

2022 National Lakes Assessment Human Health Fish Tissue Study

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Background

Obtaining statistically representative data on contaminants in fish is a priority area of interest for EPA. In the early 2000s, EPA's Office of Science and Technology (OST), within the Office of Water (OW), conducted the first national probability-based study of chemical contamination in fish from U.S. lakes, ponds, and reservoirs (hereafter referred to as lakes), known as the National Lake Fish Tissue Study. Mercury and PCBs were detected in all fish samples collected from 500 sampling locations in the lower 48 states, and it was estimated that 49% of U.S. lakes, at the time, contained fish with mercury levels that exceeded the human health screening level of 300 parts per billion (i.e., EPA's fish tissue-based water quality criterion for methylmercury).

Twenty years later, the 2022 National Lakes Assessment (NLA), one of EPA's National Aquatic Resource Surveys (NARS), has provided an opportunity to conduct a contemporary national-scale assessment of chemical contaminants in fish from U.S. lakes. The NLA is a probabilitybased national survey that includes collection and analysis of physical, chemical, and biological indicator data to allow a statistically valid characterization of the condition of the Nation's lakes. OST is collaborating with the Office of Wetlands, Oceans, and Watersheds and with the Office of Research and Development to complete the 2022 National Lakes Assessment Human Health Fish Tissue Study, which

will compare current chemical concentrations of certain chemicals in lake fish to results from the National Lake Fish Tissue Study, and will evaluate lake fish fillet contaminant changes over time.



Study Design

The NLA Human Health Fish Tissue Study involves:

- Attempting to collect whole fish samples from 636 randomly selected lakes in the coterminous U.S.
- Obtaining one fish composite sample from each lake (optimally, five similarly sized adult fish of the same species that are commonly consumed by humans)
- Shipping whole fish samples to sample repository freezers in Baltimore, Maryland for interim storage
- Transferring the frozen whole fish samples to the sample preparation laboratory in Owings Mills, MD
- Preparing fillet tissue samples by scaling and filleting each fish in the composite sample, homogenizing the fillets from all the fish in the sample, and dividing homogenates into aliquots for chemical analyses
- Analyzing the fillet samples for mercury, 40 per- and polyfluoroalkyl substances (PFAS), and 209 polychlorinated biphenyl (PCB) congeners

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413 Sampling Locations

National Lakes Assessment Human Health Fish Tissue Study

- Second statistically based assessment of contaminants in fish from inland lakes of the conterminous United States, mirroring EPA's National Lake Fish Tissue Study of 2000–2003 to assess change over time
 - Fish successfully collected from 413 of the 636 randomly selected sampling locations.

Sample Collection

- The target population consisted of all lakes and reservoirs in the lower 48 states with a surface area >1 hectare and that contain 1,000 square meters of open, unvegetated space and a permanent population of predator fish species.
- A total of 636 randomly selected lakes were targeted for fish sampling.
- Sampling procedures for collection of whole fish samples are detailed in EPA's National Lakes Assessment 2022 Quality Assurance Project Plan and National Lakes Assessment 2022 Field Operations Manual.
- Procedures included a list of 12 primary target predator fish species and 10 secondary predator fish species (all commonly consumed by humans).
- Most fish samples were collected by hook and line or electrofishing.
- A total 1,512 fish were collected, which yielded 413 valid whole fish composite samples from the 636 sites targeted for fish sampling.
- A total of 21 species were collected, and the majority of samples (66%) were Largemouth Bass.

Sample Preparation

Whole fish samples were stored in freezers

- at \leq -20° Celsius (C). The sample preparation laboratory is scaling and filleting each fish, homogenizing the fillet tissue, and preparing the required number of fish fillet tissue aliquots for
- Sample preparation quality control procedures include analysis of lipids in ground fillet tissue samples for homogeneity testing and analysis of mercury, PCBs, and PFAS in equipment rinsates to test the adequacy of equipment
- The filleting process involves removing the fillet (with skin on and "belly flap" or ventra muscle attached) from both sides of each fish, combining fillets from all fish in the composite sample, and homogenizing them together using an electric meat grinder.
- Sample preparation procedures are detailed in EPA's Quality Assurance Project Plan for 2022 National Lake Assessment (NLA) Fish Tissue Study Sample Preparation.

Analysis

- Fillet tissue homogenates will be prepared and analyzed for mercury using Appendix to Method 1631, Total Mercury in Tissue, Sludge, Sediment, and Soil by Acid Digestion and BrCl Oxidation Revision B and Revision E, respectively.
- Fillet homogenates will also be analyzed for 40 PFAS and 209 PCB congeners (Analytical method requirements for PFAS and PCB analyses will be described in the Quality Assurance Project Plan for Analysis of the 2022 National Lake Assessment Fish Fillet Samples for Mercury, Per- and Polyfluoroalkyl Substances, and Polychlorinated Biphenyls.
- Statistical analysis will include an estimation of the number and proportion of lakes in the sampled population, and an estimation of the cumulative distribution and percentile concentrations of the target chemicals in
- The percentage of the sampled population of U.S. lakes containing fish with fillet concentrations that exceed screening levels for human health protection will be estimated by overlaying these screening levels on cumulative distribution plots for the target chemicals.

Target Chemicals Metals **Target Analyte CAS Number** Mercury (total) 7439-97-6 Per- and Polyfluoroalkyl Substances (PFAS) **Target Analyte Abbreviation CAS Number** Perfluoroalkyl carboxylic acids Perfluorobutanoic acid 375-22-4 2706-90-3 Perfluoropentanoic acid 307-24-4 Perfluorohexanoic acid Perfluoroheptanoic acid 375-85-9 335-67-1 Perfluorooctanoic acid 375-95-1 Perfluorononanoic acid Perfluorodecanoic acid 335-76-2 2058-94-8 Perfluoroundecanoic acid PFDoA Perfluorododecanoic acid 307-55-1 72629-94-8 Perfluorotridecanoic acid PFTrDA Perfluorotetradecanoic acid PFTeDA 376-06-7 Perfluroalkyl sulfonic acids 375-73-5 Perfluorobutanesulfonic acid 2706-91-4 Perfluoropentansulfonic acid PFHxS Perfluorohexanesulfonic acid 355-46-4 Perfluoroheptanesulfonic acid 375-92-8 Perfluorooctanesulfonic acid 1763-23-1 Perfluorononanesulfonic acid 68259-12-1 Perfluorodecanesulfonic acid 335-77-3 PFDoS Perfluorododecanesulfonic acid 79780-39-5 Fluorotelomer sulfonic acids 4:2FTS 757124-72-4 1H,1H, 2H, 2H-Perfluorohexane sulfonic acid 6:2FTS 1H,1H, 2H, 2H-Perfluorooctane sulfonic acid 27619-97-2 8:2FTS 1H,1H, 2H, 2H-Perfluorodecane sulfonic acid 39108-34-4 **Perfluorooctane sulfonamides** PFOSA 754-91-6 Perfluorooctanesulfonamide **NMeFOSA** N-methyl perfluorooctanesulfonamide 31506-32-8 **NEtFOSA** N-ethyl perfluorooctanesulfonamide 4151-50-2 Perfluorooctane sulfonamidoacetic acids 2355-31-9 N-methyl perfluorooctanesulfonamidoacetic acid **NMeFOSAA NEtFOSAA** 2991-50-6 N-ethyl perfluorooctanesulfonamidoacetic acic **Perfluorooctane sulfonamide ethanols NMeFOSE** N-methyl perfluorooctanesulfonamidoethanol 24448-09-7 **NEtFOSE** N-ethyl perfluorooctanesulfonamidoethanol 1691-99-2 Per- and Polyfluoroether carboxylic acids HFPO-DA 13252-13-6 Hexafluoropropylene oxide dimer acid 4,8-Dioxa-3H-perfluorononanoic acid **ADONA** 919005-14-4 Perfluoro-3-methoxypropanoic acid 377-73-1 PFMBA 863090-89-5 Perfluoro-4-methoxybutanoic acid 151772-58-6 Nonafluoro-3,6-dioxaheptanoic acid NFDHA **Ether sulfonic acids** 9-Chlorohexadecafluoro-3-oxanonane-1-sulfonic acid 9CI-PF3ONS 756426-58-1 11-Chloroeicosafluoro-3-oxaundecane-1-sulfonic acid 11Cl-PF3OUdS 763051-92-9 PFEESA 113507-82-7 Perfluoro(2-ethoxyethane)sulfonic acid Fluorotelomer carboxylic acids 3:3FTCA 356-02-5 3-Perfluoropropyl propanoic acid 914637-49-3 2H,2H,3H,3H-Perfluorooctanoic acid 5:3FTCA 7:3FTCA 3-Perfluoroheptyl propanoic acid 812-70-4 **Polychlorinated Biphenyls (PCBs) Target Analyte** Full congener analysis providing results for 209 congeners

Anticipated Study Milestones

Ending Fish Preparation Completed 2022 2023

Sample Collection

Sample Collection

Beginning

2024 Fish Fillet **Analysis Completed** 2025

Reporting

Completed

Data Analysis

Completed