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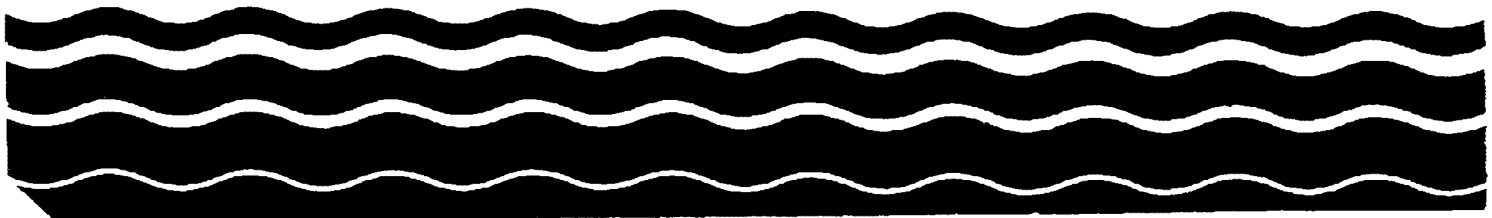
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**Ambient
Water Quality
Criteria
for**

Mercury - 1984



AMBIENT AQUATIC LIFE WATER QUALITY CRITERIA FOR
MERCURY

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FOREWORD

Section 304(a)(1) of the Clean Water Act of 1977 (P.L. 95-217) requires the Administrator of the Environmental Protection Agency to publish criteria for water quality accurately reflecting the latest scientific knowledge on the kind and extent of all identifiable effects on health and welfare which may be expected from the presence of pollutants in any body of water, including ground water. This document is a revision of proposed criteria based upon a consideration of comments received from other Federal agencies, State agencies, special interest groups, and individual scientists. The criteria contained in this document replace any previously published EPA aquatic life criteria.

The term "water quality criteria" is used in two sections of the Clean Water Act, section 304(a)(1) and section 303(c)(2). The term has a different program impact in each section. In section 304, the term represents a non-regulatory, scientific assessment of ecological effects. The criteria presented in this publication are such scientific assessments. Such water quality criteria associated with specific stream uses when adopted as State water quality standards under section 303 become enforceable maximum acceptable levels of a pollutant in ambient waters. The water quality criteria adopted in the State water quality standards could have the same numerical limits as the criteria developed under section 304. However, in many situations States may want to adjust water quality criteria developed under section 304 to reflect local environmental conditions and human exposure patterns before incorporation into water quality standards. It is not until their adoption as part of the State water quality standards that the criteria become regulatory.

Guidelines to assist the States in the modification of criteria presented in this document, in the development of water quality standards, and in other water-related programs of this Agency, have been developed by EPA.

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Introduction*

Mercury has long been recognized as one of the most toxic of the heavy metals, but only recently was it identified as a serious pollutant in the aquatic environment (National Research Council, 1978; National Research Council Canada, 1979; Nriagu, 1979). Elemental mercury is a heavy liquid at room temperature and was considered relatively inert, because it was assumed that it would quickly settle to the bottom of a body of water and remain there in an innocuous state. However, elemental mercury is oxidized to mercury(II) under natural conditions (Wood, 1974). Furthermore, mercury(II), whether discharged directly or produced from elemental mercury, can be methylated by both aerobic and anaerobic bacteria (Akagi, 1979; Beijer and Jernelov, 1979; Callahan, et al. 1979; Jernelov, 1971, 1972; Jernelov, et al. 1975; National Research Council, 1978; Summers and Silver, 1978; Thayer and Brinckman, 1982; Wright and Hamilton, 1982). Mercury(II) can also be methylated in the slime coat, liver, and intestines of fish (Jernelov, 1968; Matsumura, et al. 1975; Rudd, et al. 1980b), but methylation apparently does not occur in other tissues (Huckabee, et al. 1978; Matida, et al. 1971; Pennacchioni, et al. 1976) or in plants (Czuba and Mortimer, 1980). (The term "methylmercury" is used herein to refer only to monomethylmercury, and not to dimethylmercury or any other monoorganomercury salt or diorganomercury compound. Inorganic mercury(II) will be referred to as "mercury(II)".)

The importance of methylation may be reduced by demethylation (Bisogni, 1979; Ramamoorthy, et al. 1982). Demethylation might provide a feedback

*An understanding of the "Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses" (Stephan, et al., 1985), hereafter referred to as the Guidelines, is necessary in order to understand the following text, tables, and calculations.

mechanism that controls the concentration of methylmercury in sediment and in water. Jernelov, et al. (1975) cited a case in which low levels of methylmercury in fish from a highly contaminated area coincided with strong methylmercury degrading activity in the sediment. Demethylation also occurs in fish (Burrows and Krenkel, 1973; de Freitas, et al. 1981; Gage, 1964; Olson, et al. 1978), probably as part of the depuration mechanism.

Numerous factors such as alkalinity, ascorbic acid, chloride, dissolved oxygen, hardness, organic complexing agents, pH, sediment, and temperature probably affect the acute and chronic toxicity and bioaccumulation of the various forms of mercury (Amend, et al. 1969; Baker, et al. 1983; Feich, et al. 1972; Hahne and Kroontje, 1973; Jernelov, 1980; Ramamoorthy and Blumhagen, 1984; Ribeyre and Boudou, 1982; Rogers and Beamish, 1981, 1983; Rudd, et al. 1980a; Rudd and Turner, 1983a,b; Sharma, et al. 1982; Stokes, et al. 1983; Tsai, et al. 1975; Wren and MacCrimmon, 1983; Wright and Hamilton, 1982).

A variety of studies have been conducted on the effect of selenium on the acute toxicity of mercury (e.g., Birge, et al. 1981; Bowers, et al. 1980; De Filippis, 1979; Heisinger, 1981; Heisinger, et al. 1979; Klaverkamp, et al., 1983a; Lawrence and Holoka, 1983; Sharma and Davis, 1980c) and on the accumulation of mercury from food and water (e.g., Beijer and Jernelov, 1978; Chang, et al. 1983; Heisinger, et al. 1979; Klaverkamp, et al. 1983b; Rudd, et al. 1980a; Sharma and Davis, 1980c; Speyer, 1980; Turner and Swick, 1983). Available data do not, however, show that quantitative relationships are consistent enough for a variety of aquatic species to enable relating water quality criteria to any of these variables.

Because of the variety of forms of inorganic and organic mercury and lack of definitive information about their relative toxicities, no available

analytical measurement is known to be ideal for expressing aquatic life criteria for mercury. Previous aquatic life criteria for mercury (U.S. EPA, 1980) were specified in terms of total recoverable mercury, which would probably be measured as total mercury (U.S. EPA, 1983a), but both of these measurements are probably too rigorous in some situations. Acid-soluble mercury (operationally defined as the mercury that passes through a 0.45 μ m membrane filter after the sample is acidified to pH = 1.5 to 2.0 with nitric acid) is probably the best measurement at the present for the following reasons:

1. This measurement is compatible with all available data concerning toxicity of mercury to, and bioaccumulation of mercury by, aquatic organisms. No test results were rejected just because it was likely that they would have been substantially different if they had been reported in terms of acid-soluble mercury. For example, results reported in terms of dissolved mercury would not have been used if the concentration of precipitated mercury was substantial.
2. On samples of ambient water, measurement of acid-soluble mercury should measure all forms of mercury that are toxic to aquatic life or can be readily converted to toxic forms under natural conditions. In addition, this measurement should not measure several forms, such as mercury that is occluded in minerals, clays, and sand or is strongly sorbed to particulate matter, that are not toxic and are not likely to become toxic under natural conditions. Although this measurement (and many others) will measure soluble, complexed forms of mercury, such as the EDTA complex of mercury(II), that probably have low toxicities to aquatic life, concentrations of these forms probably are negligible in most ambient water.

3. Although water quality criteria apply to ambient water, the measurement used to express criteria is likely to be used to measure mercury in aqueous effluents. Measurement of acid-soluble mercury should be applicable to effluents because it will measure precipitates, such as carbonate and hydroxide precipitates of mercury(II), that might exist in an effluent and dissolve when the effluent is diluted with receiving water. If desired, dilution of effluent with receiving water before measurement of acid-soluble mercury might be used to determine whether the receiving water can decrease the concentration of acid-soluble mercury because of sorption.
4. The acid-soluble measurement should be useful for most metals, thus minimizing the number of samples and procedures that are necessary.
5. The acid-soluble measurement does not require filtration at the time of collection, as does the dissolved measurement.
6. The only treatment required at the time of collection is preservation by acidification to pH = 1.5 to 2.0, similar to that required for the total measurement.
7. Durations of 10 minutes to 24 hours between acidification and filtration probably will not affect the result substantially.
8. The carbonate system has a much higher buffer capacity from pH = 1.5 to 2.0 than it does from pH = 4 to 9 (Weber and Stumm, 1963).
9. Differences in pH within the range of 1.5 to 2.0 probably will not affect the result substantially.
10. After acidification and filtration of the sample to isolate the acid-soluble mercury, the analysis for total acid-soluble mercury can be performed using permanganate and persulfate oxidation and cold vapor atomic absorption (U.S. EPA, 1983a), as with the total measurement.

Acid-soluble inorganic mercury can be measured by not breaking down the organomercury compounds before using cold vapor atomic absorption.

Methylmercury has been measured using gas chromatography (Cappon, 1984; Hildebrand, et al. 1980; Paasivirta, et al. 1981), thin layer chromatography (Kudo, et al. 1982), and liquid chromatography (Gast and Kraak, 1979; MacCrehan and Durst, 1978).

Thus, expressing aquatic life criteria for mercury in terms of the acid-soluble measurement has both toxicological and practical advantages. On the other hand, because no measurement is known to be ideal for expressing aquatic life criteria for mercury or for measuring mercury in ambient water or aqueous effluents, measurement of both total acid-soluble mercury and total mercury in ambient water or effluent or both might be useful. For example, there might be cause for concern if total mercury is much above an applicable limit, even though total acid-soluble mercury is below the limit.

Unless otherwise noted, all concentrations reported herein are expected to be essentially equivalent to acid-soluble mercury concentrations. All concentrations are expressed as mercury, not as the chemical tested. The criteria presented herein supersede previous aquatic life water quality criteria for mercury (U.S. EPA, 1976, 1980) because these new criteria were derived using improved procedures and additional information. Whenever adequately justified, a national criterion may be replaced by a site-specific criterion (U.S. EPA, 1983b), which may include not only site-specific criterion concentrations (U.S. EPA, 1983c), but also site-specific durations of averaging periods and site-specific frequencies of allowed exceedences (U.S. EPA, 1985). The latest literature search for information for this document was conducted in May, 1984; some newer information was also used.

Acute Toxicity to Aquatic Animals

Table 1 contains the primary acute toxicity data for three classes of mercury compounds: mercury(II), methylmercury, and other mercury compounds, chiefly organic. The latter information exists principally because many of these compounds were considered for use in treatment of diseases and control of parasites in fish culture, although their source for environmental concern is from industrial and agricultural uses for fungus control. Both phenylmercuric acetate and pyridylmercuric acetate have been called PMA. Tests have been conducted on different formulations which contain various percentages of active ingredient and the percentages of active ingredient given by the authors were used to calculate mercury concentrations. When the percentage of active ingredient was not given for pyridylmercuric acetate, 80 percent was assumed (Allison, 1957).

The freshwater acute toxicity values indicate that the difference in sensitivity between different species to a particular mercury compound is far greater than the difference in sensitivity of a particular species to various mercury compounds. For example, the reported acute values for mercury(II) range from 2.217 $\mu\text{g/L}$ for Daphnia pulex to 2,000 $\mu\text{g/L}$ for the aquatic stages of certain insects, with a continual gradation in sensitivity among intermediate species (Table 3). On the other hand, Joshi and Rege (1980), Lock and van Overbeeke (1981) and Matida, et al. (1971) found that various species were 4 to 31 times more sensitive to various organic mercury compounds than to mercuric chloride (Table 1).

MacLeod and Pessah (1973) studied the effect of temperature on the acute toxicity of mercuric chloride to rainbow trout. At 5, 10, and 15 C, the LC50s were 400, 280, and 220 $\mu\text{g/L}$, respectively (Tables 1 and 6). Clemens and Sneed (1958b) found a similar effect of temperature on toxicity to

juvenile channel catfish; at 10, 16.5, and 24 C the acute values for phenylmercuric acetate were 1,960, 1,360, and 233 $\mu\text{g/L}$, respectively (Table 6).

The 28 Genus Mean Acute Values in Table 3 were calculated as geometric means of the available Species Mean Acute Values (Tables 1 and 3). Acute values are available for more than one species in each of two genera, and the range of Species Mean Acute Values within each genus is less than a factor of 1.6. On the other hand, a midge was among the most sensitive species, whereas other insects were the most resistant species. The most sensitive genus, Daphnia, is 756 times more sensitive than the most resistant, Acroneuria (Table 3). A freshwater Final Acute Value of 4.857 $\mu\text{g/L}$ was obtained for mercury(II) using the Genus Mean Acute Values in Table 3 and the calculation procedure described in the Guidelines. Not enough data are available to calculate a Final Acute Value for methylmercury, but the available data indicate that it is more acutely toxic than mercury(II).

Saltwater fishes and invertebrates both show wide ranges of sensitivities to mercury(II). Acute values for fishes range from 36 $\mu\text{g/L}$ for spot to 1,678 $\mu\text{g/L}$ for winter flounder (Tables 1 and 3). Among invertebrates a mysid has an acute value of 3.5 $\mu\text{g/L}$, whereas the value for the soft-shell clam is 400 $\mu\text{g/L}$. Of the 29 saltwater genera for which acute values are available, the most sensitive, Mysidopsis, is 479 times more sensitive than the most resistant, Pseudopleuronectes. Acute values are available for more than one species in each of three genera and the range of Species Mean Acute Values within each genus is less than a factor of 1.7. The saltwater Final Acute Value of 4.125 $\mu\text{g/L}$ was calculated for mercury(II) from the Genus Mean Acute Values in Table 3.

Chronic Toxicity to Aquatic Animals

Chronic toxicity tests with Daphnia magna have been conducted on three mercury compounds (Table 2). The renewal and flow-through techniques produced similar results for mercury(II), but the renewal technique produced much higher results for methylmercury, presumably because of volatility. In addition, a chronic test with brook trout on methylmercuric chloride yielded a chronic value of 0.5193 $\mu\text{g/L}$. Both an early life-stage test and a life-cycle test on mercuric chloride found adverse effects on the fathead minnow at all concentrations tested including the lowest of 0.23 $\mu\text{g/L}$. For mercuric chloride the acute-chronic ratio with Daphnia magna is less than 6, whereas that with the fathead minnow is greater than 600. For methylmercury the acute-chronic ratio with brook trout is 142.3.

A chronic value of 1.131 $\mu\text{g/L}$ was obtained in a flow-through life-cycle exposure of a mysid to mercuric chloride (Table 2). Groups of 30 juvenile mysids were reared in each of 5 concentrations for 36 days at 21 C and a salinity of 30 g/kg. Effects examined included time to first spawn and productivity (total number of young/number of available female spawning days and total number of spawns/number of available female spawning days). No spawning occurred at 2.5 $\mu\text{g/L}$. Time to spawn and productivity at 1.6 $\mu\text{g/L}$ were significantly different from the controls. The highest concentration at which no statistically significant effect on reproductive processes was detected was 0.8 $\mu\text{g/L}$. Therefore, the chronic limits are 0.8 and 1.6 $\mu\text{g/L}$ and the chronic value is 1.131 $\mu\text{g/L}$. The 96-hr LC50 for this species in the same study was 3.5 $\mu\text{g/L}$, giving an acute-chronic ratio of 3.095 (Table 2).

The species mean acute-chronic ratio for Daphnia magna is 4.498, whereas that for the mysid is 3.095 (Table 3). These are sensitive species in fresh and salt water, respectively, and the four most sensitive species in each

water are invertebrates. Thus, it seems reasonable to use the geometric mean of these two values as the Final Acute-Chronic Ratio (Table 3). Division of the Final Acute Values by 3.731 results in freshwater and saltwater Final Chronic Values of 1.302 and 1.106 $\mu\text{g/L}$, respectively. Even though the fathead minnow was considerably less sensitive than Daphnia magna in acute tests, the acute-chronic ratio for the fathead minnow is so high that its chronic value is below the Final Chronic Value and probably below the Final Residue Value (see below). If the acute-chronic ratio of greater than 649 for the fathead minnow is representative of ratios for other freshwater and saltwater fishes, then twelve of fourteen tested fish species, including the rainbow trout, coho salmon, bluegill, and haddock, would have chronic values below the Final Chronic Value. Various values for vertebrates in Table 6 are below the Final Chronic Value or are indicative of large acute-chronic ratios.

Toxicity to Aquatic Plants

Whereas some freshwater plant values for mercury(II) are about 1,000 $\mu\text{g/L}$ (Table 4), effects of mercury(II) and methylmercury have been observed at concentrations below 10 $\mu\text{g/L}$, respectively (Table 6). Some organomercury compounds have affected algae at concentrations less than 1.0 $\mu\text{g/L}$ (Table 6). Although freshwater plants are relatively insensitive to mercury(II) and sensitive to methylmercury, they do not appear to be more sensitive to methylmercury than freshwater animals.

Data concerning the toxicity of mercuric chloride to saltwater plants are from four studies with eight species of algae. The EC50s (Table 4) indicate reduction in growth at concentrations ranging from 10 to 160 $\mu\text{g/L}$. No data are available concerning the toxicities of organomercury compounds to saltwater plants.

Bioaccumulation

Bioconcentration is a function of the relative rates of uptake and depuration. The bioconcentration factor (BCF) of mercury is high for fish because uptake is relatively fast and depuration is very slow. Thus, the biological half-life of mercury in fish is approximately 2 to 3 years (de Freitas, et al. 1974, 1977; Jarvenpaa, et al. 1970; Lockhart, et al. 1972; McKim, et al. 1976; Mellinger, 1973; Ruohtula and Miettinen, 1975; Sharpe, et al. 1977). Depuration of mercury is so slow that, even in the absence of exposure to mercury, long-term reduction in the concentration of mercury in fish tissue is largely due to dilution by tissue addition from growth. Usually less than 60 percent of mercury in invertebrates is methylated, but in fish, except for young fish, usually more than 70 percent is methylated (Bache, et al. 1971; Baluja, et al. 1983; Busch, 1983; Cappon, 1984; Cappon and Smith, 1982a,b; Haccula, et al. 1978; Hildebrand, et al. 1980; Huckabee, et al. 1979; Kudo, et al. 1982; Lucen, et al. 1980; Paasivirta, et al. 1983).

The distribution of mercury within a fish is the result of the movement of mercury from the absorbing surfaces (gills, skin, and gastrointestinal tract), into the blood, then to the internal organs, and eventually either to the kidney or bile for recycling or elimination or to muscle for long-term storage. As the tissue concentration approaches steady-state, net accumulation rate is slowed either by a reduction in uptake rate, possibly due to inhibition of membrane transport, or by an increase in depuration rate, possibly because of a saturation of storage sites, or both.

High concentrations of mercury in the slime coat of certain freshwater fishes, such as burbot, eels, and northern pike, and in the skin of acutely-exposed fishes are believed to be due to the methylating activity of bacteria

prevalent in the mucous coat (Jernelov, 1968). In addition, acutely toxic concentrations of mercury have been reported to stimulate secretion of mucus (Baker, 1973; Lock, et al. 1981; McKone, et al. 1971). When acutely exposed fish are placed in mercury-free water, the skin quickly loses mercury, probably because of elimination of mercury(II) and sloughing of the slime coat (Burrows and Krenkel, 1973; Burrows, et al. 1974). The skin and mucous coat are in direct contact with mercury in water and can accumulate proportionately more mercury during short exposure than muscle. During long exposure there is sufficient time for mercury to reach more permanent storage sites.

Because sorption at the gill surface is a major pathway of mercury into aquatic organisms (Fromm, 1977), increases in temperature and activity cause increases in metabolic rate and ventilation rate and, therefore, uptake rate (de Freitas and Hart, 1975; Rogers and Beamish, 1981). The relationship between temperature and tissue residues seems to apply primarily before steady-state is reached (Reinert, et al. 1974) but also to some extent at steady-state (Boudou, et al. 1979; Cember, et al. 1978; Hartung, 1976). The latter is difficult to understand if steady-state results from saturation of available binding sites. Apparently not only are uptake and depuration accelerated by temperature (Ruohtula and Miettinen, 1975), but higher tissue residues also occur at higher temperatures, possibly because the uptake rate increases proportionately more than the depuration rate.

Similarly, low concentrations of dissolved oxygen are likely to increase both respiration rate and uptake rate. Larson (1976) found that the low concentration of dissolved oxygen in a eutrophic lake forced fishes into warmer surface waters to secure adequate oxygen. The warmer surface water apparently stimulated metabolic rate and increased mercury uptake.

Increased metabolic rate increases not only ventilation rate but also energy demand and thus increases food consumption and exposure to mercury through the food chain. Uptake through both the gills and the digestive tract is significant for fish, and some data suggest that tissue residues are higher in organisms exposed via both routes than via either separately (Boudou, et al. 1979; Phillips and Buhler, 1978). The relative importance of uptake from food for various fish species depends on such things as assimilation efficiency (Phillips and Gregory, 1979), body size, growth rate, and life span (Sharpe, et al. 1977), and diet (Murray, 1978). Although Murray (1978) did find different concentrations of mercury in different fish species, Huckabee (1972) and Huckabee, et al. (1974) found similar concentrations in both forage and game fish in the same environment.

Haines (1981) reported that acid rain tends to scour more mercury from the air. Acidification of a body of water might also increase mercury residues in fish even if no new input of mercury occurs, possibly because lower pH increases ventilation rate and membrane permeability, accelerates the rates of methylation and uptake, affects partitioning between sediment and water, or reduces growth or reproduction of fish (Akielaszek and Haines, 1981; Fromm, 1980; Hahne and Kroontje, 1973; Jernelov, 1980; Miller and Akagi, 1979; Ribeyre and Boudou, 1982; Rudd and Turner, 1983b; Scheider, et al. 1979; Stokes, et al. 1983; Tsai, et al. 1975; Wrenn and MacCrimmon, 1983). However, Heiskary and Helwig (1983) did not find a relation between pH and mercury in fish.

The available information (e.g., Boudou and Ribeyre, 1981; Boudou, et al. 1977, 1980; de Freitas, et al. 1981; Hamdy and Prabhu, 1979; Hamelink, et al. 1977; Herrick, et al. 1982; Huckabee, et al. 1979; Jernelov and Lann, 1971; Klaverkamp, et al. 1983c; MacCrimmon, et al. 1983; Norstrom, et al.

1976; Phillips, et al. 1980; Ribeyre, et al. 1980; Rogers and Beamish, 1982; Rogers and Qadri, 1982) indicates that the importance of uptake from food probably depends on the form and concentration of mercury in the diet and on the size and metabolic rate of the fish. Transfer of mercury from fish to wildlife predators has also been observed (Heinz, et al. 1980; Kucera, 1983; Wren, et al. 1983).

The available freshwater BCFs are listed in Tables 5 and 6. Table 5 contains BCFs only from those studies in which the exposure concentrations were adequately measured and the tissue residues reached steady-state or the test lasted longer than 27 days. Although the BCFs presented in Table 6 do not meet all these conditions, they do provide information on BCFs for plants and illustrate the very important influence of temperature on bioconcentration.

McKim, et al. (1976) studied the uptake of methylmercury into various tissues of brook trout. At concentrations in water of 0.93, 0.29, 0.09 and 0.03 $\mu\text{g/L}$ the resulting concentrations in muscle after 273 days were 10,000, 5,000, 1,900, and 1,000 $\mu\text{g/kg}$ and the corresponding BCFs were 11,000, 17,000, 21,000, and 33,000, respectively. Because the concentration of mercury in the muscle did not decrease as much as the concentration in water, the BCFs increased as the concentration in water decreased. A possible explanation for an inverse relationship between concentration in water and BCF is that steady-state results from saturation of available binding sites (Cember, et al. 1978). The maximum concentration in tissue would then be dependent on the number of available binding sites and would be independent of the concentration of mercury in water. If the concentration in tissue were constant, the BCF would be inversely proportional to the concentration in water. However, neither the concentration in tissue nor the BCF was constant. The

comparable BCFs for the whole body were 10,000, 12,000, 12,000, and 23,000 and are lower than those for muscle. The fish were adversely affected at 0.93 $\mu\text{g/L}$, but for both muscle and whole body, the BCF at this concentration is in line with the other BCFs. Even though concentrations up to 0.29 $\mu\text{g/L}$ did not cause statistically significant adverse effects, the concentration of mercury in fish exposed to 0.03 $\mu\text{g/L}$ were at the FDA action level.

Olson, et al. (1975) obtained much higher BCFs for methylmercury with the fathead minnow, and the BCF was also concentration-dependent. As the concentration in the water decreased from 0.247 $\mu\text{g/L}$ to 0.018 $\mu\text{g/L}$, the concentration in the fish decreased from 10,900 $\mu\text{g/kg}$ to 1,470 $\mu\text{g/kg}$, but the BCF decreased from 44,100 to 81,700. The contrast between the results with the fathead minnow at 25 C (Olson, et al. 1975) and brook trout at 9 to 15 C (McKim, et al. 1976) is one of considerable interest and potential importance. The trout were fed pelleted feed, and so had little opportunity for food chain input. In contrast, the fathead minnow is a browser and probably fed not only on the introduced food but also on the Aufwuchs growing in the test solution to which mercury had been added. Thus the higher BCFs for the fathead minnow might be more representative of field situations in which fish are exposed to methylmercury via both the water and food (Phillips and Buhler, 1978; Phillips and Gregory, 1979; Rogers and Beamish, 1982). Also, because temperature affects bioconcentration, the fathead minnow might be more representative of commonly consumed warmwater fishes.

In a 75-day test, Niimi and Lowe-Jinde (1984) found 12 mg/kg in the whole body of rainbow trout exposed to 0.15 μg methylmercury/L (0.14 μg mercury/L) in water and 18 μg mercury/kg in food. The BCF of 85,700 is higher than the highest BCF obtained by Olson, et al. (1975). However, at 0.012 $\mu\text{g/L}$ and 18 $\mu\text{g/kg}$, they found 0.053 mg/kg in the trout. This BCF of

4,077 is lower than the lowest BCF obtained by McKim, et al. (1976). Also, although both McKim, et al. (1976) and Olson, et al. (1975) found higher BCFs at lower concentrations in water, Niimi and Lowe-Jinde (1984) found just the opposite.

The FDA action level for mercury in fish and shellfish is 1.0 mg/kg (Table 5), and now only applies to the methylmercury in consumed tissues (U.S. FDA, 1984a,b). In their test on methylmercury, McKim, et al. (1976) found that brook trout exposed to 0.03 $\mu\text{g/L}$ contained 1 mg/kg in muscle tissue. However, in their test on methylmercury with the fathead minnow, Olson, et al. (1975) found that exposure to 0.018 $\mu\text{g/L}$ resulted in 1.47 mg/kg in the fish and a BCF of 81,700. Use of this BCF with the FDA action level results in a Final Residue Value of 0.012 $\mu\text{g/L}$ for methylmercury (Table 5). The concentration in the fathead minnow is for whole body, but Heisinger, et al. (1979) found no significant difference between various body compartments. Further, Huckabee, et al. (1974) found that all fishes in a particular environment acquired about the same concentrations of mercury in both the whole body and muscle tissue when they were chronically exposed to low concentrations of mercury. On the other hand, Heiskary and Helwig (1983) and McKim, et al. (1976) found higher concentrations of mercury in the edible portion of fish than in the whole body. Thus the concentration of mercury in the muscle of some edible species is likely to exceed the FDA action level when exposed to methylmercury at a concentration of 0.012 $\mu\text{g/L}$.

Although the FDA action level only applies to methylmercury in fish and shellfish, it can be used to derive a water quality criterion for mercury(II) because most of the mercury in fish is methylmercury even if the organisms were exposed to inorganic mercury (de Freitas, et al. 1974; Jernelov and Lann, 1971). A BCF of 4,994 was obtained for mercuric chloride in a

life-cycle test with the fathead minnow (Snarski and Olson, 1982). This BCF is based on the concentration of mercuric chloride in the water and the total concentration of organic and inorganic mercury in the tissue. Even though all concentrations tested caused adverse effects and the higher concentrations caused more severe effects, the BCFs were similar at all concentrations and were lower than those obtained with methylmercury by McKim, et al. (1976) and Olson, et al. (1975). Use of the BCF of 4,994 with the FDA action level of 1.0 mg/kg results in a freshwater Final Residue Value of 0.20 $\mu\text{g/L}$ for mercury(II) (Table 5). This value of 0.20 $\mu\text{g/L}$ derived for mercury(II) would, however, be too low if field BCFs are higher than laboratory BCFs, if waters contain substantial concentrations of methylmercury, or if methylation processes are accelerated in sediment.

Information on the bioconcentration of various mercury compounds by saltwater animals and plants is included in Tables 5 and 6 and by saltwater plankton in Table 6. For mercuric chloride, BCFs ranged from 853 to 10,920 with algae. In tests with the eastern oyster, BCFs of 10,000, 40,000, and 40,000 were obtained for mercuric chloride, methylmercuric chloride and phenylmercuric chloride, respectively (Kopfler, 1974). These are similar to the BCFs obtained with freshwater fish, but the BCF of 129 obtained for mercuric chloride in tail muscle of the American lobster is much lower.

To protect the marketability of saltwater shellfish for human consumption, Final Residue Values can be calculated based on the BCFs for the oyster and the FDA action level of 1.0 mg/kg. Accordingly, the Final Residue Values for mercuric chloride, methylmercuric chloride, and phenylmercuric chloride are 0.10, 0.025, and 0.025 $\mu\text{g/L}$, respectively. However, at these concentrations fifty percent of the exposed oysters would probably exceed the

FDA action level if all the mercury in the body were present as methylmercury.

Other Data

Most of the significant freshwater and saltwater results in Table 6 have already been discussed in connection with data in Tables 1-5, but a few additional items deserve special mention. Comparable tests with four species showed that mercuric cyanide was 0.67 to 50 times as toxic as mercuric chloride. Also, Birge, et al. (1979) reported that flow-through tests gave EC50s nearly two orders of magnitude lower than static tests with rainbow trout, catfish, goldfish, and largemouth bass (Table 6). Bouquegneau (1979) found that preexposure induced metallothionein production, which then protected the fish.

Molybdenum (Yamane and Koizumi, 1982) and vitamin E (Ganther, 1978, 1980) affects the toxicity of mercury to mammals, and probably many consumers of aquatic organisms, as does selenium (e.g., Alexander, et al. 1979; Berlin, 1978; National Research Council, 1978; National Research Council Canada, 1979; Stopford and Goldwater, 1975; Strom, et al. 1979). Wobeser, et al. (1976a,b) found methylmercury to be much less toxic to mink when they were fed freshwater drum, Aplodinolus grunniens, containing high mercury tissue residues than when they were fed a diet to which methylmercury chloride had been added. On the other hand, Albanus, et al. (1972) and Charbonneau, et al. (1974) found similar toxicity to cats when fed similar dietary concentrations of methylmercury, one as a tissue residue in pike and the other with methylmercury added to the ration.

Finley and Stendell (1978) and Heinz (1979a) fed black and mallard ducks, respectively, food contaminated with methylmercuric dicyandiamide.

These feeding studies extended over two and three generations, respectively, and demonstrated reduced hatching success and juvenile survival at mercury concentrations that were estimated to be equivalent to 0.5 and 0.1 mg/kg, respectively, in the natural succulent food of the wild ducks. These results were not used to estimate a Final Residue Value based on food for wildlife because the dicyandiamide compound might not represent the toxicity of methylmercury alone. Nevertheless, these tests suggest that the Final Residue Value might be an order of magnitude too high because at least one of these authors believes that the anion had little effect on the results (Heinz, 1979b).

Unused Data

Some data on the effects of mercury on aquatic organisms were not used because the studies were conducted with species that are not resident in North America (e.g., Ahsanullah, 1982; Akiyama, 1970; Dial, 1978a,b; Heisinger and Green, 1975; Jones, 1939a, 1940, 1973, 1975; Khangarot, et al. 1982; Kihlstrom and Hulch, 1972; Krishnaja and Rege, 1982; Mathur, et al. 1981; McClurg, 1984; Murci and Shukla, 1984; Nagashima, et al. 1983; Saxena and Parashari, 1983; Shaffi, 1981; Srivastava, 1982; van den Broek and Tracey, 1981; Verma, et al. 1984), or because the test species was not obtained in North America and was not identified well enough to determine if it is resident in North America (Hannerz, 1968). Brown and Ahsanullah (1971) conducted tests with brine shrimp, which species is too atypical to be used in deriving national criteria. Reviews by Chapman, et al. (1968), Eisler (1981), Eisler, et al. (1979), Phillips and Russo (1978), and Thompson, et al. (1972) only contain data that have been published elsewhere.

The 96-hr values reported by Buikema, et al. (1974a,b) were subject to error because of possible reproductive interactions (Buikema, et al. 1977). Applegate, et al. (1957) exposed only one or two organisms. Data were not used if the mode of exposure was inappropriate for deriving water quality criteria (Giblin and Massaro, 1973; Lucu and Skreblin, 1981; Schmidt-Nielson, et al. 1977; Weisbart, 1973). In addition, data were not used if mercury was a component of an effluent (Wong, et al. 1982) or if the test organisms were cultured in one water and tested in another (Bringmann and Kuhn, 1982). Bills, et al. (1977) and Passino and Cocant (1979) did not report any usable results. Jones (1935, 1938, 1939b, 1947), Miller (1980, 1981), and Nuzzi (1972) did not report a clearly defined endpoint.

Results of some laboratory tests were not used because the test was conducted in distilled water (McKone, et al. 1971), the quality of the dilution water or medium was questionable (Brkovic-Popovic and Popovic, 1977a,b; Carter and Cameron, 1973; Kim, et al. 1977a,b; Stary and Kratzer, 1982; Stary, et al. 1982, 1983), or because the test solution or culture medium contained too much EDTA which would probably complex mercury (Gutierrez-Galindo, 1981; Knowles and Zingmark, 1978; Stratton and Corke, 1979; Stratton, et al. 1979).

Data concerning concentrations of mercury in wild organisms (e.g., Abernathy and Cumbie, 1977; Armstrong and Scott, 1979; Bodaly, et al. 1984; Copeland and Ayers, 1972; DiNardi, et al. 1974; Flegal, et al. 1981; Helwig and Hora, 1983; Hildebrand, et al. 1980; Jensen, et al. 1981; Leonzio, et al. 1982; Martin, et al. 1984; May and McKinney, 1981; Mitchell, et al. 1982; Moore and Sutherland, 1980; Murray, 1978; Pennington, et al. 1982; Phillips and Buhler, 1980; Price and Knight, 1978; Ray, et al. 1984; Sheffy, 1978; Tsui and McCart, 1981; Wachs, 1982; Watling, et al. 1981) were not used to

calculate bioaccumulation factors if the concentrations of mercury in the ambient water during the period of exposure was not adequately measured. Studies using isotopic mercury (e.g., Cunningham and Tripp, 1975; Glooschenko, 1969) were not used because of the possibility of isotopic discrimination.

Results of bioconcentration tests were not used if the tests were conducted in distilled water, were not long enough, were not flow-through, or if the concentration of mercury in the test solution was not adequately measured (e.g., Cunningham and Tripp, 1973; Kim, et al. 1977a,b; McKone, et al. 1971; Medeiros, et al. 1980; Middaugh and Rose, 1974; Phillips and Gregory, 1980; Ribeyre, et al. 1980; Sharma and Davis, 1980a; Stary and Kratzer, 1980, 1982; Stary, et al. 1980, 1981, 1982, 1983; Vernberg and O'Hara, 1972).

Summary

Data are available on the acute toxicity of mercury(II) to 28 genera of freshwater animals. Acute values for invertebrate species range from 2.2 µg/L for Daphnia pulex to 2,000 µg/L for three insects. Acute values for fishes range from 30 µg/L for the guppy to 1,000 µg/L for the Mozambique tilapia. Few data are available for various organomercury compounds and mercurous nitrate, and they all appear to be 4 to 31 times more acutely toxic than mercury(II).

Available chronic data indicate that methylmercury is the most chronically toxic of the tested mercury compounds. Tests on methylmercury with Daphnia magna and brook trout produced chronic values less than 0.07 µg/L. For mercury(II) the chronic value obtained with Daphnia magna was about 1.1 µg/L and the acute-chronic ratio was 4.5. In both a life-cycle

test and an early life-stage test on mercuric chloride with the fathead minnow, the chronic value was less than 0.26 $\mu\text{g}/\text{L}$ and the acute-chronic ratio was over 600.

Freshwater plants show a wide range of sensitivities to mercury, but the most sensitive plants appear to be less sensitive than the most sensitive freshwater animals to both mercury(II) and methylmercury. A bioconcentration factor of 4,994 is available for mercury(II), but the bioconcentration factors for methylmercury range from 4,000 to 85,000.

Data on the acute toxicity of mercuric chloride are available for 29 genera of saltwater animals including annelids, molluscs, crustaceans, echinoderms, and fishes. Acute values range from 3.5 $\mu\text{g}/\text{L}$ for a mysid to 1,678 $\mu\text{g}/\text{L}$ for winter flounder. Fishes tend to be more resistant and molluscs and crustaceans tend to be more sensitive to the acute toxic effects of mercury(II). Results of a life-cycle test with the mysid show that mercury(II) at a concentration of 1.6 $\mu\text{g}/\text{L}$ significantly affected time of first spawn and productivity; the resulting acute-chronic ratio was 3.1.

Concentrations of mercury that affected growth and photosynthetic activity of one saltwater diatom and six species of brown algae range from 10 to 160 $\mu\text{g}/\text{L}$. Bioconcentration factors of 10,000 and 40,000 have been obtained for mercuric chloride and methylmercury with an oyster.

National Criteria

Derivation of a water quality criterion for mercury is more complex than for most metals because of methylation of mercury in sediment, in fish, and in the food chain of fish. Apparently almost all mercury currently being discharged is mercury(II). Thus mercury(II) should be the only important

possible cause of acute toxicity and the Criterion Maximum Concentrations can be based on the acute values for mercury(II).

The best available data concerning long-term exposure of fish to mercury(II) indicates that concentrations above 0.23 $\mu\text{g/L}$ caused statistically significant effects on the fathead minnow and caused the concentration of total mercury in the whole body to exceed 1.0 mg/kg. Although it is not known what percent of the mercury in the fish was methylmercury, it is also not known whether uptake from food would increase the concentration in the fish in natural situations. Species such as rainbow trout, coho salmon, and especially the bluegill, might suffer chronic effects and accumulate high residues of mercury about the same as the fathead minnow.

With regard to long-term exposure to methylmercury, McKim, et al. (1976) found that brook trout can exceed the FDA action level without suffering statistically significant adverse effects on survival, growth, or reproduction. Thus for methylmercury the Final Residue Value would be substantially lower than the Final Chronic Value.

Basing a freshwater criterion on the Final Residue Value of 0.012 $\mu\text{g/L}$ derived from the bioconcentration factor of 81,700 for methylmercury with the fathead minnow (Olson, et al. 1975) essentially assumes that all discharged mercury is methylmercury. On the other hand, there is the possibility that in field situations uptake from food might add to the uptake from water. Similar considerations apply to the derivation of the saltwater criterion of 0.025 $\mu\text{g/L}$ using the BCF of 40,000 obtained for methylmercury with the Eastern oyster (Kopfler, 1974). Because the Final Residue Values for methylmercury are substantially below the Final Chronic Values for mercury(II), it is probably not too important that many fishes, including the rainbow trout,

coho salmon, bluegill, and haddock might not be adequately protected by the freshwater and saltwater Final Chronic Values for mercury(II).

In contrast to all the complexities of deriving numerical criteria for mercury, monitoring for unacceptable environmental effects should be relatively straightforward. The most sensitive adverse effect will probably be exceedence of the FDA action level. Therefore, existing discharges should be acceptable if the concentration of methylmercury in the edible portion of exposed consumed species does not exceed the FDA action level.

The procedures described in the "Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses" indicate that, except possibly where a locally important species is very sensitive, freshwater aquatic organisms and their uses should not be affected unacceptably if the four-day average concentration of mercury does not exceed 0.012 $\mu\text{g}/\text{L}$ more than once every three years on the average and if the one-hour average concentration does not exceed 2.4 $\mu\text{g}/\text{L}$ more than once every three years on the average. If the four-day average concentration exceeds 0.012 $\mu\text{g}/\text{L}$ more than once in a three-year period, the edible portion of consumed species should be analyzed to determine whether the concentration of methylmercury exceeds the FDA action level.

The procedures described in the "Guidelines for Deriving Numerical National Water Quality Criteria for the Protection of Aquatic Organisms and Their Uses" indicate that, except possibly where a locally important species is very sensitive, saltwater aquatic organisms and their uses should not be affected unacceptably if the four-day average concentration of mercury does not exceed 0.025 $\mu\text{g}/\text{L}$ more than once every three years on the average and if the one-hour average concentration does not exceed 2.1 $\mu\text{g}/\text{L}$ more than once every three years on the average. If the four-day average concentration

exceeds 0.025 µg/L more than once in a three-year period, the edible portion of consumed species should be analyzed to determine whether the concentration of methylmercury exceeds the FDA action level.

EPA believes that a measurement such as "acid-soluble" would provide a more scientifically correct basis upon which to establish criteria for metals. The criteria were developed on this basis. However, at this time, no EPA approved methods for such a measurement are available to implement the criteria through the regulatory programs of the Agency and the States. The Agency is considering development and approval of methods for a measurement such as "acid-soluble". Until available, however, EPA recommends applying the criteria using the total recoverable method. This has two impacts: (1) certain species of some metals cannot be analyzed directly because the total recoverable method does not distinguish between individual oxidation states, and (2) these criteria may be overly protective when based on the total recoverable method.

The recommended exceedence frequency of three years is the Agency's best scientific judgment of the average amount of time it will take an unstressed system to recover from a pollution event in which exposure to mercury exceeds the criterion. Stressed systems, for example, one in which several outfalls occur in a limited area, would be expected to require more time for recovery. The resilience of ecosystems and their ability to recover differ greatly, however, and site-specific criteria may be established if adequate justification is provided.

The use of criteria in designing waste treatment facilities requires the selection of an appropriate wasteload allocation model. Dynamic models are preferred for the application of these criteria. Limited data or other factors may make their use impractical, in which case one should rely on a

steady-state model. The Agency recommends the interim use of 1Q5 or 1Q10 for Criterion Maximum Concentration (CMC) design flow and 7Q5 or 7Q10 for the Criterion Continuous Concentration (CCC) design flow in steady-state models for unstressed and stressed systems respectively. These matters are discussed in more detail in the Technical Support Document for Water Quality-Based Toxics Control (U.S. EPA, 1985).

Table 1. Acute Toxicity of Mercury to Aquatic Animals

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value (µg/L)**</u>	<u>Reference</u>
<u>FRESHWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Tubificid worm, Branchiura sowerbyi</u>	R, U	Mercuric chloride	80	80	Chapman, et al. 1982a
<u>Tubificid worm, Limnodrilus hoffmeisteri</u>	R, U	Mercuric chloride	180	180	Chapman, et al. 1982a,b
<u>Tubificid worm, Quistadrilus multisetosus</u>	R, U	Mercuric chloride	250	250	Chapman, et al. 1982a
<u>Tubificid worm, Rhyacodrilus montana</u>	R, U	Mercuric chloride	240	240	Chapman, et al. 1982a
<u>Tubificid worm, Spirosperma ferox</u>	R, U	Mercuric chloride	330	330	Chapman, et al. 1982a
<u>Tubificid worm, Spirosperma nikolskyl</u>	R, U	Mercuric chloride	500	500	Chapman, et al. 1982a
<u>Tubificid worm, Stylodrilus heringianus</u>	R, U	Mercuric chloride	140	140	Chapman, et al. 1982a
<u>Tubificid worm, Tubifex tubifex</u>	R, U	Mercuric chloride	140	140	Chapman, et al. 1982a,b
<u>Tubificid worm, Varlchaeta pacifica</u>	R, U	Mercuric chloride	100	100	Chapman, et al. 1982a
<u>Worm, Nais sp.</u>	S, M	Mercuric nitrate	1,000	1,000	Rehwoidt, et al. 1973
<u>Snail (embryo), Amnicola sp.</u>	S, M	Mercuric nitrate	2,100***	-	Rehwoidt, et al. 1973
<u>Snail (adult), Amnicola sp.</u>	S, M	Mercuric nitrate	80	80	Rehwoidt, et al. 1973
<u>Snail, Aplexa hypnorum</u>	S, U	Mercuric chloride	370	370	Holcombe, et al. 1983

Table 1. (Continued)

<u>Species</u>	<u>Method</u> [#]	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)</u> ^{**}	<u>Species Mean Acute Value (µg/L)</u> ^{**}	<u>Reference</u>
<u>Cladoceran, Daphnia magna</u>	S, U	Mercuric chloride	<4.4****	-	Anderson, 1948
<u>Cladoceran, Daphnia magna</u>	S, U	Mercuric chloride	5	-	Blesinger & Christensen, 1972
<u>Cladoceran, Daphnia magna</u>	S, U	Mercuric chloride	3.177	-	Canton & Adema, 1978
<u>Cladoceran, Daphnia magna</u>	S, U	Mercuric chloride	1.478	-	Canton & Adema, 1978
<u>Cladoceran, Daphnia magna</u>	S, U	Mercuric chloride	2.180	-	Canton & Adema, 1978
<u>Cladoceran (<6 hr old), Daphnia magna</u>	S, U	Mercuric chloride	4.4	-	Barera & Adams, 1983
<u>Cladoceran (<24 hr old), Daphnia magna</u>	S, U	Mercuric chloride	4.4	-	Barera & Adams, 1983
<u>Cladoceran (1-9 day old), Daphnia magna</u>	S, U	Mercuric chloride	5.2-14.8***	3.157	Barera & Adams, 1983
<u>Cladoceran, Daphnia pulex</u>	S, U	Mercuric chloride	2.217	2.217	Canton & Adema, 1978
<u>Amphipod, Gammarus sp.</u>	S, M	Mercuric nitrate	10	10	Rehboldt, et al. 1973
<u>Crayfish (male, mixed ages), Faxonella clypeatus</u>	R, M	Mercuric chloride	20	20	Helt & Fingerman, 1977; Helt, 1981
<u>Crayfish, Orconectes ilmosus</u>	S, M	Mercuric chloride	50	50	Boutet & Chaisemartin, 1973
<u>Mayfly, Ephemera subvaria</u>	S, U	Mercuric chloride	2,000	2,000	Warnick & Bell, 1969
<u>Damselfly, (Unidentified)</u>	S, M	Mercuric nitrate	1,200	1,200	Rehboldt, et al. 1973

Table 1. (Continued)

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value (µg/L)**</u>	<u>Reference</u>
Stonefly, <u>Acroneuria lycorias</u>	S, U	Mercuric chloride	2,000	2,000	Warnick & Bell, 1969
Caddisfly, <u>Hydropsyche betteni</u>	S, U	Mercuric chloride	2,000	2,000	Warnick & Bell, 1969
Caddisfly, (Unidentified)	S, M	Mercuric nitrate	1,200	1,200	Rehboldt, et al. 1973
Midge, <u>Chironomus sp.</u>	S, M	Mercuric nitrate	20	20	Rehboldt, et al. 1973
Coho salmon (juvenile), <u>Oncorhynchus kisutch</u>	R, M	Mercuric chloride	240	240	Lorz, et al. 1978
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	R, U	Mercuric chloride	155.1	-	Matida, et al., 1971
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	FT, U	Mercuric chloride	280	-	MacLeod & Pessah, 1973
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	FT, U	Mercuric chloride	220	-	MacLeod & Pessah, 1973
Rainbow trout, <u>Salmo gairdneri</u>	FT, U	Mercuric chloride	420	-	Daoust, 1981
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	FT, M	Mercuric chloride	275	275	Lock & van Overbeeke, 1981
Fathead minnow, <u>Pimephales promelas</u>	FT, M	Mercuric chloride	168	-	Snarski & Olson, 1982
Fathead minnow, <u>Pimephales promelas</u>	FT, M	Mercuric chloride	150	158.7	Call, et al. 1983
Mosquitofish (female), <u>Gambusia affinis</u>	S, U	Mercuric chloride	180	180	Joshi & Rege, 1980
Guppy (116-157 mg), <u>Poecilia reticulata</u>	R, U	Mercuric chloride	30	-	Deshmukh & Marathe, 1980

Table 1. (Continued)

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value (µg/L)**</u>	<u>Reference</u>
<u>Guppy (362-621 mg), Poecilia reticulata</u>	R, U	Mercuric chloride	53.5***	30	Deshmukh & Marathe, 1980
<u>Bluegill (juvenile), Lepomis macrochirus</u>	S, U	Mercuric chloride	160	160	Holcombe, et al. 1983
<u>Mozambique tilapia, Tilapia mossambica</u>	S, U	Mercuric chloride	1,000	1,000	Qureshi & Saksena, 1980
<u>Methylmercury</u>					
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	R, U	Methylmercuric chloride	25	-	Matida, et al. 1971
<u>Rainbow trout (larva), Salmo gairdneri</u>	R, U	Methylmercuric chloride	24	-	Wobeser, 1973
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	R, U	Methylmercuric chloride	42	-	Wobeser, 1973
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	FT, M	Methylmercuric chloride	24	24	Lock & van Overbeke, 1981; Lock, et al. 1981
<u>Brook trout (juvenile), Salvelinus fontinalis</u>	FT, M	Methylmercuric chloride	84	-	McKim, et al. 1976
<u>Brook trout (yearling), Salvelinus fontinalis</u>	FT, M	Methylmercuric chloride	65	73.89	McKim, et al. 1976
<u>Other Mercury Compounds</u>					
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	R, U	Phenylmercuric acetate	5	5	Matida, et al. 1971
<u>Rainbow trout (2 mos), Salmo gairdneri</u>	FT, M	Mercurous nitrate	33.0	33.0	Hale, 1977
<u>Goldfish, Carassius auratus</u>	S, U	Phenylmercuric lactate	82	82	Ellis, 1947

Table 1. (Continued)

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value** (µg/L)</u>	<u>Reference</u>
<u>Common carp, Cyprinus carpio</u>	R, U	2-Methoxy ethyl mercuric chloride	139	139	Das & Misra, 1982
<u>Fathead minnow, Pimephales promelas</u>	S, M	Mercuric acetate	40	40	Curtis, et al. 1979; Curtis & Ward, 1981
<u>Fathead minnow, Pimephales promelas</u>	S, M	Mercuric thiocyanate	115	115	Curtis, et al. 1979; Curtis & Ward, 1981
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Ethylmercuric p-toluene sulfonamide	51	51	Clemens & Sneed, 1959
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Ethylmercuric phosphate	49	49	Clemens & Sneed, 1959
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Phenylmercuric acetate	1,966	1,966	Clemens & Sneed, 1959
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Phenylmercuric acetate	28	28	Clemens & Sneed, 1958a, 1959
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Pyridylmercuric acetate	<176	-	Clemens & Sneed, 1958b
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Pyridylmercuric acetate	224	-	Clemens & Sneed, 1958b
<u>Channel catfish (juvenile), Ictalurus punctatus</u>	S, U	Pyridylmercuric acetate	<153	<182	Clemens & Sneed, 1958b
<u>Mosquitofish (female), Gambusia affinis</u>	S, U	Methoxy ethyl mercuric chloride	910	910	Joshi & Rege, 1980
<u>Mosquitofish (female), Gambusia affinis</u>	S, U	Phenylmercuric acetate	37	37	Joshi & Rege, 1980
<u>Mosquitofish (female), Gambusia affinis</u>	S, U	Phenylmercuric acetate (Ceresan)	44	44	Joshi & Rege, 1980

Table 1. (Continued)

<u>Species</u>	<u>Method[#]</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value** (µg/L)</u>	<u>Reference</u>
<u>SALTWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Polychaete worm (adult), Neanthes arenaceodentata</u>	S, U	Mercuric chloride	96	-	Reish, et al. 1976
<u>Polychaete worm (juvenile), Neanthes arenaceodentata</u>	S, U	Mercuric chloride	100	97.98	Reish, et al. 1976
<u>Sand worm (adult), Nereis virens</u>	S, U	Mercuric chloride	70	70	Eisler & Hennekey, 1977
<u>Polychaete worm (larva), Capitella capitata</u>	S, U	Mercuric chloride	14	14	Reish, et al. 1976
<u>Oligochaete worm, Limnodriloides verrucosus</u>	R, U	Mercuric chloride	120	120	Chapman, et al. 1982a
<u>Oligochaete worm, Monopylephorus cuticulatus</u>	R, U	Mercuric chloride	230	230	Chapman, et al. 1982a
<u>Oligochaete worm, Tubificoides gabriellae</u>	R, U	Mercuric chloride	98	98	Chapman, et al. 1982a
<u>Northern horse mussel, Modiolus modiolus</u>	S, M	Mercuric chloride	230	230	Hilmy, et al. 1981
<u>Blue mussel, Mytilus edulis</u>	S, U	Mercuric chloride	5.8	5.8	Martin, et al. 1981
<u>Bay scallop (juvenile), Argopecten irradians</u>	S, U	Mercuric chloride	89	89	Nelson, et al. 1976
<u>Pacific oyster, Crassostrea gigas</u>	S, U	Mercuric chloride	6.7	-	Martin, et al. 1981
<u>Pacific oyster, Crassostrea gigas</u>	S, M	Mercuric chloride	5.7	-	Glickstein, 1978
<u>Pacific oyster, Crassostrea gigas</u>	S, M	Mercuric nitrate	5.5	5.944	Glickstein, 1978

Table 1. (Continued)

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value (µg/L)**</u>	<u>Reference</u>
<u>Eastern oyster, Crassostrea virginica</u>	S, U	Mercuric chloride	5.6	-	Calabrese & Nelson, 1974; Calabrese, et al. 1977
<u>Eastern oyster, Crassostrea virginica</u>	S, U	Mercuric chloride	10.2	7.558	MacInnes & Calabrese, 1978
<u>Common rangia (adult), Rangia cuneata</u>	S, U	Mercuric chloride	10,000	-	Olson & Harrel, 1973
<u>Common rangia (adult), Rangia cuneata</u>	S, U	Mercuric chloride	8,700	-	Olson & Harrel, 1973
<u>Common rangia (adult), Rangia cuneata</u>	S, M	Mercuric chloride	58	-	Dillon, 1977
<u>Common rangia (adult), Rangia cuneata</u>	S, M	Mercuric chloride	122	*****	Dillon, 1977
<u>Quahog clam, Mercenaria mercenaria</u>	S, U	Mercuric chloride	4.8	4.8	Calabrese & Nelson, 1974; Calabrese, et al. 1977
<u>Soft-shell clam (adult), Mya arenaria</u>	S, U	Mercuric chloride	400	400	Eisler & Hennekey, 1977
<u>Copepod, Pseudodiaptomus coronatus</u>	S, U	Mercuric chloride	79	79	Gentile, 1982
<u>Copepod, Eurytemora affinis</u>	S, U	Mercuric chloride	158	158	Gentile, 1982
<u>Copepod, Acartia clausi</u>	S, U	Mercuric chloride	10	10	Gentile, 1982
<u>Copepod (adult), Acartia tonsa</u>	S, U	Mercuric chloride	10	-	Sosnowski & Gentile, 1978
<u>Copepod (adult), Acartia tonsa</u>	S, U	Mercuric chloride	14	-	Sosnowski & Gentile, 1978
<u>Copepod (adult), Acartia tonsa</u>	S, U	Mercuric chloride	15	-	Sosnowski & Gentile, 1978

Table 1. (Continued)

<u>Species</u>	<u>Method</u> [#]	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)</u> ^{**}	<u>Species Mean Acute Value (µg/L)</u> ^{**}	<u>Reference</u>
Copepod (adult), <u>Acartia tonsa</u>	S, U	Mercuric chloride	20	14.32	Gentile, 1982
Copepod, <u>Nitocra spinipes</u>	S, U	Mercuric chloride	230	230	Bengtsson, 1978
Mysid, <u>Mysidopsis bahia</u>	FT, M	Mercuric chloride	3.5	3.5	Gentile, et al. 1982, 1983; Lussler, et al. Manuscript
White shrimp (adult), <u>Penaeus setiferus</u>	S, U	Mercuric chloride	17	17	Green, et al. 1976
American lobster (larva), <u>Homarus americanus</u>	S, U	Mercuric chloride	20	20	Johnson & Gentile, 1979
Hermit crab (adult), <u>Pagurus longicarpus</u>	S, U	Mercuric chloride	50	50	Eisler & Hennekey, 1977
Dungeness crab (larva), <u>Cancer magister</u>	S, U	Mercuric chloride	8.2	-	Martin, et al. 1981
Dungeness crab (larva), <u>Cancer magister</u>	S, M	Mercuric chloride	6.6	7.357	Glickstein, 1978
Green crab (larva), <u>Carcinus maenas</u>	S, U	Mercuric chloride	14	14	Connor, 1972
Starfish (adult), <u>Asterias forbesi</u>	S, U	Mercuric chloride	60	60	Eisler & Hennekey, 1977
Haddock (larva), <u>Melanogrammus aeglefinus</u>	S, U	Mercuric chloride	98	98	Cardin, 1982
Mummichog, <u>Fundulus heteroclitus</u>	S, U	Mercuric chloride	300	-	Dorfman, 1977
Mummichog, <u>Fundulus heteroclitus</u>	S, U	Mercuric chloride	200	-	Dorfman, 1977
Mummichog, <u>Fundulus heteroclitus</u>	S, U	Mercuric chloride	300	-	Dorfman, 1977

Table 1. (Continued)

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value (µg/L)**</u>	<u>Reference</u>
<u>Mummichog, Fundulus heteroclitus</u>	S, U	Mercuric chloride	300	-	Dorfman, 1977
<u>Mummichog (adult), Fundulus heteroclitus</u>	S, U	Mercuric chloride	800	-	Eisler & Hennekey, 1977
<u>Mummichog (adult), Fundulus heteroclitus</u>	S, U	Mercuric chloride	2,000	*****	Klaunig, et al. 1975
<u>Mummichog (embryo), Fundulus heteroclitus</u>	S, M	Mercuric chloride	67.4	67.4	Sharp & Neff, 1982
<u>Atlantic silverside (larva), Menidia menidia</u>	S, U	Mercuric chloride	144	-	Cardin, 1982
<u>Atlantic silverside (larva), Menidia menidia</u>	S, U	Mercuric chloride	125	-	Cardin, 1982
<u>Atlantic silverside (juvenile), Menidia menidia</u>	S, U	Mercuric chloride	86	115.7	Cardin, 1982
<u>Tidewater silverside (juvenile), Menidia peninsulae</u>	S, U	Mercuric chloride	71	71	Hansen, 1983
<u>Fourspine stickleback (adult), Apeltes quadracus</u>	S, U	Mercuric chloride	315	315	Cardin, 1982
<u>Spot (juvenile), Leiostomus xanthurus</u>	S, U	Mercuric chloride	36	36	Hansen, 1983
<u>Winter flounder (larva), Pseudopleuronectes americanus</u>	S, U	Mercuric chloride	1,820	-	Cardin, 1982
<u>Winter flounder (larva), Pseudopleuronectes americanus</u>	S, U	Mercuric chloride	1,560	-	Cardin, 1982

Table 1. (Continued)

<u>Species</u>	<u>Method*</u>	<u>Chemical</u>	<u>LC50 or EC50 (µg/L)**</u>	<u>Species Mean Acute Value (µg/L)**</u>	<u>Reference</u>
<u>Winter flounder (larva), Pseudopleuronectes americanus</u>	S, U	Mercuric chloride	1,810	-	Cardin, 1982
<u>Winter flounder (larva), Pseudopleuronectes americanus</u>	S, U	Mercuric chloride	1,320	-	Cardin, 1982
<u>Winter flounder (larva), Pseudopleuronectes americanus</u>	S, U	Mercuric chloride	1,960	1,678	Cardin, 1982
<u>Methylmercury</u>					
<u>Amphipod (adult), Gammarus duebeni</u>	S, U	Methylmercuric chloride	150	150	Lockwood & Inman, 1975
<u>Mummichog (embryo), Fundulus heteroclitus</u>	S, M	Methylmercuric chloride	51.1	51.1	Sharp & Neff, 1982
<u>Other Mercury Compounds</u>					
<u>Grass shrimp (adult), Palaemonetes pugio</u>	S, M	Mercuric acetate	47	47	Curtis, et al. 1979; Curtis & Ward, 1981
<u>Grass shrimp (adult), Palaemonetes pugio</u>	S, M	Mercuric thiocyanate	76	76	Curtis, et al. 1979; Curtis & Ward, 1981
<u>Mummichog, Fundulus heteroclitus</u>	S, U	Mercurous sulfate	6,800	-	Dorfman, 1977
<u>Mummichog, Fundulus heteroclitus</u>	S, U	Mercurous sulfate	300	*****	Dorfman, 1977

* S = static, R = renewal, FT = flow-through, U = unmeasured, M = measured.

** Results are expressed as mercury, not as the chemical.

*** Not used in calculation of Species Mean Acute Value because data are available for a more sensitive life stage.

**** "Less than" values were not used in calculations.

*****No Species Mean Acute Value calculated because acute values are too divergent for this species.

Table 2. Chronic Toxicity of Mercury to Aquatic Animals

<u>Species</u>	<u>Test*</u>	<u>Chemical</u>	<u>Limits (µg/L)**</u>	<u>Chronic Value (µg/L)**</u>	<u>Reference</u>
<u>FRESHWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Cladoceran, Daphnia magna</u>	LC***	Mercuric chloride	0.72-1.28	0.96	Biesinger, et al. 1982
<u>Cladoceran, Daphnia magna</u>	LC****	Mercuric chloride	0.91-1.82	1.287	Biesinger, et al. 1982
<u>Fathead minnow, Pimephales promelas</u>	LC	Mercuric chloride	<0.26*****	<0.26	Snarski & Olson, 1982
<u>Fathead minnow, Pimephales promelas</u>	ELS	Mercuric chloride	<0.23*****	<0.23	Call, et al. 1983
<u>Methylmercury</u>					
<u>Cladoceran, Daphnia magna</u>	LC***	Methylmercuric chloride	<0.04*****	<0.04	Biesinger, et al. 1982
<u>Cladoceran, Daphnia magna</u>	LC****	Methylmercuric chloride	0.52-0.87	0.6726	Biesinger, et al. 1982
<u>Brook trout, Salvelinus fontinalis</u>	LC	Methylmercuric chloride	0.29-0.93	0.5193	McKim, et al. 1976.
<u>Other Mercury Compounds</u>					
<u>Cladoceran, Daphnia magna</u>	LC****	Phenylmercuric acetate	1.12-1.90	1.459	Biesinger, et al. 1982
<u>SALTWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Mysid, Mysidopsis bahia</u>	LC	Mercuric chloride	0.8-1.6	1.131	Gentile, et al. 1982, 1983; Lussier, et al. Manuscript

Table 2. (Continued)

* LC = life cycle or partial life cycle, ELS = early life stage.

** Results are expressed as mercury, not as the chemical.

*** Flow-through

**** Renewal

***** Adverse effects occurred at this concentration, which was the lowest concentration tested.

<u>Species</u>	<u>Acute-Chronic Ratio</u>		
	<u>Acute Value</u> <u>(µg/L)</u>	<u>Chronic Value</u> <u>(µg/L)</u>	<u>Ratio</u>
	<u>Mercury(II)</u>		
<u>Cladoceran,</u> <u>Daphnia magna</u>	5	0.96	5.208
<u>Cladoceran,</u> <u>Daphnia magna</u>	5	1.287	3.885
<u>Fathead minnow,</u> <u>Pimephales promelas</u>	168	<0.26	>646.2
<u>Fathead minnow,</u> <u>Pimephales promelas</u>	150	<0.23	>652.2
<u>Mysid,</u> <u>Mysidopsis bahia</u>	3.5	1.131	3.095
	<u>Methylmercury</u>		
<u>Brook trout</u> <u>Salvelinus fontinalis</u>	73.89 [†]	0.5193	142.3

[†] Geometric mean of 2 values from McKim, et al. (1976) in Table 1.

Table 3. Ranked Genus Mean Acute Values with Species Mean Acute-Chronic Ratios

<u>Rank[#]</u>	<u>Genus Mean Acute Value (µg/L)</u>	<u>Species</u>	<u>Species Mean Acute Value (µg/L)</u>	<u>Species Mean Acute-Chronic Ratio</u>
<u>FRESHWATER SPECIES</u>				
<u>Mercury(II)</u>				
28	2,000	Stonefly, <u>Acroneuria lycorias</u>	2,000	-
27	2,000	Mayfly, <u>Ephemera subvaria</u>	2,000	-
26	2,000	Caddisfly, <u>Hydropsyche betteni</u>	2,000	-
25	1,200	Caddisfly, (Unidentified)	1,200	-
24	1,200	Damselfly, (Unidentified)	1,200	-
23	1,000	Worm, <u>Nais sp.</u>	1,000	-
22	1,000	Mozambique tilapia, <u>Tilapia mossambica</u>	1,000	-
21	406.2	Tubificid worm, <u>Spirosperma ferox</u>	330	-
		Tubificid worm, <u>Spirosperma nikolskyi</u>	500	-
20	370	Snail, <u>Aplexa hypnorum</u>	370	-
19	275	Rainbow trout, <u>Salmo gairdneri</u>	275	-
18	250	Tubificid worm, <u>Quistadrilus multisetosus</u>	250	-
17	240	Tubificid worm, <u>Rhyacodrilus montana</u>	240	-

Table 3. (Continued)

<u>Rank*</u>	<u>Genus Mean Acute Value (µg/L)</u>	<u>Species</u>	<u>Species Mean Acute Value (µg/L)</u>	<u>Species Mean Acute-Chronic Ratio</u>
16	240	Coho salmon, <u>Oncorhynchus kisutch</u>	240	-
15	180	Tubificid worm, <u>Limnodrilus hoffmeisteri</u>	180	-
14	180	Mosquitofish, <u>Gambusia affinis</u>	180	-
13	160	Bluegill, <u>Lepomis macrochirus</u>	160	-
12	158.7	Fathead minnow, <u>Pimephales promelas</u>	158.7	>649.2**
11	140	Tubificid worm, <u>Tubifex tubifex</u>	140	-
10	140	Tubificid worm, <u>Stylodrilus heringianus</u>	140	-
9	100	Tubificid worm, <u>Varichaeta pacifica</u>	100	-
8	80	Tubificid worm, <u>Branchiura sowerbyi</u>	80	-
7	80	Snail, <u>Amnicola sp.</u>	80	-
6	50	Crayfish, <u>Orconectes limosus</u>	50	-
5	30	Guppy, <u>Poecilia reticulata</u>	30	-
4	20	Crayfish, <u>Faxonella clypeatus</u>	20	-
3	20	Midge, <u>Chironomus sp.</u>	20	-

Table 3. (Continued)

<u>Rank[#]</u>	<u>Genus Mean Acute Value (µg/L)</u>	<u>Species</u>	<u>Species Mean Acute Value (µg/L)</u>	<u>Species Mean Acute-Chronic Ratio</u>
2	10	Amphipod, <u>Gammarus</u> sp.	10	-
1	2.646	Cladoceran, <u>Daphnia magna</u>	3.157	4.498**
		Cladoceran, <u>Daphnia pulex</u>	2.217	-
<u>SALTWATER SPECIES</u>				
<u>Mercury(II)</u>				
29	1,678	Winter flounder, <u>Pseudopleuronectes americanus</u>	1,678	-
28	400	Soft-shell clam, <u>Mya arenaria</u>	400	-
27	315	Fourspine stickleback, <u>Apeltes quadracus</u>	315	-
26	230	Northern horse mussel, <u>Modiolus modiolus</u>	230	-
25	230	Copepod, <u>Nitocra spinipes</u>	230	-
24	230	Oligochaete worm, <u>Monopylephorus cuticulatus</u>	230	-
23	158	Copepod, <u>Eurytemora affinis</u>	158	-
22	120	Oligochaete worm, <u>Limnodriloides verrucosus</u>	120	-

Table 3. (Continued)

<u>Rank*</u>	<u>Genus Mean Acute Value (µg/L)</u>	<u>Species</u>	<u>Species Mean Acute Value (µg/L)</u>	<u>Species Mean Acute-Chronic Ratio</u>
21	98	Oligochaete worm, <u>Tubificoides gabriellae</u>	98	-
20	98	Haddock, <u>Melanogrammus aeglefinus</u>	98	-
19	97.98	Polychaete worm, <u>Neanthes arenaceodentata</u>	97.98	-
18	90.63	Atlantic silverside, <u>Menidia menidia</u>	115.7	-
		Tidewater silverside, <u>Menidia peninsulae</u>	71	-
17	89	Bay scallop, <u>Argopecten irradians</u>	89	-
16	79	Copepod, <u>Pseudodiaptomus coronatus</u>	79	-
15	70	Sand worm, <u>Nereis virens</u>	70	-
14	67.4	Mummichog, <u>Fundulus heteroclitus</u>	67.4	-
13	60	Starfish, <u>Arterias forbesi</u>	60	-
12	50	Hermit crab, <u>Pagurus longicarpus</u>	50	-
11	36	Spot, <u>Leiostomus xanthurus</u>	36	-
10	20	American lobster, <u>Homarus americanus</u>	20	-
9	17	White shrimp, <u>Penaeus setiferus</u>	17	-

Table 3. (Continued)

<u>Rank*</u>	<u>Genus Mean Acute Value (µg/L)</u>	<u>Species</u>	<u>Species Mean Acute Value (µg/L)</u>	<u>Species Mean Acute-Chronic Ratio</u>
8	14	Green crab, <u>Carcinus maenas</u>	14	-
7	14	Polychaete worm, <u>Capitella capitata</u>	14	-
6	11.97	Copepod, <u>Acartia clausi</u>	10	-
		Copepod, <u>Acartia tonsa</u>	14.32	
5	7.357	Dungeness crab, <u>Cancer magister</u>	7.357	-
4	6.703	Pacific oyster, <u>Crassostrea gigas</u>	5.944	-
		Eastern oyster, <u>Crassostrea virginica</u>	7.558	-
3	5.8	Blue mussel, <u>Mytilus edulis</u>	5.8	-
2	4.8	Quahog clam, <u>Mercenaria mercenaria</u>	4.8	-
1	3.5	Mysid, <u>Mysidopsis bahia</u>	3.5	3.095

* Ranked from most resistant to most sensitive based on Genus Mean Acute Value.

**Geometric mean of two values in Table 2.

Table 3. (Continued)

Mercury(II)

Final Acute-Chronic Ratio = 3.731 (see text)

Fresh water

Final Acute Value = 4.857 $\mu\text{g/L}$

Criterion maximum concentration = $(4.857 \mu\text{g/L}) / 2 = 2.428 \mu\text{g/L}$

Final Chronic Value = $(4.857 \mu\text{g/L}) / 3.731 = 1.302 \mu\text{g/L}$

Salt water

Final Acute Value = 4.125 $\mu\text{g/L}$

Criterion maximum concentration = $(4.125 \mu\text{g/L}) / 2 = 2.062 \mu\text{g/L}$

Final Chronic Value = $(4.125 \mu\text{g/L}) / 3.731 = 1.106 \mu\text{g/L}$

Table 4. Toxicity of Mercury to Aquatic Plants

<u>Species</u>	<u>Chemical</u>	<u>Effect</u>	<u>Result</u> ($\mu\text{g/L}$) [#]	<u>Reference</u>
<u>FRESHWATER SPECIES</u>				
<u>Mercury(II)</u>				
Alga, <u>Chlorella vulgaris</u>	Mercuric chloride	33-day EC50 (cell division inhibition).	1,030	Rosko & Rachlin, 1977
Alga, <u>Chlorella vulgaris</u>	Mercuric chloride	LC50	100-1,000	Gipps & Biro, 1978
Alga, <u>Chlorella vulgaris</u>	Mercuric chloride	LC50	148-296	Rai, 1979; Rai, et al. 1981
Alga, <u>Chlorella vulgaris</u>	Mercuric chloride	15-day EC50 (growth)	443-592	Rai, et al. 1981
Alga, <u>Anabaena flos-aquae</u>	Mercuric chloride	EC50 (growth)	53	Thomas & Montes, 1978
Blue alga, <u>Microcystis aeruginosa</u>	Mercuric chloride	8-day Incipient inhibition	5	Bringmann, 1975; Bringmann & Kuhn, 1976, 1978a,b
Green alga, <u>Scenedesmus quadricauda</u>	Mercuric chloride	8-day Incipient inhibition	70	Bringmann, 1975; Bringmann & Kuhn, 1976, 1978a,b, 1979, 1980b
Alga, <u>Selenastrum capricornutum</u>	Mercuric chloride	Inhibited growth	59	Slooff, et al. 1983
Eurasian watermilfoil, <u>Myriophyllum spicatum</u>	Mercuric chloride	32-day EC50 (root weight)	3,400	Stanley, 1974
<u>Methylmercury</u>				
Alga, <u>Anabaena flos-aquae</u>	Methylmercuric chloride	EC50 (growth)	6.0	Thomas & Montes, 1978
Alga, <u>Chlorella vulgaris</u>	Methylmercuric chloride	15-day EC50 (growth)	0.8-4.0	Rai, et al. 1981

Table 4. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>Other Mercury Compounds</u>				
<u>Alga,</u> <u>Anabaena flos-aquae</u>	Phenylmercuric acetate	EC50 (growth)	2.8	Thomas & Montes, 1978
<u>SALTWATER SPECIES</u>				
<u>Mercury(II)</u>				
<u>Alga,</u> <u>Thalassiosira aestivalis</u>	Mercuric chloride	Reduced chlorophyll a	10	Hollibaugh, et al. 1980
<u>Seaweed,</u> <u>Ascophyllum nodosum</u>	Mercuric chloride	10-day EC50 (growth)	100	Strömgren, 1980
<u>Diatom,</u> <u>Ditylum brightwellii</u>	Mercuric chloride	5-day EC50 (growth)	10	Canterford & Canterford, 1980
<u>Seaweed,</u> <u>Fucus serratus</u>	Mercuric chloride	10-day EC50 (growth)	160	Strömgren, 1980
<u>Seaweed,</u> <u>Fucus spiralis</u>	Mercuric chloride	10-day EC50 (growth)	80	Strömgren, 1980
<u>Seaweed,</u> <u>Fucus vesiculosus</u>	Mercuric chloride	10-day EC50 (growth)	45	Strömgren, 1980
<u>Giant kelp,</u> <u>Macrocystis pyrifera</u>	Mercuric chloride	4-day EC50 (growth)	50	Clendenning & North, 1959
<u>Seaweed,</u> <u>Pelvetia canaliculata</u>	Mercuric chloride	10-day EC50 (growth)	130	Strömgren, 1980

* Results are expressed as mercury, not as the chemical.

Table 5. Bioaccumulation of Mercury by Aquatic Organisms

<u>Species</u>	<u>Tissue</u>	<u>Chemical</u>	<u>Duration (days)</u>	<u>Bioconcentration Factor[#]</u>	<u>Reference</u>
<u>FRESHWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Rainbow trout, <i>Salmo gairdneri</i></u>	Whole body	Mercuric chloride	60	1,800	Boudou & Ribeyre, 1984
<u>Fathead minnow, <i>Pimephales promelas</i></u>	Whole body	Mercuric chloride	287	4,994**	Snarski & Olson, 1982
<u>Methylmercury</u>					
<u>Rainbow trout, <i>Salmo gairdneri</i></u>	Whole body	Methylmercuric chloride	60	11,000	Boudou & Ribeyre, 1984
<u>Rainbow trout, <i>Salmo gairdneri</i></u>	Whole body	Methylmercuric chloride	75	85,700	Nimi & Lowe-Jinde, 1984
<u>Brook trout, <i>Salvelinus fontinalis</i></u>	Muscle	Methylmercuric chloride	273	11,000-33,000	McKim, et al. 1976
<u>Brook trout, <i>Salvelinus fontinalis</i></u>	Whole body	Methylmercuric chloride	273	10,000-23,000	McKim, et al. 1976
<u>Brook trout, <i>Salvelinus fontinalis</i></u>	Muscle and whole body	Methylmercuric chloride	756	12,000	McKim, et al. 1976
<u>Fathead minnow, <i>Pimephales promelas</i></u>	Whole body	Methylmercuric chloride	336	44,130-81,670	Olson, et al. 1975
<u>SALTWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Eastern oyster (adult), <i>Crassostrea virginica</i></u>	Soft parts	Mercuric chloride	74	10,000	Kopfler, 1974
<u>American lobster (adult), <i>Homarus americanus</i></u>	Tail muscle	Mercuric chloride	30	129	Thurberg, et al. 1977

Table 5. (Continued)

<u>Species</u>	<u>Tissue</u>	<u>Chemical</u>	<u>Duration (days)</u>	<u>Bioconcentration Factor[#]</u>	<u>Reference</u>
<u>Methylmercury</u>					
Eastern oyster (adult), <u>Crassostrea virginica</u>	Soft parts	Methylmercuric chloride	74	40,000	Kopfler, 1974
<u>Other Mercury Compounds</u>					
Eastern oyster (adult), <u>Crassostrea virginica</u>	Soft parts	Phenylmercuric chloride	74	40,000	Kopfler, 1974

* Results are based on mercury, not the chemical.

**From concentrations that caused adverse effects in a life-cycle test.

Maximum Permissible Tissue Concentration

<u>Consumer</u>	<u>Action Level or Effect</u>	<u>Concentration (mg/kg)</u>	<u>Reference</u>
Man	Action level for edible fish or shellfish	1.0	U.S. FDA, 1984a,b
Mink, <u>Mustela vison</u>	Histological evidence of injury	<1.1	Wobeser, 1976a,b
Brook trout, <u>Salvelinus fontinalis</u>	Death (700 days)	5-7	McKim, et al. 1976

Methylmercury

Freshwater Final Residue Value = (1.0 mg/kg) / 81,700 = 0.000012 mg/kg = 0.012 µg/L (see text)

Saltwater Final Residue Value = (1.0 mg/kg) / 40,000 = 0.000025 mg/kg = 0.025 µg/L

Mercury(II)

Freshwater Final Residue Value = (1.0 mg/kg) / 4,994 = 0.00020 mg/kg = 0.20 µg/L (see text)

Saltwater Final Residue Value = (1.0 mg/kg) / 10,000 = 0.00010 mg/kg = 0.10 µg/L (see text)

Table 6. Other Data on Effects of Mercury on Aquatic Organisms

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>FRESHWATER SPECIES</u>					
<u>Mercury(II)</u>					
Alga, (Spring assemblages, predominately diatoms)	Mercuric chloride	2 hrs	EC50 (reduced photosynthesis)	80	Blinn, et al. 1977
Alga, <u>Ankistrodesmus braunii</u>	Mercuric chloride	168-240 hrs	EC50 (inhibited lipid biosynthesis)	2,590	Matson, et al. 1972
Alga, <u>Ankistrodesmus braunii</u>	Mercuric chloride	24 days	Inhibited growth	74	Trevors, 1982
Alga, <u>Ankistrodesmus sp.</u>	Mercuric chloride	10 days	More toxic at pH = 5 than pH = 7	5	Baker, et al. 1983
Alga, <u>Synedra ulna</u>	Mercuric chloride	0.29 days	BCF=29,000	-	Fujita & Hashizume, 1972
Green alga, <u>Scenedesmus quadricauda</u>	Mercuric chloride	96 hrs	Incipient inhibition	30**	Bringmann & Kuhn, 1959a,b
Green alga, <u>Scenedesmus quadricauda</u>	Mercuric cyanide	96 hrs	Incipient inhibition	150**	Bringmann & Kuhn, 1959a,b
Bacteria, <u>Escherichia coli</u>	Mercuric chloride	-	Incipient inhibition	200	Bringmann & Kuhn, 1959a
Bacteria, <u>Escherichia coli</u>	Mercuric cyanide	-	Incipient inhibition	200	Bringmann & Kuhn, 1959a
Bacteria, <u>Pseudomonas putida</u>	Mercuric chloride	16 hrs	Incipient inhibition	10	Bringmann & Kuhn, 1976, 1977a, 1979, 1980b
Protozoan, <u>Entosiphon sulcatum</u>	Mercuric chloride	72 hrs	Incipient inhibition	18	Bringmann, 1978; Bringmann & Kuhn, 1979, 1980b, 1981
Protozoan, <u>Chilomonas paramecium</u>	Mercuric chloride	48 hrs	Incipient inhibition	15	Bringmann, et al. 1980; Bringmann & Kuhn, 1981
Protozoan, <u>Uronema parduczi</u>	Mercuric chloride	20 hrs	Incipient inhibition	67	Bringmann & Kuhn, 1980a, 1981

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)^a</u>	<u>Reference</u>
Protozoan, <u>Microregma heterostoma</u>	Mercuric chloride	28 hrs	Incipient inhibition	150	Bringmann & Kuhn, 1959b
Protozoan, <u>Microregma heterostoma</u>	Mercuric cyanide	28 hrs	Incipient inhibition	160	Bringmann & Kuhn, 1959b
Hydra, <u>Hydra oligactis</u>	Mercuric chloride	48 hrs	LC50	56	Slooff, 1983; Slooff, et al. 1983
Planarian, <u>Dugesia lugubris</u>	Mercuric chloride	48 hrs	LC50	55	Slooff, 1983
Tubificid worm, <u>Tubifex tubifex</u>	Mercuric chloride	48 hrs	LC50	3,200	Qureshi, et al. 1980
Snail, <u>Lymnaea stagnalis</u>	Mercuric chloride	48 hrs	LC50	443	Slooff, 1983; Slooff, et al. 1983
Mussel, <u>Margaritifera margaritifera</u>	Mercuric nitrate	39 days	BCF=302	-	Møllinger, 1973
Cladoceran, <u>Diaphanosoma sp.</u>	Mercuric chloride	3 wks	Reduced population density	2.8	Marshall, et al. 1981
Cladoceran, <u>Daphnia galeata mendotae</u>	Mercuric chloride	3 wks	Reduced population density	2.2	Marshall, et al. 1981
Cladoceran, <u>Daphnia magna</u>	Mercuric chloride	3 wks	Reproductive impairment	3.4	Blesinger & Christensen, 1972
Cladoceran, <u>Daphnia magna</u>	Mercuric chloride	48 hrs	EC50	30**	Bringmann & Kuhn, 1959a,b
Cladoceran, <u>Daphnia magna</u>	Mercuric cyanide	48 hrs	EC50	20**	Bringmann & Kuhn, 1959a,b
Cladoceran, <u>Daphnia magna</u>	Mercuric chloride	24 hrs	LC50	13	Bringmann & Kuhn, 1977b
Cladoceran, <u>Bosmina longirostris</u>	Mercuric chloride	3 wks	Reduced population density	2.8	Marshall, et al. 1981

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
Natural copepod assemblages	Mercuric chloride	7 days	Reduced growth rate	28.3	Borgmann, 1980
Amphipod, <u>Gammarus sp.</u>	Mercuric chloride	7 days	BCF=2,500	-	Zubarik & O'Connor, 1978
Amphipod, <u>Gammarus sp.</u>	Mercuric nitrate	7 days	BCF=2,500	-	Zubarik & O'Connor, 1978
Crayfish (male, mixed ages), <u>Faxonella clypeatus</u>	Mercuric chloride	72 hrs	LC50	200	Helt & Fingerman, 1977
Crayfish (0.2 g), <u>Faxonella clypeatus</u>	Mercuric chloride	24 hrs	LC50	1,000	Helt & Fingerman, 1977
Crayfish (1.2 g), <u>Faxonella clypeatus</u>	Mercuric chloride	672 hrs	LC50	1,000	Helt & Fingerman, 1977
Crayfish (adult), <u>Orconectes ilmosus</u>	Mercuric chloride	96 hrs	LC60	740	Doyle, et al. 1976
Crayfish (juvenile), <u>Orconectes ilmosus</u>	Mercuric chloride	30 days	LC50 (unfed)	2	Boutet & Chalsemartin, 1973
Crayfish (juvenile), <u>Orconectes ilmosus</u>	Mercuric chloride	30 days	LC50 (fed)	<2	Boutet & Chalsemartin, 1973
Crayfish (male, mixed ages), <u>Procambarus clarki</u>	Mercuric chloride	72 hrs	LC50	200	Helt & Fingerman, 1977
Freshwater community (primary producers, herbivores and carnivorous midges)	Mercuric chloride	1 yr	Reduced algal standing stock and diversity; no evidence of effects on midges	0.1	Sigmon, et al. 1977
Mosquito, <u>Aedes aegypti</u>	Mercuric chloride	48 hrs	LC50	4,100	Slooff, et al. 1983
Mosquito, <u>Aedes aegypti</u>	Mercuric chloride	48 hrs	LC50	776	Slooff, et al. 1983

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)^a</u>	<u>Reference</u>
<u>Pink salmon (embryo), <i>Oncorhynchus gorboscha</i></u>	Mercuric sulfate	2 days <time from fertiliza- tion to hatch	EC32 to EC81 (deformity)	5.2	Servizi & Martens, 1978
<u>Pink salmon (pre-eyed embryo), <i>Oncorhynchus gorboscha</i></u>	Mercuric sulfate	2 days <time from fertiliza- tion to stage	LC100	8.5	Servizi & Martens, 1978
<u>Pink salmon (larva), <i>Oncorhynchus gorboscha</i></u>	Mercuric sulfate	168 hrs	LC50	140	Servizi & Martens, 1978
<u>Sockeye salmon (embryo), <i>Oncorhynchus nerka</i></u>	Mercuric sulfate	2 days <time from fertiliza- tion to hatch	EC45.6 (deformity)	4.3	Servizi & Martens, 1978
<u>Sockeye salmon (pre-eyed embryo), <i>Oncorhynchus nerka</i></u>	Mercuric sulfate	2 days <time from fertiliza- tion to stage	LC100	9.3	Servizi & Martens, 1978
<u>Sockeye salmon (larva), <i>Oncorhynchus nerka</i></u>	Mercuric sulfate	168 hrs	LC50	290	Servizi & Martens, 1978
<u>Sockeye salmon (juvenile), <i>Oncorhynchus nerka</i></u>	Mercuric sulfate	168 hrs	LC50	190	Servizi & Martens, 1978
<u>Rainbow trout (juvenile), <i>Salmo gairdneri</i></u>	Mercuric chloride	24 hrs	LC50	903	Wobeser, 1973
<u>Rainbow trout (juvenile), <i>Salmo gairdneri</i></u>	Mercuric chloride	2 hrs	Depressed olfactory bulbar response	74	Hara, et al. 1976
<u>Rainbow trout (juvenile), <i>Salmo gairdneri</i></u>	Mercuric chloride	-	Growth inhibition	2.1-21	Matida, et al. 1971
<u>Rainbow trout, <i>Salmo gairdneri</i></u>	Mercuric chloride	1 wk	Effected osmo- regulation	100	Lock, et al. 1981
<u>Rainbow trout, <i>Salmo gairdneri</i></u>	Mercuric chloride	80 min	Avoidance threshold	0.2	Black & Birge, 1980
<u>Rainbow trout (embryo, larva), <i>Salmo gairdneri</i></u>	Mercuric chloride	28 days	EC50 (death and deformity)	4.7 5.0	Birge, et al. 1979, 1980

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>Rainbow trout (embryo, larva), Salmo gairdneri</u>	Mercuric chloride	28 days	EC50 (death and deformity)	<0.1	Birge, et al. 1979, 1980
<u>Rainbow trout (embryo, larva), Salmo gairdneri</u>	Mercuric chloride	28 days	EC10 (death and deformity)	0.9	Birge, et al. 1981
<u>Rainbow trout, Salmo gairdneri</u>	Mercuric chloride	5, 18 mo	Substantial mortality	0.12-0.24	Birge, et al. 1979
<u>Rainbow trout, Salmo gairdneri</u>	Mercuric chloride	48 hrs	LC50	480	Slooff, et al. 1983
<u>Goldfish (embryo, larva), Carassius auratus</u>	Mercuric chloride	7 days	EC50 (death and deformity)	121.9	Birge, 1978; Birge, et al. 1979
<u>Goldfish (embryo, larva), Carassius auratus</u>	Mercuric chloride	7 days	EC50 (death and deformity)	0.7	Birge, 1978; Birge, et al. 1979
<u>Common carp, Cyprinus carpio</u>	Mercuric chloride	60-72 hrs	Reduced hatching success	≥3,000	Huckabee & Griffith, 1974
<u>Fathead minnow, Pimephales promelas</u>	Mercuric chloride	48 hrs	LC50	37	Slooff, et al. 1983
<u>White sucker (adult), Catostomus commersoni</u>	Mercuric chloride	36 min	Blood enzyme (LDH) inhibition 20%	8,000	Christensen, 1972
<u>White sucker (adult), Catostomus commersoni</u>	Mercuric chloride	46 min	Blood enzyme (GOT) inhibition 20%	10,000	Christensen, 1972
<u>Channel catfish (embryo, larva), Ictalurus punctatus</u>	Mercuric chloride	10 days	EC50 (death and deformity)	30	Birge, et al. 1979
<u>Channel catfish (embryo, larva), Ictalurus punctatus</u>	Mercuric chloride	10 days	EC50 (death and deformity)	0.3	Birge, et al. 1979
<u>Channel catfish (embryo, larva), Ictalurus punctatus</u>	Mercuric chloride	-	Increased albinism	0.5	Westerman & Birge, 1978
<u>Channel catfish (embryo, larva), Ictalurus punctatus</u>	Mercuric chloride	10 days	BCF=441-2071	-	Birge, et al. 1979

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
Channel catfish (embryo, larva), <u>Ictalurus punctatus</u>	Mercuric chloride	10 days	BCF-4.4-353	-	Birge, et al. 1979
Mosquitofish, <u>Gambusia affinis</u>	Mercuric chloride	>10 days	LC50	500	Boudou, et al. 1979
Guppy, <u>Poecilia reticulata</u>	Mercuric chloride	24 hrs	LC50	13	Handy, 1977
Guppy, <u>Poecilia reticulata</u>	Mercuric chloride	48 hrs	LC50	303	Slooff, et al. 1983
Bluegill (embryo, larva), <u>Lepomis macrochirus</u>	Mercuric chloride	7-8 days	EC50 (death and deformity)	88.7	Birge, et al. 1979
Redear sunfish (embryo, larva), <u>Lepomis microlophus</u>	Mercuric chloride	7-8 days	EC50 (death and deformity)	137.2	Birge, et al. 1979
Largemouth bass (embryo, larva), <u>Micropterus salmoides</u>	Mercuric chloride	8 days	EC50 (death and deformity)	130 140	Birge, et al. 1978, 1979
Largemouth bass (embryo, larva), <u>Micropterus salmoides</u>	Mercuric chloride	8 days	EC50 (death and deformity)	5.3	Birge, et al. 1979
Largemouth bass, <u>Micropterus salmoides</u>	-	24 hrs	Affected opercular rhythm	10	Morgan, 1979
Mozambique tilapia, <u>Tilapia mossambica</u>	Mercuric nitrate	35 days	Clinical symptoms	310	Panigrahi & Misra, 1980
Mozambique tilapia, <u>Tilapia mossambica</u>	Mercuric chloride	48 hrs	LC50	1,000	Menezes & Qasim, 1983
Pig frog (embryo, larva), <u>Rana gryllo</u>	Mercuric chloride	7 days	EC50 (death and deformity)	67.2	Birge, et al. 1979
River frog (embryo, larva), <u>Rana heckscheri</u>	Mercuric chloride	7 days	EC50 (death and deformity)	59.9	Birge, et al. 1979
Leopard frog (embryo, larva), <u>Rana pipiens</u>	Mercuric chloride	7 days	EC50 (death and deformity)	7.3	Birge, et al. 1979

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
Narrow-mouthed toad (embryo, larva), <u>Gastrophryne carolinensis</u>	Mercuric chloride	7 days	EC50 (death and deformity)	1 1.3	Birge, et al. 1978, 1979
Green toad (embryo, larva), <u>Bufo debilis</u>	Mercuric chloride	7 days	EC50 (death and deformity)	40.0	Birge, et al. 1979
Fowler's toad (embryo, larva), <u>Bufo fowleri</u>	Mercuric chloride	7 days	EC50 (death and deformity)	65.9	Birge, et al. 1979
Red-spotted toad (embryo, larva), <u>Bufo punctatus</u>	Mercuric chloride	7 days	EC50 (death and deformity)	36.8	Birge, et al. 1979
Northern cricket frog (embryo, larva), <u>Acris crepitans</u>	Mercuric chloride	7 days	EC50 (death and deformity)	10.4	Birge, et al. 1979
Southern gray treefrog (embryo, larva), <u>Hyla chrysoscelis</u>	Mercuric chloride	7 days	EC50 (death and deformity)	2.4	Birge, et al. 1979
Spring peeper (embryo, larva), <u>Hyla crucifer</u>	Mercuric chloride	7 days	EC50 (death and deformity)	2.8	Birge, et al. 1979
Barking treefrog (embryo, larva), <u>Hyla gratiosa</u>	Mercuric chloride	7 days	EC50 (death and deformity)	2.5	Birge, et al. 1979
Squirrel treefrog (embryo, larva), <u>Hyla squirella</u>	Mercuric chloride	7 days	EC50 (death and deformity)	2.4	Birge, et al. 1979
Gray treefrog (embryo, larva), <u>Hyla versicolor</u>	Mercuric chloride	7 days	EC50 (death and deformity)	2.6	Birge, et al. 1979
African clawed frog, <u>Xenopus laevis</u>	Mercuric chloride	11 mos	Substantial mortality	0.16-0.2	Birge, et al. 1978
African clawed frog, <u>Xenopus laevis</u>	Mercuric chloride	48 hrs	LC50	74	Slooff & Baerseman, 1980; Slooff, et al. 1983

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)*</u>	<u>Reference</u>
Marbled salamander (embryo, larva), <u>Ambystoma opacum</u>	Mercuric chloride	7-8 days	EC50 (death and deformity)	108 107.5	Birge, et al. 1978, 1979
<u>Methylmercury</u>					
Alga, <u>Ankistrodesmus braunii</u>	Methylmercuric chloride	168-240 hrs	Lipid biosynthesis, >EC50	1,598	Matson, et al. 1972
Alga, <u>Coelastrum microporum</u>	Methylmercuric chloride	-	EC50 (growth inhibition)	>2.4-<4.8	Holderness, et al. 1975
Alga, <u>Scenedesmus obliquus</u>	Methylmercuric chloride	14 days	BCF=2,100 (Maxi- mum by third day)***	-	Havlik, et al. 1979
Alga, <u>Microcystis incerta</u>	Methylmercuric chloride	14 days	BCF=990 (Maximum by third day)***	-	Havlik, et al. 1979
Planarian, <u>Dugesia dorotocephala</u>	Methylmercuric chloride	4 days	LC50	200-500	Best, et al. 1981
Mussel, <u>Margaritifera margaritifera</u>	Methylmercuric chloride	57 days	BCF=2,463	-	Møllinger, 1973
Amphipod, <u>Gammarus</u> sp.	Methylmercuric chloride	7 days	BCF=8,000 (approx.)	-	Zubarik & O'Connor, 1978
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	Methylmercuric chloride	84 days****	BCF=4,530 (whole fish, 5 C)	-	Reinert, et al. 1974
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	Methylmercuric chloride	84 days****	BCF=6,620 (whole fish, 10 C)	-	Reinert, et al. 1974
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	Methylmercuric chloride	84 days****	BCF=8,049 (whole fish, 15 C)	-	Reinert, et al. 1974
Rainbow trout, <u>Salmo gairdneri</u>	Methylmercuric chloride	-	Inhibited growth	0.0037-0.037	Matida, et al. 1971
Rainbow trout, <u>Salmo gairdneri</u>	Methylmercuric chloride	14 days	approximate LC80	8	Blanc, 1973

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>Rainbow trout, Salmo gairdneri</u>	Methylmercuric chloride	1 wk	Effected osmo-regulation	5	Lock, et al. 1981
<u>Rainbow trout, Salmo gairdneri</u>	Methylmercuric chloride plus inorganic mercury	120 days	Loss of appetite	48 mg/kg in food	Matida, et al. 1971
<u>Rainbow trout (embryo, larva), Salmo gairdneri</u>	-	24 days	EC50 (death and deformity)	5	Birge & Black, 1977
<u>Rainbow trout, Salmo gairdneri</u>	Methylmercuric chloride plus inorganic mercury	269 days	Loss of nervous control	48 mg/kg in food	Matida, et al. 1971
<u>Rainbow trout, Salmo gairdneri</u>	Methylmercuric chloride	30 min	EC50 (Reduced viability of sperm)	1,000	McIntyre, 1973
<u>Brook trout (embryo), Salvelinus fontinalis</u>	Methylmercuric chloride	16-17 days	Decreased enzyme (GOT) activity	0.88	Christensen, 1975
<u>Brook trout (alevin), Salvelinus fontinalis</u>	Methylmercuric chloride	Incubation period + 21 days	Reduced growth	0.7	Christensen, 1975
<u>Brook trout (alevins), Salvelinus fontinalis</u>	Methylmercuric chloride	30 days	Increased enzyme (GOT) activity	0.79	Christensen, 1975
<u>Brook trout (juvenile), Salvelinus fontinalis</u>	Methylmercuric chloride	14 days	Increased blood plasma chloride	2.93	Christensen, et al. 1977
<u>Brook trout, Salvelinus fontinalis</u>	Methylmercuric chloride	8 days	Increased cough frequency	≥3	Drummond, et al. 1974
<u>Common carp, Cyprinus carpio</u>	Methylmercuric chloride	16 days	Reduced protein synthesis	0.05	Sharma & Davis, 1980b
<u>Mosquitofish, Gambusia affinis</u>	Methylmercuric chloride	<24 hrs	LC50	500	Boudou, et al. 1979

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>Mosquitofish, Gambusia affinis</u>	Methylmercuric chloride	30 days	BCF=2,500 (whole fish, 10 C)	-	Boudou, et al. 1979
<u>Mosquitofish, Gambusia affinis</u>	Methylmercuric chloride	30 days	BCF=4,300 (whole fish, 18 C)	-	Boudou, et al. 1979
<u>Mosquitofish, Gambusia affinis</u>	Methylmercuric chloride	30 days	BCF=3,000 (whole fish, 164 mg/kg in food, 10 C)	-	Boudou, et al. 1979
<u>Mosquitofish, Gambusia affinis</u>	Methylmercuric chloride	30 days	BCF=27,000 (whole fish, 238 mg/kg in food, 26 C)	-	Boudou, et al. 1979
<u>Bluegill (juvenile), Lepomis macrochirus</u>	Methylmercuric chloride	28.5 days	BCF=373***** (whole fish, 9 C)	-	Cember, et al. 1978
<u>Bluegill (juvenile), Lepomis macrochirus</u>	Methylmercuric chloride	28.5 days	BCF=921***** (whole fish, 21 C)	-	Cember, et al. 1978
<u>Bluegill (juvenile), Lepomis macrochirus</u>	Methylmercuric chloride	28.5 days	BCF=2,400***** (whole fish, 33 C)	-	Cember, et al. 1978
<u>Leopard frog (tadpole), Rana pipiens</u>	Methylmercuric chloride	48 hrs	LC100	50-100	Chang, et al. 1974
<u>Leopard frog, Rana pipiens</u>	Methylmercuric chloride	4 mos	Failure to metamorphose	1-10	Chang, et al. 1974
<u>Leopard frog (blastula embryo), Rana pipiens</u>	Methylmercuric chloride	5 days	LC50	12-16	Dial, 1976

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
Leopard frog (gastrula embryo), <u>Rana pipiens</u>	Methylmercuric chloride	5 days	LC50	8-12	Dial, 1976
Leopard frog (neural plate embryo), <u>Rana pipiens</u>	Methylmercuric chloride	5 days	LC50	12-16	Dial, 1976
Leopard frog (blastula embryo), <u>Rana pipiens</u>	Methylmercuric chloride	96 hrs	EC50 (teratogenesis)	0-4	Dial, 1976
Leopard frog (gastrula embryo), <u>Rana pipiens</u>	Methylmercuric chloride	96 hrs	EC50 (teratogenesis)	8-12	Dial, 1976
Leopard frog (neural plate embryo), <u>Rana pipiens</u>	Methylmercuric chloride	96 hrs	EC50 (teratogenesis)	12	Dial, 1976
Newt, <u>Triturus viridescens</u>	Methylmercuric chloride	>2 days	Delayed limb regeneration	8	Chang, et al. 1976
Newt, <u>Triturus viridescens</u>	Methylmercuric chloride	17 days	Death	300	Chang, et al. 1976
Newt, <u>Triturus viridescens</u>	Methylmercuric chloride	8 days	Death	1,000	Chang, et al. 1976
Mink (adult), <u>Mustela vison</u>	Methylmercuric chloride	93 days	Histologic evidence of injury	1,100	Wobeser, 1973
Mink (adult), <u>Mustela vison</u>	Methylmercuric chloride	93 days	LC50 in brain tissue	11,000	Wobeser, 1973
<u>Other Mercury Compounds</u>					
Alga, (Florida Lake assemblage)	Methylmercuric dicyandiamide	125 hrs	Reduced biomass	0.8 (approx.)	Harriss, et al. 1970

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result</u> <u>($\mu\text{g/L}$)^a</u>	<u>Reference</u>
Alga, (Florida Lake assemblage)	N-Methylmercuric- 1,2,3,6-tetrahydro- 3,6-methano-3,4,5,6, 7,7,-hexachloro- phthalimide	125 hrs	Reduced biomass	0.3 (approx.)	Harriss, et al. 1970
Alga, Cladophoraceae	Ethylmercuric phosphate	1 hr	Nuisance control	38.6	Burrows & Combs, 1958
Alga, Ulothrichaceae	Ethylmercuric phosphate	1 hr	Nuisance control	38.6	Burrows & Combs, 1958
Alga, (Florida Lake assemblage)	Phenylmercuric acetate	125 hrs	Reduced biomass	0.5 (approx.)	Harriss, et al. 1970
Alga, (Florida Lake assemblage)	Diphenyl mercury	125 hrs	Reduced biomass	2.8 (approx.)	Harriss, et al. 1970
Alga, <u>Scenedesmus obliquus</u>	Phenylmercuric chloride	14 days	BCF=13,000 (Maxi- mum by third day)	-	Havlik, et al. 1979
Alga, <u>Microcystis incerta</u>	Phenylmercuric chloride	14 days	BCF=4,000 (Maximum by third day)	-	Havlik, et al. 1979
Sponge, <u>Ephydatia fluviatilis</u>	Mercury	30 days	Malformed gemmo- scleres	1	Mysing-Gubala & Poirrier, 1981
Sponge, <u>Ephydatia fluviatilis</u>	Mercury	30 days	LC50	100-500	Mysing-Gubala & Poirrier, 1981
Amphipod, <u>Gammarus sp.</u>	Phenylmercuric acetate	7 days	BCF=8,000 (approx.)	-	Zubarik & O'Connor, 1978
Crayfish (juvenile), <u>Procambarus clarki</u>	Methylmercuric dicyandimide	120 hrs	LC50	56	Hendrick & Everett, 1965
Sockeye salmon (juvenile), <u>Oncorhynchus nerka</u>	Pyridylmercuric acetate	1.5 hrs	LC50	10,500- 15,700	Burrows & Palmer, 1949
Sockeye salmon (juvenile), <u>Oncorhynchus nerka</u>	Pyridylmercuric acetate	1 hr	Safe for disease control	<954	Rucker, 1948

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)*</u>	<u>Reference</u>
<u>Sockeye salmon (juvenile), Oncorhynchus nerka</u>	Pyridylmercuric acetate	1 hr	Safe for disease control	<4,752	Rucker & Whipple, 1951
<u>Chinook salmon (fingerling), Oncorhynchus tshawytscha</u>	Ethylmercuric phosphate	1 hr	Distress	77	Burrows & Combs, 1958
<u>Chinook salmon, Oncorhynchus tshawytscha</u>	Ethylmercuric phosphate	20 hrs	Safe for disease control	39	Burrows & Combs, 1958
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Pyridylmercuric acetate	1 hr	LC100	1,030	Allison, 1957
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Pyridylmercuric acetate	1 hr	LC0	967	Allison, 1957
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Pyridylmercuric acetate	1 hr	LC33 (8.3 C) (13.3 C)	4,750 4,750	Rodgers, et al. 1951
<u>Rainbow trout (alevin), Salmo gairdneri</u>	Pyridylmercuric acetate	1 hr	Safe for disease control	<4,750	Rucker & Whipple, 1951
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Pyridylmercuric acetate	1 hr	LC60	517	Allison, 1957
<u>Rainbow trout, Salmo gairdneri</u>	Phenylmercuric acetate	12 wks	Growth inhibition	0.11-1.1	Matida, et al. 1971
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Ethylmercuric phosphate	48 hrs	LC50	43	Matida, et al. 1971
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Ethylmercuric p-toluene sulfonamide	-	Retarded learning	5 $\mu\text{g/g}$ in feed daily or 10 $\mu\text{g/g}$ feed every fifth day	Hartman, 1978
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Pyridylmercuric acetate	24 hrs	LC50	25	MacLeod & Pessah, 1973
<u>Rainbow trout (juvenile), Salmo gairdneri</u>	Phenylmercuric acetate	48 hrs	LC50	1,780	Willford, 1966

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)[#]</u>	<u>Reference</u>
Rainbow trout (juvenile), <u>Salmo gairdneri</u>	Merthiolate	48 hrs	LC50	10,500	Willford, 1966
Brown trout (juvenile), <u>Salmo trutta</u>	Pyridylmercuric acetate	1 hr	Safe for disease control	4,750	Rodgers, et al. 1951
Brown trout (juvenile), <u>Salmo trutta</u>	Pyridylmercuric acetate	48 hrs	LC50	2,950	Willford, 1966
Brown trout (juvenile), <u>Salmo trutta</u>	Merthiolate	48 hrs	LC50	26,800	Willford, 1966
Brook trout (juvenile), <u>Salvelinus fontinalis</u>	Pyridylmercuric acetate	1 hr	Safe for disease control	2,070	Allison, 1957
Brook trout (juvenile), <u>Salvelinus fontinalis</u>	Pyridylmercuric acetate	1 hr	Safe for disease control	4,750	Rodgers, et al. 1951
Brook trout (juvenile), <u>Salvelinus fontinalis</u>	Pyridylmercuric acetate	48 hrs	LC50	5,080	Willford, 1966
Brook trout (juvenile), <u>Salvelinus fontinalis</u>	Merthiolate	48 hrs	LC50	36,900	Willford, 1966
Lake trout (juvenile), <u>Salvelinus namaycush</u>	Pyridylmercuric acetate	48 hrs	LC50	3,610	Willford, 1966
Lake trout (juvenile), <u>Salvelinus namaycush</u>	Merthiolate	48 hrs	LC50	1,060	Willford, 1966
Channel catfish (juvenile), <u>Ictalurus punctatus</u>	Pyridylmercuric acetate	72 hrs	LC50	232	Clemens & Sneed, 1958a, 1959
Channel catfish (juvenile), <u>Ictalurus punctatus</u>	Pyridylmercuric acetate	48 hrs	LC50 (10 C) (16.5 C) (24 C)	1,960 1,340 234	Clemens & Sneed, 1958b
Channel catfish (yolk sac fry), <u>Ictalurus punctatus</u>	Pyridylmercuric acetate	48 hrs	LC50 (23 C)	178	Clemens & Sneed, 1958b
Channel catfish (1 wk-old), <u>Ictalurus punctatus</u>	Pyridylmercuric acetate	48 hrs	LC50 (23 C)	<148	Clemens & Sneed, 1958b

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)*</u>	<u>Reference</u>
<u>Channel catfish,</u> <u>Ictalurus punctatus</u>	Pyridylmercuric acetate	48 hrs	LC50	1,370	Willford, 1966
<u>Channel catfish,</u> <u>Ictalurus punctatus</u>	Merthiolate	48 hrs	LC50	2,800	Willford, 1966
<u>Bluegill (juvenile),</u> <u>Lepomis macrochirus</u>	Pyridylmercuric acetate	48 hrs	LC50	7,600	Willford, 1966
<u>Bluegill (juvenile),</u> <u>Lepomis macrochirus</u>	Merthiolate	48 hrs	LC50	32,000	Willford, 1966
<u>Largemouth bass,</u> <u>Micropterus salmoides</u>	Mercury	21 days	Threshold of effect opercu- lar rhythm	10	Morgan, 1979
<u>SALTWATER SPECIES</u>					
<u>Mercury(II)</u>					
<u>Red alga,</u> <u>Antithamnion plumula</u>	Mercuric chloride	30 min	LC50 after 7 days	5,000	Boney & Corner, 1959
<u>Alga,</u> <u>Chaetoceros galvestonensis</u>	Mercuric chloride	4 days	About 30% reduction in growth	10	Hannan, et al. 1973b
<u>Alga,</u> <u>Chaetoceros galvestonensis</u>	Mercuric chloride	4 days	No growth of culture	100	Hannan, et al. 1973b
<u>Alga,</u> <u>Chaetoceros galvestonensis</u>	Mercuric chloride	4 days	BCF=10,920	-	Hannan, et al. 1973b
<u>Alga,</u> <u>Chroomonas salina</u>	Mercuric chloride	2 days	BCF=853	-	Parrish & Carr, 1976
<u>Alga,</u> <u>Cyclotella sp.</u>	Mercuric chloride	3 days	No growth of culture	100	Hannan & Patouillet, 1972
<u>Alga,</u> <u>Dunaliella sp.</u>	Mercuric chloride	-	75% reduction in CO_2	2,500	Mills & Colwell, 1977

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)^a</u>	<u>Reference</u>
Alga, <u>Dunaliella tertiolecta</u>	Mercuric chloride	8 days	About 10% increase in maximum chloro- phyll <u>a</u> concentra- tion	100	Betz, 1977
Alga, <u>Dunaliella tertiolecta</u>	Mercuric chloride	8 days	About 45% increase in maximum chloro- phyll <u>a</u> concentra- tion	220	Betz, 1977
Diatom, <u>Nitzschia acicularis</u>	Mercuric chloride	7 days	Prevented growth	150-200	Mora & Fabregas, 1980
Diatom, <u>Skeletonema costatum</u>	Mercuric chloride	15 days	Reduced cell density	0.08	Cloutier-Mantha & Harrison, 1980
Alga, <u>Dunaliella tertiolecta</u>	Mercuric chloride	3 days	About 15% reduction in growth	10	Davies, 1976
Alga, <u>Dunaliella tertiolecta</u>	Mercuric chloride	8 days	No effect on growth	2	Davies, 1976
Alga, <u>Isochrysis galbana</u>	Mercuric chloride	15 days	About 10% reduction in growth	5.1	Davies, 1974
Alga, <u>Isochrysis galbana</u>	Mercuric chloride	15 days	About 60% reduction in growth	10.5	Davies, 1974
Alga, <u>Isochrysis galbana</u>	Mercuric chloride	28 days	Growth rate recovery to near normal after day 5	10.5	Davies, 1974
Kelp (zoospores, gametophytes, sporophytes), <u>Laminaria hyperborea</u>	Mercuric chloride	28 days	Lowest concentration causing growth inhibition	10	Hopkins & Kain, 1971
Kelp (zoospores, gametophytes, sporophytes), <u>Laminaria hyperborea</u>	Mercuric chloride	22 hrs	EC50 respiration	about 450	Hopkins & Kain, 1971
Kelp (zoospores, gametophytes, sporophytes), <u>Laminaria hyperborea</u>	Mercuric chloride	28 hrs	About 80% reduc- tion in respiration	10,000	Hopkins & Kain, 1971

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)*</u>	<u>Reference</u>
Alga, <u>Phaeodactylum tricornutum</u>	Mercuric chloride	4 days	About 50% reduction in growth	50	Hannan, et al. 1973b
Alga, <u>Phaeodactylum tricornutum</u>	Mercuric chloride	4 days	No growth of culture	120	Hannan, et al. 1973a
Alga, <u>Phaeodactylum tricornutum</u>	Mercuric chloride	4 days	BCF=7,120	-	Hannan, et al. 1973b
Red alga (sporling), <u>Plumaria elegans</u>	Mercuric chloride	24 hrs	40% reduction in growth over 21 days	120	Boney, 1971
Red alga (sporling), <u>Plumaria elegans</u>	Mercuric chloride	1 hr	40% reduction in growth over 21 days	1,000	Boney, 1971
Red alga (sporling), <u>Plumaria elegans</u>	Mercuric chloride	18 hrs	LC50 after 7 days	3,170	Boney, et al. 1959
Red alga, <u>Plumaria elegans</u>	Mercuric chloride	30 min	LC50 after 7 days	6,700	Boney & Corner, 1959
Red alga, <u>Polysiphonia lanosa</u>	Mercuric chloride	30 min	LC50 after 7 days	8,000	Boney & Corner, 1959
Alga (mixed), <u>Asterionella japonica</u> plus <u>Dicogenes</u> sp.	Mercuric chloride	8 days	BCF=3,467	-	Laumond, et al. 1973
5 seaweed species, <u>Ascophyllum nodosum</u> , <u>Fucus spiralis</u> , <u>F. vesiculosus</u> , <u>F. serratus</u> , <u>Pelvetia canaliculata</u>	Mercuric chloride	10 days	10-30% reduction in growth	10	Strangren, 1980
Algae, (eighteen species)	Mercuric chloride	17 days	Growth inhibition	<5-15	Berland, et al. 1976

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/l}$)*</u>	<u>Reference</u>
Algae, (eighteen species)	Mercuric chloride	17 days	Death	10-50	Berland, et al. 1976
Algae, (three species)	Mercuric chloride	168 hrs	Depressed growth	30-350	Sick & Windom, 1975
Algae, (three species)	Mercuric chloride	168 hrs	No further bioaccumulation	40	Sick & Windom, 1975
Algae, (three species)	Mercuric chloride	168 hrs	Changes in cell chemistry	30-350	Sick & Windom, 1975
Natural phytoplankton populations	Mercuric chloride	120 hrs	Reduced chlorophyll a	6	Hollibaugh, et al. 1980
Natural phytoplankton populations	Mercuric chloride	96 hrs	Reduced biomass	2	Hollibaugh, et al. 1980
Phytoplankton, (Natural assemblages)	Mercuric chloride	21 days	Inhibited growth	1	Thomas, et al. 1977
Protozoan, <u>Cristigera</u> sp.	Mercuric chloride	12 hrs	Reduced growth	2.5-5	Gray & Ventilla, 1973
Protozoan, <u>Euplotes vannus</u>	Mercuric chloride	48 hrs	Inhibition of reproduction	1,000	Persone & Uyttersprot, 1975
Sand worm (adult), <u>Nereis virens</u>	Mercuric chloride	168 hrs	LC50	60	Eisler & Hennekey, 1977
Sand worm (adult), <u>Nereis virens</u>	Mercuric chloride	168 hrs	LC100	125	Eisler & Hennekey, 1977
Polychaete worm (adult), <u>Ophryotrocha diadema</u>	Mercuric chloride	96 hrs	LC13	50	Reish & Carr, 1978
Polychaete worm (adult), <u>Ophryotrocha diadema</u>	Mercuric chloride	96 hrs	LC60	100	Reish & Carr, 1978
Polychaete worm (adult), <u>Ophryotrocha diadema</u>	Mercuric chloride	96 hrs	LC100	500	Reish & Carr, 1978
Polychaete worm, <u>Ophryotrocha diadema</u>	Mercuric chloride	48 hrs	LC50	30-100	Parker, 1984

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)^a</u>	<u>Reference</u>
Blue mussel (larva), <u>Mytilus edulis</u>	Mercuric chloride	24 hrs	Abnormal development	32	Okubo & Okubo, 1962
Pacific oyster (larva), <u>Crassostrea gigas</u>	Mercuric chloride	24 hrs	Abnormal development	32	Okubo & Okubo, 1962
Eastern oyster (embryo), <u>Crassostrea virginica</u>	Mercuric chloride	12 days	LC50	12	Calabrese & Nelson, 1974; Calabrese, et al. 1977
Eastern oyster (embryo), <u>Crassostrea virginica</u>	Mercuric chloride	48 hrs	LC0	1	Calabrese, et al. 1973
Eastern oyster (embryo), <u>Crassostrea virginica</u>	Mercuric chloride	19 days	Trace metal upset	50	Kopfler, 1974
Clam, <u>Mulinna lateralis</u>	Mercuric chloride	72 hrs	Reduced calcium uptake	26.5	Ming-Shan & Zubkoff, 1982
Common rangia, <u>Rangia cuneata</u>	Mercuric chloride	96 hrs	LC50 (<1 g/kg salinity)	5,100	Olson & Harrel, 1973
Common rangia, <u>Rangia cuneata</u>	Mercuric chloride	14 days	BCF=1,130 (whole animal)	-	Dillon & Neff, 1978
Quahog clam (larva), <u>Mercenaria mercenaria</u>	Mercuric chloride	8-10 days	LC50	14	Calabrese & Nelson, 1974; Calabrese, et al. 1977
Quahog clam (larva), <u>Mercenaria mercenaria</u>	Mercuric chloride	42-48 hrs	LC0	2.5	Calabrese, et al. 1973
Soft-shell clam (adult), <u>Mya arenaria</u>	Mercuric chloride	168 hrs	LC0	1	Eisler & Hennekey, 1977
Soft-shell clam (adult), <u>Mya arenaria</u>	Mercuric chloride	168 hrs	LC50	4	Eisler & Hennekey, 1977
Soft-shell clam (adult), <u>Mya arenaria</u>	Mercuric chloride	168 hrs	LC100	30	Eisler & Hennekey, 1977
Copepods (adult), (5 genera)	Mercuric chloride	10 days	90% decrease in egg production	10	Reeve, et al. 1977
Copepods (adult), (5 genera)	Mercuric chloride	10 days	70% decrease in fecal pellet	10	Reeve, et al. 1977

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)[*]</u>	<u>Reference</u>
Copepods (adult), (5 genera)	Mercuric chloride	48 hrs	Hg-Cu Interactions on LC50	17 (Hg in mixture)	Reeve, et al. 1977
Copepod (adult), <u>Pseudocalanus minutus</u>	Mercuric chloride	70 days	No growth of culture	5	Sonntag & Greve, 1977
Copepod (adult), <u>Pseudocalanus minutus</u>	Mercuric chloride	70 days	No growth inhibition	1	Sonntag & Greve, 1977
Copepod (adult), <u>Acartia clausi</u>	Mercuric chloride	1.9 hrs	LC50	50	Corner & Sparrow, 1956
Copepod (adult), <u>Acartia clausi</u>	Mercuric chloride	24 hrs	BCF=7,500	-	Relichiro, et al. 1983
Barnacle (adult), <u>Balanus balanoides</u>	Mercuric chloride	48 hrs	LC90	1,000	Clarke, 1947
Barnacle (cyprid), <u>Balanus balanoides</u>	Mercuric chloride	6 hrs	About 10% reduction in substrate attach- ment over 19 days	10	Pyefinch & Mott, 1948
Barnacle (cyprid), <u>Balanus balanoides</u>	Mercuric chloride	6 hrs	LC50	90	Pyefinch & Mott, 1948
Barnacles (nauplius), <u>Balanus crenatus</u>	Mercuric chloride	6 hrs	LC50	60	Pyefinch & Mott, 1948
Barnacle (cyprid), <u>Balanus improvisus</u>	Mercuric chloride	48 hrs	About 50% abnormal development	16,600	Clarke, 1947
White shrimp (adult), <u>Penaeus setiferus</u>	Mercuric chloride	60 days	No effect on respiration, growth, or molting	1	Green, et al. 1976
Grass shrimp (larva), <u>Palaemonetes vulgaris</u>	Mercuric chloride	<24 hrs	LC100	56	Shealy & Sandifer, 1975
Grass shrimp (larva), <u>Palaemonetes vulgaris</u>	Mercuric chloride	48 hrs	LC0	<5.6	Shealy & Sandifer, 1975
Grass shrimp, <u>Palaemonetes pugio</u>	Mercuric chloride	120 hrs	LC50	148	Barthalmus, 1977

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)*</u>	<u>Reference</u>
<u>Grass shrimp,</u> <u>Palaemonetes pugio</u>	Mercuric chloride	24 hrs	Impaired condi- tioned avoidance response	37	Barthalmus, 1977
<u>Grass shrimp (larva),</u> <u>Palaemonetes vulgaris</u>	Mercuric chloride	48 hrs	LC50	10	Shealy & Sandifer, 1975
<u>Grass shrimp (larva),</u> <u>Palaemonetes vulgaris</u>	Mercuric chloride	48 hrs	Abnormal development	10-18	Shealy & Sandifer, 1975
<u>Hermit crab (adult),</u> <u>Pagurus longicarpus</u>	Mercuric chloride	168 hrs	LC0	10	Eisler & Hennekey, 1977
<u>Hermit crab (adult),</u> <u>Pagurus longicarpus</u>	Mercuric chloride	168 hrs	LC50	50	Eisler & Hennekey, 1977
<u>Hermit crab (adult),</u> <u>Pagurus longicarpus</u>	Mercuric chloride	168 hrs	LC100	125	Eisler & Hennekey, 1977
<u>Green crab (adult),</u> <u>Carcinus maenas</u>	Mercuric chloride	48 hrs	LC50	1,000	Portmann, 1968
<u>Green crab (adult),</u> <u>Carcinus maenas</u>	Mercuric chloride	48 hrs	LC50	1,200	Connor, 1972
<u>Green crab (larva),</u> <u>Carcinus maenas</u>	Mercuric chloride	47 hrs	LC50	10	Connor, 1972
<u>Green crab (larva),</u> <u>Carcinus maenas</u>	Mercuric chloride	20-30 hrs	LC50	33	Connor, 1972
<u>Green crab (larva),</u> <u>Carcinus maenas</u>	Mercuric chloride	4.3-13.5 hrs	LC50	100	Connor, 1972
<u>Green crab (larva),</u> <u>Carcinus maenas</u>	Mercuric chloride	2.7 hrs	LC50	1,000	Connor, 1972
<u>Green crab (larva),</u> <u>Carcinus maenas</u>	Mercuric chloride	0.5 hrs	LC50	3,300	Connor, 1972
<u>Green crab (larva),</u> <u>Carcinus maenas</u>	Mercuric chloride	0.22 hrs	LC50	10,000	Connor, 1972

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)[#]</u>	<u>Reference</u>
<u>Fiddler crab (adult), Uca pugilator</u>	Mercuric chloride	28 days	Low survival, inhibited limb regeneration	1,000	Wels, 1976
<u>Fiddler crab (adult), Uca pugilator</u>	Mercuric chloride	6 days	20-25% reduction in percent survival	180	Vernberg & Vernberg, 1972
<u>Fiddler crab (adult), Uca pugilator</u>	Mercuric chloride	6 days	20-25% reduction in percent survival	180	Vernberg & Vernberg, 1972
<u>Fiddler crab (adult), Uca pugilator</u>	Mercuric chloride	24 hrs	Increased oxygen consumption	180	Vernberg & Vernberg, 1972
<u>Fiddler crab (zoea), Uca pugilator</u>	Mercuric chloride	8 days	LC50	1.8	DeCoursey & Vernberg, 1972
<u>Fiddler crab (zoea), Uca pugilator</u>	Mercuric chloride	24 hrs	20-100% increase in metabolic rate after stage I zoea	1.8	DeCoursey & Vernberg, 1972
<u>Fiddler crab (zoea), Uca pugilator</u>	Mercuric chloride	5 days	About 40% increase in swimming activity of stage V zoea	1.8	DeCoursey & Vernberg, 1972
<u>Starfish (adult), Asterias forbesi</u>	Mercuric chloride	168 hrs	LC0	10	Eisler & Hennekey, 1977
<u>Starfish (adult), Asterias forbesi</u>	Mercuric chloride	168 hrs	LC50	20	Eisler & Hennekey, 1977
<u>Starfish (adult), Asterias forbesi</u>	Mercuric chloride	168 hrs	LC100	125	Eisler & Hennekey, 1977
<u>Sea urchin (spermatzoa), Arbacia punctulata</u>	Mercuric chloride	8 min	About 150% increase in swimming speed	20	Young & Nelson, 1974
<u>Sea urchin (spermatzoa), Arbacia punctulata</u>	Mercuric chloride	24 min	About 80% decrease in swimming speed	2,000	Young & Nelson, 1974
<u>Sea urchin (embryo), Arbacia punctulata</u>	Mercuric chloride	13 hrs	Abnormal development	92	Waterman, 1937
<u>Haddock (embryo), Melanogrammus aeglefinus</u>	Mercuric chloride	96 hrs	LC50	918	Cardin, 1982

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	168 hrs	LC0	100	Eisler & Hennekey, 1977
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	168 hrs	LC50	800	Eisler & Hennekey, 1977
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	168 hrs	LC100	1,000	Eisler & Hennekey, 1977
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	24 hrs	Disrupted osmoregulation	125	Renfro, et al. 1974
<u>Mummichog (adult), Fundulus heteroclitus</u>	-	96 hrs	Affected liver enzymes	200	Jackim, et al. 1970
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	28 days	Up to 40% reduction in enzyme activity before recovery	12	Jackim, 1973
<u>Mummichog (embryo), Fundulus heteroclitus</u>	Mercuric chloride	3 days	Many developmental abnormalities	30-40	Wels & Wels, 1977
<u>Mummichog (embryo), Fundulus heteroclitus</u>	Mercuric chloride	3 days	Some developmental abnormalities	10-20	Wels & Wels, 1977
<u>Mummichog (embryo), Fundulus heteroclitus</u>	Mercuric chloride	12 hrs	Some developmental abnormalities	30-40	Wels & Wels, 1977
<u>Mummichog (embryo), Fundulus heteroclitus</u>	Mercuric chloride	32 days	EC50	67.4	Sharp & Neff, 1980
<u>Mummichog (larva), Fundulus heteroclitus</u>	Mercuric chloride	-	No effect	50	Wels & Wels, 1983
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	-	Mercury redistribution organs following Se pretreatment	1,000 µg Hg/kg body wt plus 400 µg Se/kg body wt	Shelline & Schmidt-Nielsen, 1977
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	96 hrs	Cellular degeneration	250-5,000	Gardner, 1975
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	48 hrs	LC100	2,000	Eisler, et al. 1972

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
<u>Mummichog (adult), Fundulus heteroclitus</u>	Mercuric chloride	96 hrs	Sluggish, uncoordinated swimming	1,150	Klaunig, et al. 1975
<u>Shiner perch, Cymatogaster aggregata</u>	Mercuric chloride	-	45% reduction of brain cholinesterase activity	33,900	Abou-Donia & Menzel, 1967
<u>Striped bass (adult), Morone saxatilis</u>	Mercuric chloride	30 days	Decreased respiration 30 days post exposure	5	Dawson, et al. 1977
<u>Winter flounder (adult), Pseudopleuronectes americanus</u>	Mercuric chloride	60 days	Decreased respiration	10	Calabrese, et al. 1975
<u>Methylmercury</u>					
<u>Alga, Dunaliella tertiolecta</u>	Methylmercuric chloride	10 min	EC50 (photosynthesis)	about 170	Overnell, 1975
<u>Alga, Phaeodactylum tricornutum</u>	Methylmercuric chloride	25 days	EC50 (photosynthesis)	about 190	Overnell, 1975
<u>Red alga (sporling), Plumaria elegans</u>	Methylmercuric chloride	18 hrs	LC50 after 7 days	44	Boney, et al. 1959
<u>Alga, Tetraselmis succica</u>	Methylmercuric chloride	3 days	Inhibited growth	25	Mora & Fabregas, 1980
<u>Alga, Chaetoceros sp.</u>	Dimethylmercury	3 days	About 75% reduction in growth	100	Hannan & Patouillet, 1972
<u>Alga, Cyclotella sp.</u>	Dimethylmercury	3 days	About 15% reduction in growth	500	Hannan & Patouillet, 1972
<u>Alga, Phaeodactylum sp.</u>	Dimethylmercury	3 days	About 45% reduction in growth	500	Hannan & Patouillet, 1972
<u>Red alga (sporling), Plumaria elegans</u>	Methylmercuric chloride	25 min	EC50 (growth over 21 days)	40	Boney, 1971
<u>Diatom, Nitzschia acicularis</u>	Methylmercuric chloride	3 days	Inhibited growth	25	Mora & Fabregas, 1980

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)[#]</u>	<u>Reference</u>
Dinoflagellate, <u>Scippsilella faeroense</u>	Mercuric acetate	14 days	No growth of culture	1,000	Kayser, 1976
Alga, <u>Chlorella</u> sp.	Ethylmercuric phosphate	10 days	22% reduction in growth	0.6	Ukeles, 1962
Alga, <u>Chlorella</u> sp.	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Alga, <u>Dunaliella euchlora</u>	Ethylmercuric phosphate	10 days	36% reduction in growth	0.6	Ukeles, 1962
Alga, <u>Dunaliella euchlora</u>	Ethylmercuric phosphate	10 days	100% lethal to culture	60	Ukeles, 1962
Alga, <u>Monochrysis lutheri</u>	Ethylmercuric phosphate	10 days	No reduction in growth	0.6	Ukeles, 1962
Alga, <u>Monochrysis lutheri</u>	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Alga, <u>Phaeodactylum tricornutum</u>	Ethylmercuric phosphate	10 days	45% reduction in growth	0.6	Ukeles, 1962
Alga, <u>Phaeodactylum tricornutum</u>	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Alga, <u>Protococcus</u> sp.	Ethylmercuric phosphate	10 days	14% reduction in growth	0.6	Ukeles, 1962
Alga, <u>Protococcus</u> sp.	Ethylmercuric phosphate	10 days	100% lethal to culture	6	Ukeles, 1962
Red alga (sporling), <u>Plumaria elegans</u>	Mercuric iodide	18 hrs	LC50 after 7 days	156	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	Ethylmercuric chloride	18 hrs	LC50 after 7 days	26	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	Phenylmercuric chloride	18 hrs	LC50 after 7 days	54	Boney, et al. 1959

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result ($\mu\text{g/L}$)[#]</u>	<u>Reference</u>
Diatom, <u>Nitzschia delicatissima</u>	Methylmercuric dicyandiamide	24 hrs	EC50 (photosynthesis)	0.4	Harriss, et al. 1970
Blue mussel (adult), <u>Mytilus edulis</u>	Methylmercuric chloride	24 hrs	About 90% reduced feeding rate	400	Dorn, 1976
Eastern oyster (adult), <u>Crassostrea virginica</u>	Methylmercuric chloride	19 days	Trace metal upset	50	Kopfler, 1974
Copepod (adult), <u>Acartia clausi</u>	Methylmercuric chloride	24 hrs	BCF=56,000-350,000	-	Reilchiro, et al. 1983
Amphipod (adult), <u>Gammarus duebeni</u>	Methylmercuric chloride	3 days	Induced diuresis	56	Lockwood & Inman, 1975
Fiddler crab (adult), <u>Uca</u> sp.	Methylmercuric chloride	32 days	No limb regeneration	300-500	Weis, 1977
Fiddler crab (adult), <u>Uca</u> sp.	Methylmercuric chloride	32 days	Melanin absent in regerated limbs	100	Weis, 1977
Mummichog (adult), <u>Fundulus heteroclitus</u>	Methylmercuric chloride	24 hrs	Disrupted osmoregulation	125	Renfro, et al. 1974
Mummichog (embryo), <u>Fundulus heteroclitus</u>	Methylmercuric chloride	7 days	Teratological effects	50	Weis, et al. 1981
Mummichog (larva), <u>Fundulus heteroclitus</u>	Methylmercuric chloride	-	Reduced LT50s	50	Weis & Weis, 1983
Mummichog (embryo), <u>Fundulus heteroclitus</u>	Methylmercuric chloride	-	Developed resistance in a pond	50	Weis & Weis, 1984
Striped mullet, <u>Mugil cephalus</u>	Methylmercuric chloride	13 days	Inhibited fin regeneration	1	Weis & Weis, 1978
<u>Other Mercury Compounds</u>					
Dinoflagellate, <u>Gymnodinium spöndens</u>	Mercuric acetate	11 days	55% reduction in growth	10	Kayser, 1976
Dinoflagellate, <u>Scrippsiella faeroense</u>	Mercuric acetate	25 days	45% reduction in growth, morphological variation	10	Kayser, 1976

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
Red alga (sporling), <u>Plumaria elegans</u>	Phenylmercuric iodide	18 hrs	LC50 after 7 days	104	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	Isoamylmercuric chloride	18 hrs	LC50 after 7 days	19	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	n-amylmercuric chloride	18 hrs	LC50 after 7 days	13	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	Isopropylmercuric chloride	18 hrs	LC50 after 7 days	28	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	n-propylmercuric chloride	18 hrs	LC50 after 7 days	13	Boney, et al. 1959
Red alga (sporling), <u>Plumaria elegans</u>	n-butylmercuric chloride	18 hrs	LC50 after 7 days	13	Boney, et al. 1959
Diatom, <u>Nitzschia delicatissima</u>	N-methylmercuric- 1,2,3,6-tetrahydro- 3,6-methano-3,4,5,6, 7,7-hexachloro- phthalimine	24 hrs	EC50 (photosynthesis)	0.3	Harriss, et al. 1970
Diatom, <u>Nitzschia delicatissima</u>	Phenylmercuric acetate	24 hrs	EC50 (photosynthesis)	1.5	Harriss, et al. 1970
Diatom, <u>Nitzschia delicatissima</u>	Diphenylmercury	24 hrs	EC50 (photosynthesis)	18	Harriss, et al. 1970
Diatom, <u>Nitzschia acicularis</u>	Phenylmercuric acetate	7 days	Inhibited growth	25	Mora & Fabregas, 1980
Eastern oyster (adult), <u>Crassostrea virginica</u>	Mercuric acetate	12 hrs daily for 15 days	33% reduction in shell growth	10	Cunningham, 1976
Eastern oyster (adult), <u>Crassostrea virginica</u>	Mercuric acetate	60 days	LC55	100	Cunningham, 1976
Eastern oyster (adult), <u>Crassostrea virginica</u>	Phenylmercuric chloride	19 days	Trace metal upset	50	Kopfler, 1974

Table 6. (Continued)

<u>Species</u>	<u>Chemical</u>	<u>Duration</u>	<u>Effect</u>	<u>Result (µg/L)*</u>	<u>Reference</u>
Copepod (adult), <u>Acartia clausi</u>	Mercuric acetate	1.9 hrs	LC50	50	Corner & Sparrow, 1956
Copepod (adult), <u>Acartia clausi</u>	Ethylmercuric chloride	1.9 hrs	LC50	50	Corner & Sparrow, 1956
Coho salmon (adult), <u>Oncorhynchus kisutch</u>	Pyridylmercuric acetate	12-15 wks, 1 hr wkly as juven- iles	0.03 mg Hg/kg wet wt muscle 2 yrs post-exposure	1,000	Amend, 1970
Sockeye salmon (juvenile), <u>Oncorhynchus nerka</u>	Pyridylmercuric acetate	12-15 wks, 1 hr wkly	1.2 mg Hg/kg wet wt muscle 12 weeks post-exposure	1,000	Amend, 1970
Sockeye salmon (adult), <u>Oncorhynchus nerka</u>	Pyridylmercuric acetate	12-15 wks, 1 hr wkly as juven- iles	0.24 mg Hg/kg wet wt muscle 3 yrs post-exposure	1,000	Amend, 1970
Sockeye salmon (adult), <u>Oncorhynchus nerka</u>	Pyridylmercuric acetate	12 1-hr exposures as juven- iles	0.04 mg Hg/kg wet wt muscle 4 yrs post-exposure	1,000	Amend, 1970
Chinook salmon (adult), <u>Oncorhynchus tshawytscha</u>	Pyridylmercuric acetate	35 wks, 1 hr wkly as juveniles	up to 0.12 mg Hg/kg muscle 4 yrs later	1,000	Amend, 1970
Threespine stickleback, <u>Gasterosteus aculeatus</u>	Phenylmercuric acetate	370 min	LC100	100	Boetius, 1960

* Results are expressed as mercury, not as the chemical.

** In river water.

*** Static, continual loss over time.

**** Not at steady-state.

*****BCF Independent of concentration in water over range tested.

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