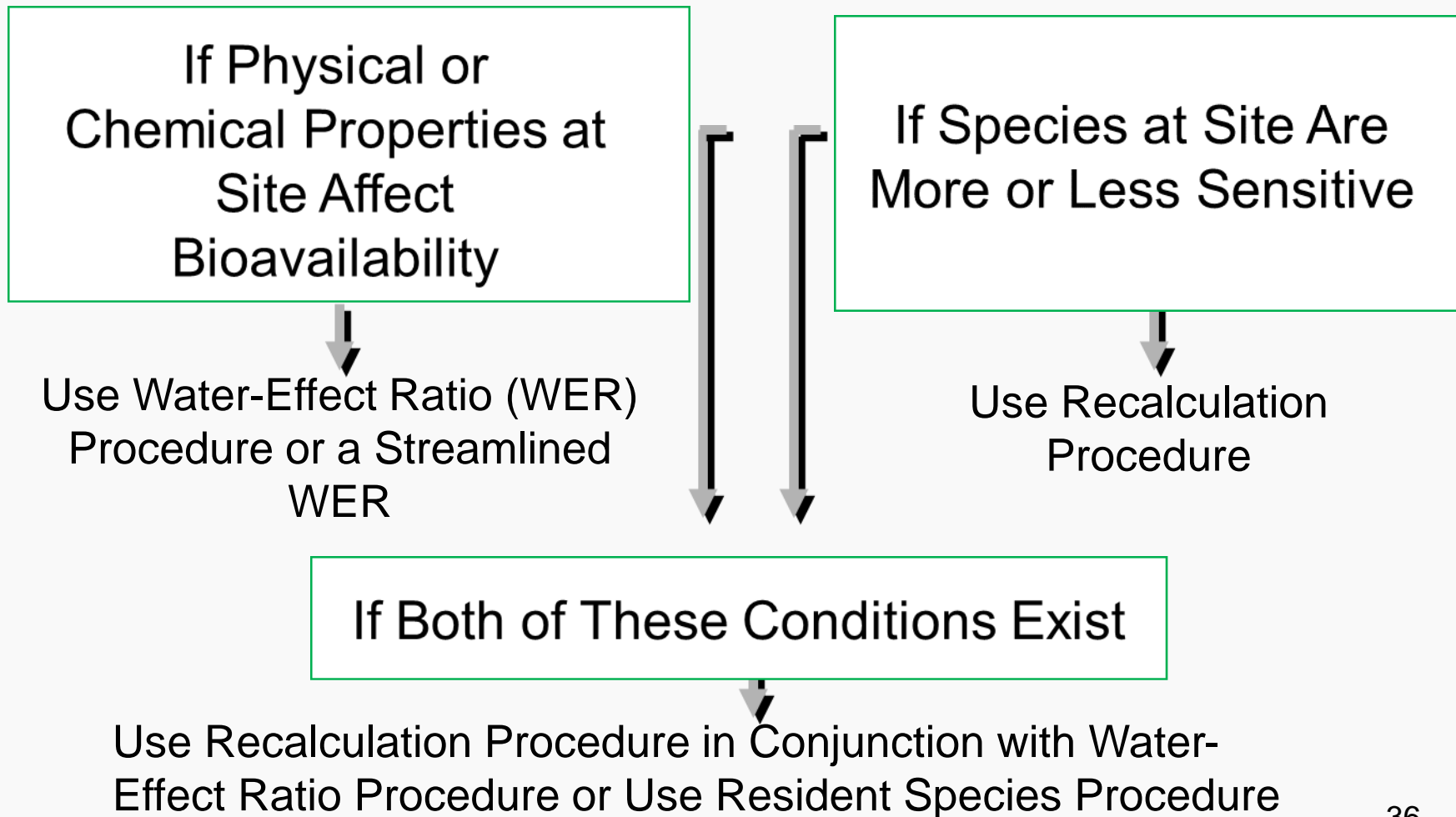
The sensitivities of the site-specific species differ from the national data set (e.g., that in the criteria document).

and/or

The physical/chemical characteristics of the site alter the bioavailability/toxicity of the pollutant.

# Site Specific Criteria (2)

## PROCEDURES



# BLMs, MLRs, & WERs

- Since the 1990s, the WER has been used to adjust hardness-based criteria for metals.
- In 2007, the biotic ligand model (BLM) was developed for copper.
  - The BLM models the bioavailability of copper based on 10 water chemistry parameters (pH, DOC, temperature, alkalinity and 6 geochemical ions (GI))
  - Implementation issues have slowed its widespread adoption and use
- Recently, a multiple linear regression method (MLR) was developed for aluminum, that simplifies the modeling necessary based on 3-4 water chemistry parameters.
  - EPA published a Final Revised Aluminum (December 2018) based on 3 water chemistry parameters (pH, DOC, and hardness)

# BLMs, MLRs, & WERs

- EPA recommends the use of the Biotic Ligand Model (BLM) for copper.
- EPA recommends the use of the Multiple Linear Regression (MLR) approach for aluminum.
- For other metals (Cd, Ni, Pb, Zn), a WER based on appropriately-conducted studies is still a potential approach for adjusting the respective hardness-based criteria at a site, until the criteria are updated to reflect fuller effects of water chemistry through bioavailability modeling ( e.g., MLR or BLM).
- EPA has entered into a Cooperative Research and Development Agreement (CRADA) with 8 metals associations to further the development of updated aquatic life criteria for the other metals.
  - <https://www.epa.gov/sites/production/files/2018-01/documents/metals-crada-summary-2018.pdf>

## EPA Review

- Internal Review
  - EPA Peer Review
    - Data quality of studies used in criteria derivation
    - Intra-agency workgroup
- External Review
  - External Peer Review (external experts – multiple views)
- Publication of Draft Recommended 304(a) criteria via the Federal Register
  - Request for scientific views on draft criteria from the public via Federal Register
- Publication of Final Recommended 304(a) criteria via the Federal Register
  - Response to Comments Document (Published with Final)

# State/Tribal Review and Adoption Process

- State Triennial Review/Adoption of criteria into WQS
- Public comments on draft WQS according to State/Tribal regulatory adoption process (Administrative Procedure Act)
- EPA approval of WQS
- WQS implementation – permits, TMDLs, monitoring and assessments



# Aquatic Life Exercise

*See Exercise 6*



# Data Table for Mineral X

Genus	species	Common name	SMAV (ug/L)	GMAV	ACR	taxa
Bufo	americanus	American toad	9805			chordata
Cragonyx	pseudogracilis	Amphipod	9190			benthic crustacean
Hyaella	azteca	Amphipod	1455			benthic crustacean
Salmo	salar	Atlantic salmon	879			salmonid
Salvelinus	fontinalis	Brook trout	6965			salmonid
Ictalurus	punctatus	Channel catfish	761			fish
Oncorhynchus	tshawytscha	Chinook salmon	11827			salmonid
Ceriodaphnia	dubia	Cladoceran	1718		4.31	planktonic crustacean
Daphnia	magna	Cladoceran	12329			planktonic crustacean
Enallagma		Damselfly	1075			insect
Pimephales	promelas	Fathead minnow	14100		6.06	fish
Lepomis	cyanellus	Green sunfish	52273			fish
Hyla	cinerea	Green tree frog	6171			chordata
Rana	pipiens	Leopard frog	8709			chordata
Tanytarus	dissimilis	Midge	186860		34.08	insect taxa 1
Pisidium	casertanum	Pea cockle	1072			mollusc
Oncorhynchus	mykiss	Rainbow trout	6901			salmonid
Pisidium	compressum	Ridged-beak peaclam	1072			mollusc
Hybognathus	amarus	Rio Grande minnow	25074			fish
Micropterus	dolomieu	Smallmouth bass	1662			fish
Amnicola	limosa	Snail	1080			mollusc
Physa		Snail	32907			mollusc

# Question 1

1. Are the required eight taxa represented in this dataset?

Genus	species	Common name	SMAV (ug/L)	GMAV	ACR	taxa
Ictalurus	punctatus	Channel catfish	761			fish
Salmo	salar	Atlantic salmon	879			salmonid
Pisidium	casertanum	Pea cockle	1072			mollusc
Enallagma		Damselfly	1075			insect
Amnicola	limosa	Snail	1080			mollusc
Pisidium	compressum	Ridged-beak peaclam	1110			mollusc
Hyalella	azteca	Amphipod	1455			benthic crustacean
Micropterus	dolomieu	Smallmouth bass	1662			fish
Ceriodaphnia	dubia	Cladoceran	1718		4.31	planktonic crustacean
Hyla	cinerea	Green tree frog	6171			chordata
Oncorhynchus	mykiss	Rainbow trout	6901			salmonid
Salvelinus	fontinalis	Brook trout	6965			salmonid
Rana	pipiens	Leopard frog	8709			chordata
Cragonyx	pseudogracilis	Amphipod	9190			benthic crustacean
Bufo	americanus	American toad	9805			chordata
Oncorhynchus	tshawytscha	Chinook salmon	11827			salmonid
Daphnia	magna	Cladoceran	12329			planktonic crustacean
Pimephales	promelas	Fathead minnow	14100		6.06	fish
Acronuria		Stonefly	23627			insect
Hybognathus	amarus	Rio Grande minnow	25074			fish
Physa		Snail	32907			mollusc
Lepomis	cyaneus	Green sunfish	52273			fish
Tanytarsus	dissimilis	Midge	186860		34.08	insect taxa 1

YES!

1. Salmonid
2. 2<sup>nd</sup> Fish
3. Chordate
4. Planktonic crustacean
5. Benthic crustacean
6. Insect
7. Rotifer/annelid/mollusc
8. Other insect/mollusc

# Question 2

## 2. What is the N for this data set?

Sort

Column Sort On Order

Sort by Genus Values A to Z

Genus	species	Common name	SMAV (ug/L)	GMAV	ACR	taxa
Acronuria		Stonefly	23627			insect
Amnicola	limosa	Snail	1080			mollusc
Bufo	americanus	American toad	9805			chordata
Ceriodaphnia	dubia	Cladoceran	1718		4.31	planktonic crustacean
Cragonyx	pseudogracilis	Amphipod	9190			benthic crustacean
Daphnia	magna	Cladoceran	12329			planktonic crustacean
Enallagma		Damselfly	1075			insect
Hyalella	azteca	Amphipod	1455			benthic crustacean
Hybognathus	amarus	Rio Grande minnow	25074			fish
Hyla	cinerea	Green tree frog	6171			chordata
Ictalurus	punctatus	Channel catfish	761			fish
Lepomis	cyaneus	Green sunfish	52273			fish
Micropterus	dolomieu	Smallmouth bass	1662			fish
Oncorhynchus	mykiss	Rainbow trout	6901			salmonid
Oncorhynchus	tshawytscha	Chinook salmon	11827			salmonid
Physa		Snail	32907			mollusc
Pimephales	promelas	Fathead minnow	14100		6.06	fish
Pisidium	casertanum	Pea cockle	1072			mollusc
Pisidium	compressum	Ridged-beak peaclam	1110			mollusc
Rana	pipiens	Leopard frog	8709			chordata
Salmo	salar	Atlantic salmon	879			salmonid
Salvelinus	fontinalis	Brook trout	6965			salmonid
Tanytarsus	dissimilis	Midge	186860		34.08	insect taxa 1

21

Number (N) is the number of GMAVs.  
GMAV = genus mean acute values

# Questions 4 and 5

4 & 5. What is the most/least sensitive Genus?

Most = Ictalurus

Least = Tanytarus

Table 1a. Acute Values with Species Mean Acute-Chronic Ratios.  
Freshwater normalized to a hardness of 50 mg/L.

Mineral X				
Genus	species	Common name	SMAV (uq/L)	GMAV
Ictalurus	punctatus	Channel catfish	761	761
Salmo	salar	Atlantic salmon	879	879
Enallagma		Damselfly	1075	1075
Amnicola	limosa	Snail	1080	1080
Pisidium	casertanum	Pea cockle	1072	1090
Hyaella	azteca	Amphipod	1455	1455
Micropterus	dolomieu	Smallmouth bass	1662	1662
Ceriodaphnia	dubia	Cladoceran	1718	1718
Hyla	cinerea	Green tree frog	6171	6171
Salvelinus	fontinalis	Brook trout	6965	6965
Rana	pipiens	Leopard frog	8709	8709
Oncorhynchus	mykiss	Rainbow trout	9001	9034
Crangonyx	pseudogracilis	Amphipod	9190	9190
Bufo	americanus	American toad	9805	9805
Daphnia	magna	Cladoceran	12329	12329
Pimephales	promelas	Fathead minnow	14100	14100
Acronuria		Stonefly	23627	23627
Hybognathus	amarus	Rio Grande minnow	25074	25074
Physa		Snail	32907	32907
Lepomis	cyanellus	Green sunfish	52273	52273
Tanytarus	dissimilis	Midge	186860	186860
Pisidium	compressum	Ridged-beak peaclam	1110	
Oncorhynchus	tshawytscha	Chinook salmon	11827	

